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No. **420** September 1993

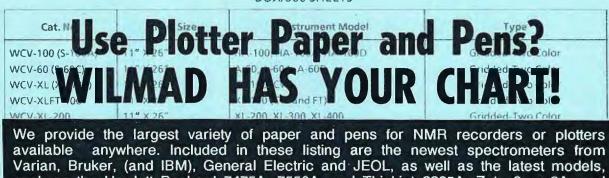
History of NMR: Request for Information and Anecdotes	2
Electronic Substituent Effects on $^2\Delta$ Isotope Shifts of 4-Substituted Benzanilides	
Morales-Ríos, M. S., Zepeda, L. G., and Joseph-Nathan, P.	5
¹³ C Chemical Shift Tensors from Carbon-13 Depleted Compounds	
Maliniak, A., Greenbaum, G., Poupko, R., Zimmermann, H., and Luz, Z.	9
25th Southeastern Magnetic Resonance Conference	10
AM-DISCO Janssen, J. W. G., and van Os, J. W. M.	13
Conformational Interchange in Dicyclohexanodiethyleneglycol by ¹³ C NMR Buchanan, G. W.	14
An Unusual Velocity Calibration Device	17
Nitrogen Chemical Shift Assignments for D-Fructose Oxime	18
Applications of Pulsed Field Gradients in High-Resolution NMR Spectroscopy	
Reid, D. G., MacLachlan, L. K., Hubbard, J. A., Doddrell, D. M., and Brereton, I. M.	21
Adiabatic Pulses on a GBS2	25
Equipment Available	26
Structural Assignment of a Regioisomer of C ₆₀ H ₄ Henderson, C. C., Cahill, P. A., and Assink, R. A.	29
Proton Exchange - Selective Pulses	33
Flat Baselines in 1D and Baseplanes in 2D Spectra	34
Continued on page	e 52

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TEXAS A&M NMR NEWS	LETTER NO. 420	, SEPTEMBER 1993	AUTHOR INDEX
Altobelli, S 17	Doddrell, D. M 21	Landucci, L 37	Ralph, J 37, 38
Assink, R. A 29	Fukushima, E 17	Landucci, W 37	Ralph, S 37
Becker, E. D 2	Greenbaum, G 9	Laukien, F. H 41	Reid, D. G 21
Bigler, B 26	Hajek, M 25	Luz, Z 9	Sadek, M 34
Brereton, I. M 21	Henderson, C. C 29	Maas, W. E 41	Schaefer, T 51
Brey, W. S 10	Herynek, V 25	MacLachlan, L. K 21	Snyder, J. R 18
Brownlee, B 34	Hubbard, J. A 21	Maliniak, A 9	Sterk, H 33
Buchanan, G. W 14	Janssen, J. W. G 13	Morales-Ríos, M. S 5	Zepeda, L.G 5
Cahill, P. A 29	Joseph-Nathan, P. 5	van Os, J. W. M 13	Zimmermann, H 9
Cory, D. G 41	Krawanja, I 33	Poupko, R 9	
TEXAS A&M NMR NEWS	SLETTER NO. 420	, SEPTEMBER 1993	ADVERTISER INDE
Acorn NMR	19	JEOL	
Acorn NMR	19	JEOL	outside back cover
Acorn NMR . American Microwave Technology . Bruker Instruments, Inc		JEOL	outside back cover
Acorn NMR		JEOL	outside back cover
Acorn NMR		JEOL	outside back cover
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FORTHCOMING NMR MEETINGS

- 1993 FACSS Meeting, Detroit, Michigan, October 17-22, 1993; Contact: H. N. Cheng, Hercules, Inc., Research Center, 500 Hercules Road, Wilmington, DE 19808; Phone: (302) 995-3505; Fax: (302) 995-4117. See TAMU NMR Newsletter 411, 10.
- 1993 Pacific Conference on Chemistry and Spectroscopy, (Joint Meeting with the American Chemical Society's Western Regional Meeting), Pasadena, California, October 19- 23, 1993; Contact: Ms. B. Belmont, Pacific Conf., 14934 S. Figueroa St., Gardena, CA 90248; Phone: (310) 538-9709.
- 1993 CABM Fall Symposium on "Macromolecular Recognition", Piscataway, NJ, October 21-22, 1993; Program Organizer: G. T. Montelione; Contact: Linda Van Derveer, Symposium Coordinator, Center for Advanced Biotechnology and Medicine, 679 Hoes Lane, Piscataway, NJ 08854; Phone: (908) 235-5309; Fax: (908) 235-4850. See Newsletter 419, 30.
- 35th ENC (Experimental NMR Conference). Asilomar Conference Center, Pacific Grove, California, April 10 15, 1994; Contact: ENC, 815 Don Gaspar, Santa Fe, NM 87501; (505) 989-4573; Fax: (505) 989-1073.
- 12th International Meeting on NMR Spectroscopy, Sponsored by the Royal Society of Chemistry, Manchester, England, July 2 7, 1995 [sic]; Contact: Dr. J. F. Gibson or Ms. G. B. Howlett See TAMU NMR Newsletter 415, 5; Phone: (44-71) 437-8656; Fax: (44-71) 437-8883.
- ISMAR 1995, Sydney, NSW, Australia, July 16-21, 1995 [sic]; Contact: Dr. Wm. A. Bubb, Secretary, Univ. of Sydney, Dept. of Biochemistry, Sydney, NSW 2006, Australia. See TAMU NMR Newsletter 419, 26.

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July 26, 1993

(received 8/2/93)

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Dr. B. L. Shapiro TAMU NMR Newsletter 966 Elsinore Court

Palo Alto, CA 94303

Dear Barry:

History of NMR: Request for Information and Anecdotes

Readers of the Newsletter may be interested to know that an Encyclopedia of NMR is being developed for publication in 1995, the 50th anniversary of the first successful observations of NMR in bulk materials. Although the Encyclopedia will consist predominantly of technical articles arranged in typical alphabetical fashion, one volume will be devoted to the history of NMR. We plan to include a number of short, largely autobiographical sketches from leading figures in NMR and an overall account of the development of NMR, from its "pre-history" in molecular beams to contemporary topics.

As editor of this historical volume, I am concerned that we tell the story of NMR as objectively and completely as possible. I would be interested in receiving suggestions of topics that might not occur to the Editorial Board and anecdotes that will spice up the historical account. Since space will be limited, I cannot promise that any specific item will be included, but I surely would like to have as much information as possible to draw from.

Please contact me with any interesting items. My address, phone and fax numbers are given above.

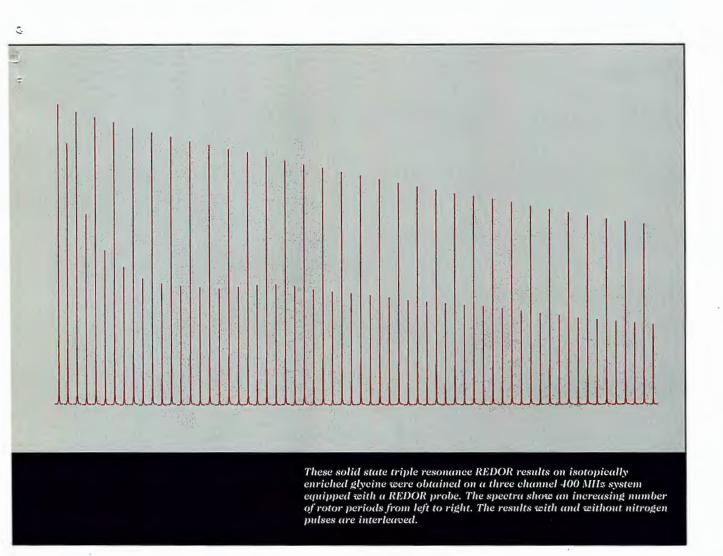
Sincerely,

Edwin D. Becker Section Editor



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July 20, 1993 (received 7/26/93)

Dr. Bernard L. Shapiro Editor/Publisher TAMU NMR Newsletter 966 Elsinore Ct. Palo Alto, CA 94303 USA

Electronic substituent effects on ²Δ isotope shifts of 4-substituted benzanilides

Dear Professor Shapiro:

We have been studying a series of para-substituted benzanilides in terms of theoretical interest concerning the substituent influence on isotope shifts transmitted across the D-labelled nitrogen. In these series of compounds a trend towards larger ²\Delta isotope shift with increase of the electron-attracting character of the substituent is clearly evident at the carbonyl carbon $^2\Delta C = O(ND)$ and at C-1' of the anilinio ring $^2\Delta C - 1'(ND)$ (see Table 1). This behavior differs from the finding in acetanilides substituted at the para position in the anilinio ring¹, in which the size of the two $^2\Delta$ isotope shifts owing to the neighboring carbons to the NH(D) site vary in opposite direction. This means that an acceptor substituent induces larger ² \Delta shifts at CAr and smaller shifts at C=O. The change in the response of the two two-bond isotope shifts in these systems can be related to the corresponding changes in the contribution of mesomeric structures induced by the electronic effects of the substituent. In fact, an electron-withdrawing substituent at the para position in the benzoyl ring of benzanilides is expected to cause an increase of the mesomeric interaction between the nitrogen atom lone pair and the carbonyl group, with the consequent increase of the C-N amide bond order, whereas the same electron-withdrawing group at the para position in the anilinio ring, such as in acetanilides 1, is expected to cause a competitive delocalization of the nitrogen lone pair in the anilinio ring, which in turn reduces the C-N amide bond order. This reasoning can explain why for an amide carbonyl group the large $^2\Delta C$ =0(ND) isotope effect is not always related to a large ${}^{2}\Delta C_{Ar}(ND)^{2}$.

From the above qualitative discussion it might be expected that the two $^2\Delta$ isotope shifts in benzanilides are mutually interdependent. This interdependence is expected to occur via a cross-conjugated pathway arising from the D-labelled nitrogen, and thus, the observed magnitudes of the $^2\Delta$ shifts at the C=O and C-1' could be the result of a balance between them, rather than a proportionality. Consequently, the influence of the substituent on the two two-bond isotope shifts observed at the carbons sharing a D-labelled nitrogen of the type $C(sp^2)-NH(D)-C(sp^2)$ could be better described by a single correlation embracing both $^2\Delta$ isotope shifts. In fact, linear relationships exist between the average values of $^2\Delta C=0$ and $^2\Delta C=1$ with either $\delta^1H(NH)$ (Eq. 1, r = 0.993) or with ^{15}N SCS (Eq. 2, r = 0.998). This behavior suggests that $^2\Delta$ isotope shifts could be used as an independent tool to prove the substituent

perturbation of the Π -electron structure in conjugated systems.

Equation		У	m	X	Ь	r
1	`	$^2\Delta(C=0+C-1')/2ppm$	12.15	δ ¹ H(NH)ppm	-34.53	0.993
2		$^{2}\Delta(C=0+C-1')/2ppm$	1.38	¹⁵ N SCS ppm	90.60	0.998

Table 1. Deuterium isotope shifts (in ppb)^a and NMR parameters (in ppm) of 4-R-benzanilides.

$$\begin{array}{c|c}
R & 3 & 2 & H(D) \\
4 & 1 & 1 & 6' \\
5 & 0 & 2' & 3'
\end{array}$$

R	$^{2}\Delta(C=0)$	²∆(C-1')	δ ¹ H(NH) ^b	15N SCS
$N0_2$	88.1	99.4	10.59	2.3
CO ₂ Me	85.3	100.1	10.48	-
Br	84.8	98.1	10.35	-
C1	84.4	98.9	10.35	0.7
F	83.1	98.6	10.29	-
Н	82.4	98.3	10.29	0.0
Me	81.4	97.7	10.20	-0.9
0Me	80.0	96.5	10.11	-1.8
NMe ₂	76.9	94.1	9.90	-3.6

 a 0.6 M in DMSO- d_{6} ; negative sign: deshielding; b 0.6 M in DMSO- d_{6} ; c In DMSO- d_{6} ; values calculated from Ref. 3; a positive value indicates a downfield shift from R=H.

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- 2. P.E. Hansen, A. Kolonicny and A. Lycka, Magn. Reson. Chem. 30, 786 (1992).
- H. Suezawa, T. Yuzuri, M. Hirota, Y. Ito and Y. Hamada, Bull. Chem. Soc. Jpn. <u>63</u>, 328 (1990).

Sincerely yours,

Martha S. Morales-Rios

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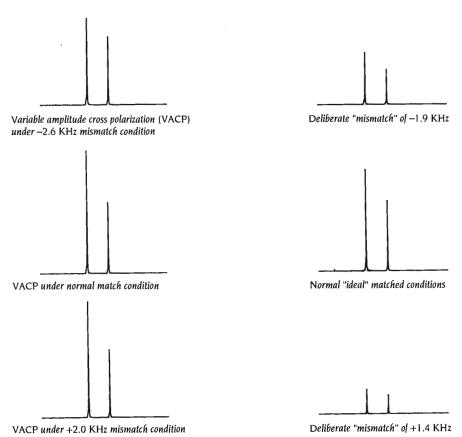
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The introduction of the Variable Amplitude Cross Polarization (VACP) technique by Professor Steve Smith at the Yale University at the 1993 ENC may completely change the approach to optimization of solid state NMR experiments. This technique produces high quality solid state CP/MAS data, even on samples where the match condition may be impossible to optimize, such as weakly coupled systems. This opens up great potential for effective quantitation and comparison of data from different samples.

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Chemagnetics would like to thank Professor Steve Smith for suggestion of this work and useful discussions during its implementation.



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August 3, 1993 (received 8/11/93)

Dr. Bernard L. Shapiro, Editor TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303.

Carbon-13 Chemical Shift Tensors from Carbon-13 Depleted Compounds

Dear Barry,

Carbon-13 chemical shift tensors provide information on the dynamic state of the molecules in the solid state. Aromatic sites are usually difficult to label by chemical methods and one then needs to record the signal using compounds of natural isotopic abundance. Beside the long averaging time that such experiments require there is often a difficulty due to overlap between signals from other carbons having similar chemical shifts values. Specific examples are the hexaalkanoyloxy derivatives of benzene, BHAn, which in the solid state exhibit several different phases.1 To check whether these solid-solid transitions involve the setting-in of whole-molecule reorientation we were interested to record the aromatic carbon-13 signal from the benzene ring in the various solid phases. This signal falls however in the same range as that of the carboxylic carbon. To eliminate the signal of the latter we prepared the homologue BHA6 with carbon depleted from the 13-isotope using CO₂ with more than 99.9% carbon-12 that we happened to have in stock. The carbon-13 NMR spectra obtained from this compound at two temperatures corresponding to solid I and solid II are shown in the figure.² In solid II a spectrum corresponding to a biaxial tensor was obtained with $\sigma_{11}=197.0$, $\sigma_{22}=139.2$, σ_{33} =68.9ppm (relative to TMS), while in solid I the spectrum was that of a uniaxial tensor with $\sigma_{11} = \sigma_{22} = 168.9$, $\sigma_{33} = 67.4$ ppm. The results clearly indicate that in solid II the benzene moiety is static while in solid I it rapidly reorients about the σ_{33} direction (normal to the benzene plane).

Sincerely yours,

A. Maliniak

G. Greenbaum

R. Poupko

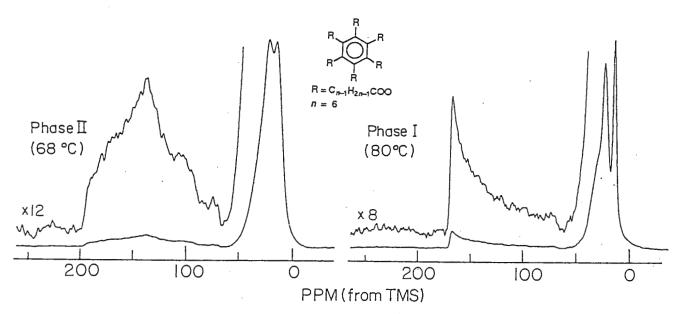
R. Fourflo.

H. Zimmermann

Z. Luz

^{1.} M. Sorai, K. Tsuji, H. Suga, H. Seki, Molec. Cryst. Liq. Cryst. <u>59</u>, 33 (1980).

^{2.} A. Maliniak, S. Greenbaum, R. Poupko, H. Zimmermann, Z. Luz, J. Phys. Chem. <u>97</u>, 4832 (1993).



Carbon-13 NMR spectra at 75 MHz of a powder sample of BHA6 in which the carboxylic carbon is depleted to less than 0.1% ¹³C. Delay time between pulses, 90s; total accumulation time, 3 days for each spectrum.

August 16, 1993

Dear Barry:

The Twenty-Fifth Southeastern Magnetic Resonance Conference will be hosted by the Departments of Chemistry, Biochemistry, and Physics of the University of Florida on October 29-30, 1993. The program will include invited lectures, contributed papers, posters and exhibits. Topics will include NMR of proteins; solid-state NMR; magnetic resonance imaging; NMR spectroscopy in vivo; magnet and coil design; ion cyclotron resonance; EPR spectroscopy; NMR at very low temperatures. Keynote talks will be given by Alex Pines, Charles Johnson, Stan Opella, Alan Marshall, Raymond Andrew, and Angela Gronenborn. Jack Crow of the National High Magnetic Field Lab will give an overview of developments in the Laboratory and Denis Markiewicz will discuss design of high-field magnets.

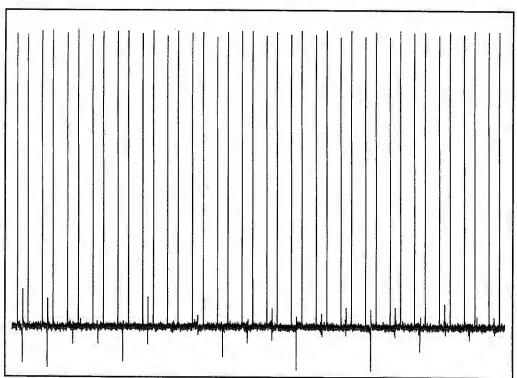
Rooms for Thursday through Saturday nights have been reserved at the Holiday Inn, University Center, Gainesville, FL, and those planning to attend the conference may call the hotel at 904-376-1661. A registration reception at the Holiday Inn is planned for Thursday evening, and the sessions will continue until mid-afternoon, Saturday. Gainesville is served by Delta and U.S. Air. Further information and registration materials may be obtained by writing or calling Wallace S. Brey, Chemistry Department, University of Florida, Gainesville, Florida, 32611-2046; phone 904-392-0520; FAX 904-392-8758.



SPECTROMETER STABILITY AND THE DIGITAL LOCK

The AVANCETM DMX series of spectrometers features the innovative Digital LockTM technology, which is also available as an option on AMX and ARX series spectrometers. The advantages of the Digital Lock in difficult environments, such as in the vicinity of subways, elevators, compressors, or other magnetic fluctuations found in most major facilities, has been extensively documented.

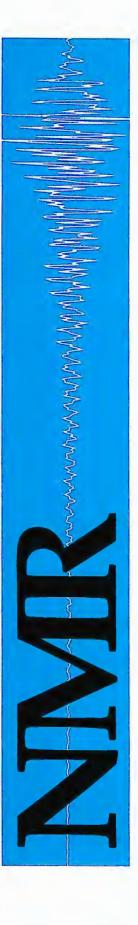
However, the **Digital Lock fundamentally improves the stability** of the AVANCE DMX spectrometer, even under favorable and near-ideal environmental conditions. In order to prove this point, we recorded the most demanding version of a Spin-Echo Difference (SED) experiment on a well shimmed sample of 3% chloroform. We used the absolute minimum of 2 scans (not 8-16 scans), no line broadening, and the Digital Lock. The 600 MHz system was located in a good environment without known magnetic and other disturbances.



20 Spin-Echo Difference experiments; 3% CHCL₃, NS=2, LB=0 (Digital Lock, good environment)



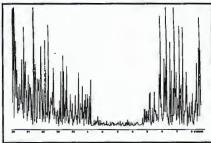




We found in 20 out of 20 experiments, the peak-to-peak height of the suppressed centerband never exceeded 50% of the height of the satellites. With a traditional analog lock system in a similar environment, it was more typical to quote the following specification: "in 7 out of 10 experiments the centerband peak-topeak height will not exceed two times the height of the satellites".

Thus, under fairly good conditions the Digital Lock still provides an improvement of a factor of 4, and on 20 out of 20 experiments! The very stringent requirements for stability of modern 2D/3D/4D NMR experiments, especially those involving cancellations and inverse detection, hardly require any discussion. Suffice it to point out, that the new generation of gradient versions of these experiments are even less forgiving, because there is no phase-cycling which would attenuate any imperfections and instabilities.

The most impressive example, comparing the conventional analog system with the digital lock under unfavorable conditions, was provided from an AMX 600 in Professor Richard Ernst's lab at the ETH next to the Zürich street cars. In the presence of magnetic perturbations the Digital Lock gives an improvement of approximately a factor of 10-25 over traditional analog lock technology in spin-echo difference suppression experiments. (TAMU NMR Newsletter No. 408, 9/92, pages 12-14)



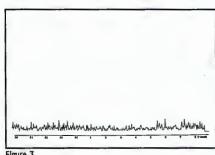


Figure 3

Both figures show an inverse spectrum of chloroform. The center line is plotted every 3 minutes, during a 24 hour period. Figure 2 is using the analog lock, figure 3 is using the digital lock. Both are run under conditions of external field disturbances from the Zürich street cars.



NSR Center

Nijmegen SON Research Center for Molecular Structure, Design and Synthesis SON National HF-NMR Facility Toernooiveld, 6525 ED Nijmegen The Netherlands

A joint Institute of the University of Nijmegen and the Netherlands Foundation for Chemical Research Nijmegen, July 29, 1993 (received 8/2/93)

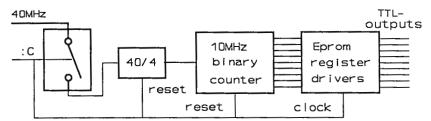
AM-DISCO

Dear Professor Shapiro,

In recent years a number of novel experiments for multinuclear NMR have been developed that cannot be performed on standard AM- or older type spectrometers. A case in point is a set of triple resonance experiments developed by A. Bax for assigning 13C/15N-enriched protein spectra in particular. In this set there are two, the HCACO and the HCA(CO)N experiments, that require, besides powerswitching, two 13C frequency sources with independent phase-cycling. To get these experiments smoothly running on an AM-type spectrometer we have built a double digital synthesizer. When one wants to exploit all of its possibillities, one ends up with ten different functions just for its phase- and frequency control. So we ran into trouble because of the limited number of TTL-outputs on our Bruker AM-spectrometers. These spectrometers "only" have a total of eight to ten freely programmable TTL-outputs available.

To solve this problem now and to avoid more of this kind in the future, we have built a device, that extends the possibilities of TTL-control of any type of spectrometer by a factor of almost infinity. Triggered by the number of monitoring LED's on its front we called this device "AM-DISCO" and thought that the abbreviation might stand for "Decoder for Innovative Spectrometer COntrol".

The basic idea of this apparatus is to use just one TTL-output to control one entire device or external RF-channel, regardless of the number of functions it has. To get this done, we use a pulsetime-decoder. Depending upon the length of a TTL-pulse (a ":C"-pulse in our case) we activate a series of TTL-outputs to control the external devices. The principles of operation are explained with the help of the simplified blockdiagram below.



The blockdiagram shows a binary counter with a timebase of 100ns that is synchronised to the main clock (40MHz) of the timing device (process controller) of the actual spectrometer. When the input pulse (":C") is "high", for instance for 1.4 μ s, the counter will reach its binary value fourteen. This number now acts as an address for the EPROM. The contents of this memorylocation is the desired combination of active and non-active TTL-outputs. Both the number of addresses and the wordlength of this EPROM can be virtually infinite. This means that the possibillities of the device are only limited by the decoding time it takes to activate the different output combinations. (100ns. per combination).

May this contribution give new hope to desperate spectroscopists. Best regards,

J.W.G. Janssen (haja@nmr.kun.nl)

I/W.M. van Os (jaos@nmr.kun.nl)

P.S. Please contribute this letter to E. de Boer's account.



Department of Chemistry

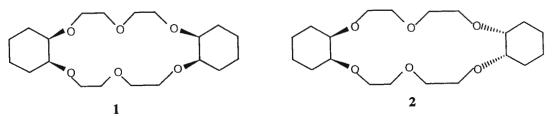
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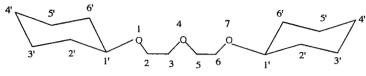
Dr. B. L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto California, 94303 USA July 6,1993 (received 7/31/93)

Title: Conformational Interchange In Dicyclohexanodiethyleneglycol As Studied By ¹³C NMR

Conformational equilibria in <u>cis-syn-cis</u> and <u>cis-anti-cis</u> dicyclohexano-18-crown-6 ethers (1 and 2 below) are amenable to examination via 13 C NMR at low temperature. Generally, the detected processes are cyclohexane ring inversions and conformational free energy differences are small (< 1.2 kJ mol⁻¹), due to the <u>cis-1,2-disposition</u> of the oxygenated substituents.

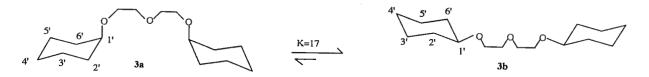


The 1,7-dicyclohexanodiethyleneglycol molecule (3,below) is a structural constituent of 1 and 2 and it is of interest to determine if it possesses any conformational equilibria detectable by NMR at low temperature.



3

The 125 MHz ¹³C NMR spectrum of **3** shows 6 lines at 275K, indicating that all conformational interconversions are rapid on the NMR timescale. At 205K in CD₂Cl₂ solution, resonances for C1', C2',6' and C3',5' and C4' appear as doublets with relative intensities in each case of <u>ca</u>. 17:1. Of course the usual precautions regarding nOe differences and T₁ variations must be taken, but it appears from these results that we have an interconvesion between **3a** and **3b** (below) in which the conformational free energy difference is ca. 5 kJmol⁻¹.



We are currently exploring the influence of alkali metal cations on this and related equilbria.

G.W.Buchanan 746 13.
Professor of Chemistry,

Director, Ottawa-Carleton Chemistry Institute

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Input VSWR Pulse width Duty cycle

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Amplitude droop

Phase change/output power

Phase error overpulse

Noise figure

Output noise (blanked)

Blanking delay

Protection

10–90 MHz 1000 W 100 W

0-900 W

 $65 \, dB$ $\pm 2 \, dB$

50 ohms < 2:1

20 ms Up to 10% 250 ns typ.

5% to 20 ms typ.

10° to rated power, typ. 4° to 20 ms duration, typ.

11 dB typ.

< 20 dB over thermal

< 2 μs on/off, TTL signal

1. VSWR: infinite VSWR

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4. Over temperature

Supplemental characteristics:

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2. RF output: Type N (F) 3. Noise blanking: BNC (F)

4. Interface: 25 pin D(F), EMI filtered

Indicators, front panel

1. AC power on

5. Over temperature

2. Peak power meter

6. Over drive

3. Over pulse width

7. CW mode

4. Over duty cycle

System monitors

1. Forward/Reflected RF power

2. Over pulse width/duty cycle

3. DC power supply fault

4. Thermal fault

Front panel controls

1. AC power

3. Duty cycle

2. Pulse width

Cooling

Operating temperature

AC line voltage

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■ LOVELACE MEDICAL FOUNDATION

Dr. B. L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

July 23, 1993 (received 7/28/93)

An Unusual Velocity Calibration Device

Dear Dr. Shapiro,

We recently measured the velocity of water flowing in a curved duct of rectangular cross-section, which we installed in special slots cut into an 80.3 MHz birdcage rf probe. Figure 1 shows the geometry. Three orthogonal velocity components were measured in a slice perpendicular to the duct axis at the site where the duct had curved 90°. We used a "phase contrast" velocity measurement technique to encode velocity. Phase contrast velocimetry is a difference method that relies on phase shifts induced by motion along a field gradient. The velocity sensitivity of a gradient waveform is proportional to the "first moment" of the gradient

waveform (e.g., $\int t \cdot G_x dt$ for the x-direction, with TE = echo time). 2-D images with and without velocity sensitivity are used to

compute velocity. In our view, it is necessary to calibrate imaging sequences used for this type of measurement, and therein lies the utility of the device shown in figure 2, which was assembled from pieces supplied by Greg, Lauren and Peter Icenogle. This material is non-magnetic, non-conductive, easily assembled, and inexpensive. The shaft extending to the right was rotated via a remote motor with the device positioned so that a cross-section of the water bottle could be imaged. The motor speed was adjusted to produce 15 rpm rotation of the bottle, and after a few hundred seconds (several times R^2/ν) velocity images were acquired. The water rotated as a rigid body: velocity components were $v_x=1.6$ y and $v_v=-1.6$ x cm/s. This provided calibration for the transverse, or secondary, velocity components in the duct cross-section. Because access to the imaging volume is limited it is difficult to produce controlled motions in these directions without a device such as this. The axial, or streamwise, component was calibrated by timed collection with the square duct in situ.

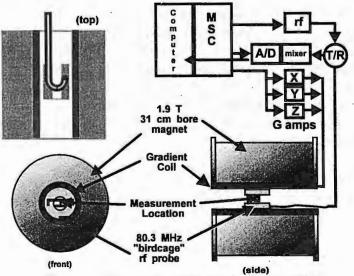


Figure 1. Three cross-sections of the layout of duct installed in the modified birdcage.

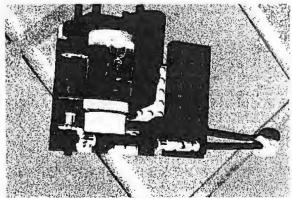


Figure 2. LEGO ™ right angle turn velocity calibrator.

Sincerely,

Steve altobell

Steve Altobelli

La Chile dine

ZENECA

Dr. B.L. Shapiro, Editor TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

(received 8/13/93)

ZENECA Ag Products

Western Research Center 1200 S. 47th Street Box 4023 Richmond California 94804-0023

Telephone (510) 231-1000 Fax (510) 231-1368 Telex 172197

Subject: Nitrogen Chemical Shift Assignments of D-Fructose Oxime

Dear Barry:

Keto-monosaccarides exist in many different tautomeric forms in aqueous solution. The linear "keto" tautomeric form is normally the least populated. Oxime derivatives of monosaccarides lock the sugar into predominantly the linear form and may serve as a good first approximation of the underivatized linear form's conformation.

D-(2-15N)Fructose oxime was synthesized using N-15 enriched hydroxyl amine hydrochloride, for a mass spectrometry experiment. Since the material was available, it was of interest to measure the nitrogen chemical shifts of D-fructose oxime. The spectrum was recorded at ambient temperature and two resonances appear at 345.1 and 341.4 ppm with similar intensities (referenced to external formamide @ 112.4 ppm).

It has been shown that ¹⁵N resonances of Z isomers of acyclic ketoximes appear upfield of the resonances of the corresponding E isomers¹. To verify whether this trend was consistent with carbohydrate oximes a long-range ¹⁵N-¹H 2D HETCOR experiment was run. Indeed the results are consistent with this trend. The resonance at 345.1 ppm correlates with H-3E and H-1E. The upfield resonance correlates with H-3Z, H-1Z and H-4Z.

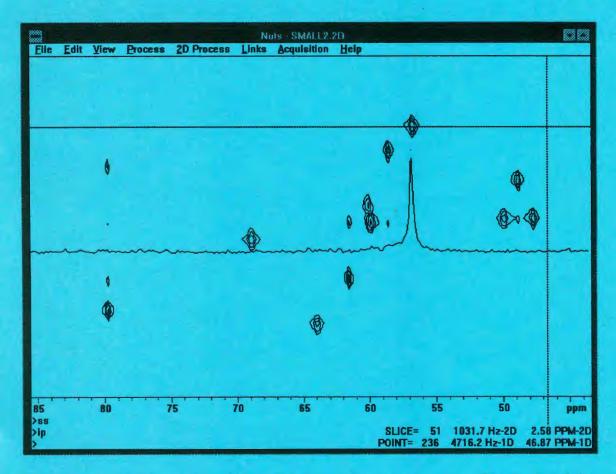
One bond carbon-nitrogen coupling constants were measured from the 13 C NMR spectrum. The results indicate that 1 J(C-N)E=(-)2.98 Hz, whereas 1 J(C-N)Z=(-)3.15 Hz. The values are expected to be negative based upon earlier work.²

1)R.E. Botto, P.W. Westerman and J.D. Roberts, *Organic Mag. Res.*, <u>11</u>, 550, 1978. 2)G.W. Buchanan and B.A. Dawson, *Can J. Chem.*, <u>56</u>, 2200, 1978.

Sincerely,

Joseph R. Snyder Ph.D. Senior Research Chemist

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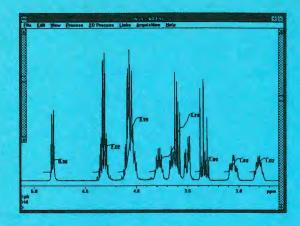
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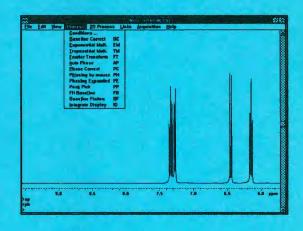
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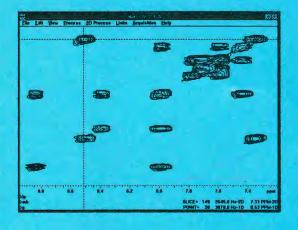
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46560 Fremont Blvd., #418 Fremont CA 94538-6482 Telephone: (415) 683-8595 FAX: (415) 683-6784 Dr B L Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto California 94303 United States of America

(received 8/2/93)

Dear Barry,

Applications of pulsed field gradients in high-resolution NMR spectroscopy.

Last year we acquired a pre-emphasis and Z₀ compensation unit for our AMX500 spectrometer and have finally managed to obtain a working configuration (after a new analogue router board and other miscellaneous but essential modifications - beware retrofitting one of these to an AMX!) Currently we are testing it out on the standard experiments, before moving on to more advanced things.

We have been exploring the use of gradient pulses to discriminate between protons bound to an isotopically enriched nucleus (eg. ¹⁵N) and those bound to the abundant but not interesting isotope (¹⁴N). In the figure, the bottom spectrum is a ¹H spectrum of the peptide VTHRLAGLLSRSGGVVKNNFVPTNVGSKAF, where all valine, leucine, alanine, glycine and phenylalanine residues have been selectively labelled with ¹⁵N. The middle spectrum is ¹⁵N edited, ie. displaying protons bound to ¹⁵N only, acquired with a standard Bruker AMX pulse sequence. The top spectrum is ¹⁵N-suppressed, ie. displaying protons not bound to ¹⁵N. The pulse sequence used was:

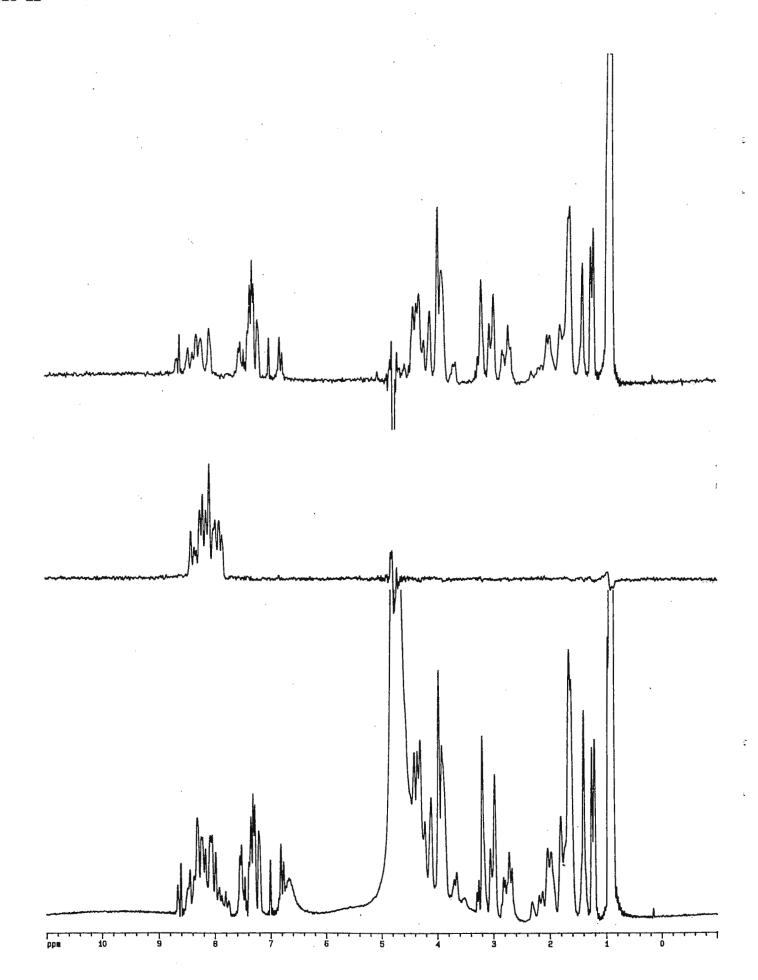
This filter may also be used as an input for various nD experiments, to give X-edited or X-suppressed spectra. We are currently applying this method to the study of protein-ligand interactions.

Yours sincerely,

ave Keid dosley Macdachlon Introdud.

David G Reid Lesley K MacLachlan Julia A Hubbard David M Doddrell

Ian M Brereton



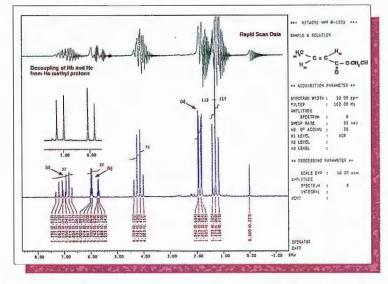
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IKE INSTITUTE FOR CLINICAL AND EXPERIMENTAL MEDICINE

DEPARTMENT OF DIAGNOSTIC AND INTERVENTIONAL RADIOLOGY HEAD: JAN PEREGRIN, MD, PhD

> June 22, 1993 (received 8/2/93)

Dr. B. L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, California 94303

Dear Dr. Shapiro,

ADIABATIC PULSE ON GBS2

We are concerning in phosphorus in vivo spectroscopy using surface coil with large diameter. Because our old Siemens GBS 2 system is not too flexible to generate new pulses, we developed new pulse generator on IBM/PC, which extends feasibilities of our system.

First pulse, which we tested, was adiabatic pulse BIR-1 [1]. It is modulated by functions cosinus in amplitude and sinus in phase. For testing we have been using phosphorus concentration of Na $^+$, H $^+$, PO $_4^{\ 3-}$ determined fantoms. The different determined the pH and consequently chemical shift of the samples.

The first comparison of the pulses has been performed using the simplest excitation sequence. For sinc and BIR-1 pulses we have measured signal intensity dependence on the transmitter voltage. According to published works the adiabatic pulse needs much more higher transmitter voltage then sinc one. The shape of the dependence and maximum intensity obtained for both pulses shows no significant differences.

Practically same results we have obtained with sequence ISIS, we have been using rectangular respective adiabatic $\pi/2$ -pulses for the excitation and hypersecant π -pulse for the inversion. Slightly higher signal intensity in the case of the adiabatic pulse hardly balances the need of the considerably higher voltage, which can make impossible to use it in clinical in vivo practice.

Significant improvement of the signal/noise ratio has been brought by the adiabatic pulse used for excitation in the presence of the magnetic field gradient (2mT/m). Gradient has influenced the spectra obtained with adiabatic pulse only slightly, while spectra obtained with sinc pulses (used in the absolutely same sequence) were practically disturbed by the noise (Figs. 1-3).

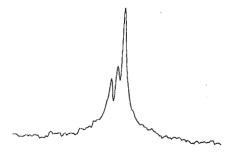
In the view of our results we can expect much more homogeneous excitation in the presence of magnetic gradient using adiabatic pulses designed by our new pulse generator in comparison with by now used pulses.

Yours sincerely,

Mil Hijek

hit Herguel

 Ugurbil K., Garwood M., Bendall M. R.: J. Magn. Reson. 72, 177-185 (1987)



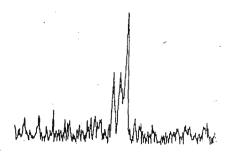


Fig. 1: Excitation by SINC pulse without gradient

Fig. 2: Excitation by SINC pulse gradient = 2mT/m

Fig. 3: Excitation by pulse B[R-1 gradient = 2mT/m



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Institut für organische Chemie

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PD Dr. P. Bigler Institute of Organic Chemistry University of Berne Frelestrasse 3 3012 Bern Switzerland

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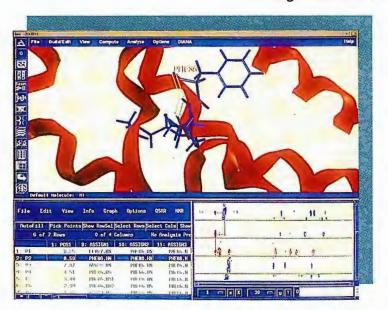
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Sandia National Laboratories

Albuquerque, New Mexico 87185

Departments 1811 and 1812 July 22, 1993

(received 8/4/93)

Dr. Barry L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

Dear Dr. Shapiro,

Structural Assignment of a Regioisomer of C₆₀H₄

We are currently investigating the chemistry of buckminsterfullerene, C_{60} , particularly the hydrogenation of C_{60} to yield a single regioisomer of $C_{60}H_2$ and several regioisomers of $C_{60}H_4$. During this work, we have found 1H NMR to be extremely valuable in the characterization of the $C_{60}H_4$ isomers.

Treatment of C_{60} with borane-THF complex in toluene followed by protonolysis with water yields 1,2- $C_{60}H_2$ (addition of hydrogen across a 6,6 ring fusion) as the major hydrogenated product. The reaction mixture also yields three isomers of $C_{60}H_4$ separable by preparative HPLC (less than 2% total yield based on the uncorrected peak heights). The 400 MHz ^{1}H NMR of one of these regioisomers is shown in Figure 1. This isomer of $C_{60}H_4$ (Figure 1a) has a distinctive ABB'A' type of coupling pattern centered at 5.03 ppm in toluene- d_8 . This type of coupling pattern is only possible for the 1,2,3,4 isomer of $C_{60}H_4$ (see Figure 2), assuming hydrogenation only occurs across 6,6-ring fusions.

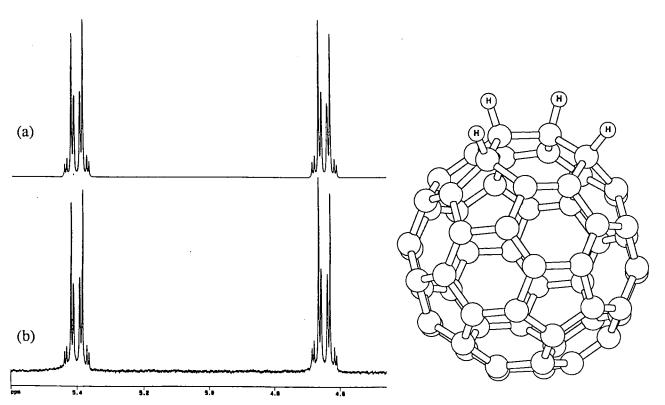


Figure 1: Simulated (a) and experimental (b) 400 ¹H NMR spectra of 1,2,3,4-C₆₀H₄ in toluene-d₈.

Figure 2: 1,2,3,4 isomer of of C₆₀H₄.

We verified this structural assignment by simulating the ^{1}H NMR spectrum of 1,2,3,4- $^{\circ}C_{60}H_4$ using the NMRSIM program for the GE 1280 data station. The following parameters were used to generate the spectrum in Figure 1b: $\Delta\delta = 300$ Hz, $^{3}J_{AB} = 14.1 \pm 0.5$ Hz, $^{3}J_{BB'} = 9.8 \pm 0.5$ Hz, $^{4}J_{AB'} = -0.2 \pm 0.5$ Hz, and $^{5}J_{AA'} = 1.7 \pm 0.2$ Hz (see Figure 3). Line broadenings of 1.0 Hz and 0.2 Hz were used in the simulated and experimental spectra respectively. The agreement between the experimental and calculated spectra is excellent even the fine splittings and relative intensities are reproduced. Changing the parameters slightly results in significant differences between the experimental and simulated spectra.

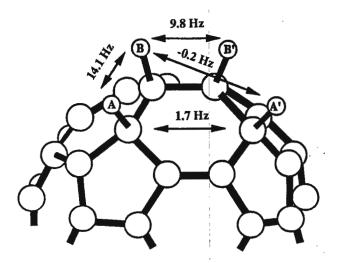


Figure 3: Coupling constants of 1,2,3,4-C₆₀H₄.

The AB (=A'B') coupling of 14.1 Hz is in agreement with the value of 15.7 Hz observed for the vicinal hydrogens of $C_{60}H_2$. Although smaller in magnitude, the value of 9.8 Hz for the BB' coupling is also reasonable for cis, vicinal hydrogens. The AB' coupling is almost zero, consistent with the geometry between the two C-H bonds, and the AA' coupling is 1.7 Hz due to small coupling through the double bond in the six-membered ring. Overall, the observed coupling pattern is clearly unique to the 1,2,3,4 isomer of $C_{60}H_4$.

The formation of 1,2,3,4-C₆₀H₄ as the major tetrahydro isomer is intriguing because this isomer is calculated by Matsuzawa, et al.,² to be the second *highest* energy isomer of the eight possible isomers resulting from hydrogenation across the 6,6-ring fusions of C₆₀. This may be a result of the reaction kinetics or a limitation of semi-empirical calculation theory which predicts all eight isomers to lie within 3.5 kcal/mole of each other. Further investigations into this question of preferred regiochemistry will certainly involve the use of high field ¹H NMR to identify the isomers of hydrogenated fullerenes.

Best Regards,

Craig C. Henderson

Paul A. Cahill

Roger A. Assink

¹ Henderson, C. C.; Cahill, P. A., Science 1993, 259, 1885.

² Matsuzawa, N.; Fukunaga, T.; Dixon, D. A., J. Phys. Chem. **1992**, 96, 10747.

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83-12327-3	L-Alanine-13C3, N-t-BOC Derivative	99 atom%
85-12266-1	L-Alanine-15N, F-MOC Derivative	99 atom%
85-12241-4	L-Alanine-15N, N-t-BOC Derivative	99 atom%
83-12353-9	L-Aspartic-3-13C Acid, N-t-BOC Derivative	99 atom%
83-12354-7	L-Aspartic-4-13C Acid, N-t-BOC Derivative	99 atom%
85-12259-6	L-Aspartic-15N Acid, F-MOC Derivative	99 atom%
85-12244-8	L-Aspartic-15N Acid, N-t-BOC Derivative	99 atom%
83-12322-4	Glycine-1-13C, N-t-BOC Derivative	99 atom%
83-12365-3	Glycine-2-13C, F-MOC Derivative	99 atom%
83-12326-5	Glycine-2-13C, N-t-BOC Derivative	99 atom%
83-12339-8	Glycine- ¹³ C ₂ , N-t-BOC Derivative	99 atom%
85-12260-4	L-Glutamic-15N Acid, F-MOC Derivative	99 atom%
85-12261-2	L-Glutamic-15N Acid, N-t-BOC Derivative	99 atom%
85-12267-9	L-Glutamine-15N ₂ , α-N-t-BOC Derivative	99 atom%
85-12263-8	L-4-Hydroxyphenylalanine-15N, N-t-BOC Deriv. (Tyrosine)	99 atom%
83-12359-6	L-Leucine-1-13C, F-MOC Derivative	99 atom%
83-12323-2	L-Leucine-1-13C, N-t-BOC Derivative	99 atom%
85-12245-5	L-Leucine-15N, N-t-BOC Derivative	99 atom%
81-12258-2	L-Leucine-2-13C,15N, N-t-BOC Derivative	99 atom% 13C, 15N
83-12324-0	L-Methionine-1-13C, N-t-BOC Derivative	99 atom%
83-12337-2	L-Methionine- ¹³ C ₁ (methyl- ¹³ C), N-t-BOC Deriv.	99 atom%
83-12357-0	L-Phenylalanine, N-t-BOC- ¹³ C, Derivative (carbonyl- ¹³ C)	99 atom%
83-12356-2	L-Phenylalanine-1-13C, N-t-BOC Derivative	99 atom%
83-12372-9	L-Phenylalanine-2-13C, F-MOC Derivative	99 atom%
83-12351-3	L-Phenylalanine-3-13C, N-t-BOC Derivative	99 atom%
83-12350-5	L-Phenly-1-13C-alanine, N-t-BOC Derivative	99 atom%
85-12243-0	L-Phenylalanine-15N, N-t-BOC Derivative	99 atom%
83-12325-7	L-Valine-1-13C, N-t-BOC Derivative	99 atom%
85-12265-3	L-Valine-15N, F-MOC Derivative	99 atom%
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82-00815-2	DL-1,4-Dithiothreitol-d ₁₀ (Cleland's Reagent)	98 atom%
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83-30005-3	D-Lactose-1- ¹³ C · H₂O (Glucose-1- ¹³ C)	98 atom%
81-12211-1	L-Lysine-6- ¹³ C, 6- ¹⁵ N HCI	99 atom% ¹³ C, ¹⁵ N
83-00804-5	2-Mercaptoethanol-1-13C	99 atom%
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Dr.Heinz Sterk

Unser Zeichen:

Dr. Bernhard Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

Proton exchange - selective pulses

Dear Dr. Shapiro:

Proton exchange between e.g. OH, and NH groups has been studied widely in the past, mostly by saturation transfer, solvent saturation or other techniques. Within the context of some investigations concerned on peptides we were interested in proton exchange rates between NH groups and the solvent(H_2O). To unravel overlapping NH protons the selective HOHAHA technique proposed by the group of Bodenhausen came into use. To check the mathematics and the usefulness of a selective inversion instead of a presaturation, a selective inversion (G3) pulse and selective detection pulse, were applied onto a mixture of hydroxy-acetophenone and salicylic-aldehyde where the following exchange can occur.

The measured OH intensities of the salicylic aldehyde at different variable delays (0.001 - 30 s) where used to calculate the lifetime, of the hydroxylic group of the hydroxy-acetophenone and vice versa. The lifetimes, obtained from the fit procedure, are 9s for the hydroxy-acetophenone and 30s for the hydroxyl proton of the salicylic- aldehyde which is in good agreement with the data obtained by Forsen and coworkers. That shows that as long as pulse duration of a selective pulse is negligible compared to the relaxation time, selective techniques can be successfully applied.

Yours sincerely

l.Krawanja

H.Sterl

Dr. Barry Shapiro TAMU Newsletter 966 Elsinor Court Palo Alto, CA 94303 USA

LA TROBE UNIVERSITY CHEMISTRY DEPARTMENT

PS 3 Room 130 La Trobe University BUNDOORA Victoria 3083 Australia

PHONE: (03) 479 2582 FAX: (03) 479 1399 International: 61-3

E-mail: brownlee@latrobe.edu.au



Dear Barry:

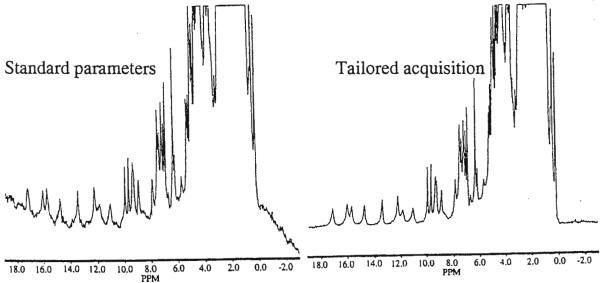
(received 8/5/93)

Flat baselines in 1D and Baseplanes in 2-D Spectra.

Although the standard default acquisition parameters in modern spectrometers are suitable for most samples there are a number of situations in which attention to detail can result in significant improvement of the experimental data resulting in the ability of detecting otherwise unseen peaks.

Our group is involved with the assignments of the contact shifted protons in paramagnetic proteins. Some of these peaks can be seen in the spectra of ferredoxin shown below at shifts from 10-18 ppm. The contact shifted peaks have short relaxation times and hence broad linewidths (100Hz) resulting in peaks of about 2% in height of the resonances from the diamagnetic protons. In 2-D spectra, the identification of cross peaks from these signals depends critically on the quality of data.

Part of the rolling baseline in 1-D spectra results from the distortion of the acquired signal by the filters in the spectrometer, and part is the result of using a real Fourier transform in our Bruker AM system required by the sequential acquisition mode. These problems can be overcome by a) resetting the receiver phase to zero prior to acquisition so that the spectrum requires no zero order phasing, and b) by choosing the dead time and filter width so that the spectrum requires no first order phasing and the spectrum has no baseline roll. In a 2-D spectrum any residual baseline slope can be corrected after F2 transformation.



The acquisition of data in this manner (tailored acquisition) requires considerable patience and care, but the results are worth the effort as can be seen from a comparison of the spectra shown below acquired without (standard parameters) and with (tailored acquisition) this procedure. It is to be hoped that a new generation of spectrometers will soon be available with better digitizers and distortionless filters.

Monoffer

Please credit this contribution to David Kellys subscription.

With best wishes,

Bob Brownlee

Maruse Sadek

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Agricultural Research Service Midwest Area U.S. Dairy Forage Research Center 1925 Linden Drive West University of Wisconsin Madison, WI 53706-1108

(608) 264-5407 E-Mail RALPHJ@MACC.WISC.EDU August 4, 1993 (received 8/7/93)

Dr. B.L. Shapiro 966 Elsinore Court Palo Alto, CA 94303

Dear Dr. Shapiro,

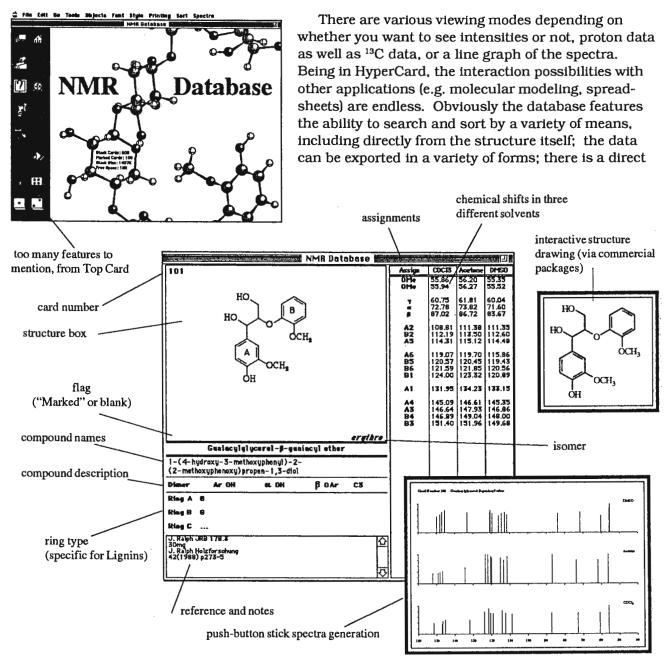
Release of a Plant Cell Wall Compound NMR Database

About 5 years ago, personnel at the US Dairy Forage Research Center and the US Forest Products Laboratory decided that an NMR Database of plant cell wall model compounds, with particular emphasis on lignin model compounds, would be created. This has been a need for quite some time and has been suggested by a number of groups at meetings for the last 15 years. Finally, we could wait no longer for someone else to do it.

Because of the agreement between chemical shifts of carbons in good model compounds and in the polymer itself, assignments in the complex lignin polymer have been substantially made from model compound data. However, lignin spectra are run in a variety of solvents, including DMSO and 9:1 acetone:water for underivatized lignins, and CDCl₃ and acetone for acetylated lignins. For that reason, and because ¹³C chemical shifts can vary substantially with solvent, it was decided from the outset that all compounds would be run in the three solvents, acetone-d₆, CDCl₃, and DMSO-d₆, and under a fixed set of conditions. Most spectra have been run routinely and assigned by reference to the literature and by comparison with related compounds in this database but we couldn't resist fully authenticating some of the more interesting models by the usual complement of COSY, long-range COSY, (perhaps TOCSY), HMQC, and HMBC experiments — it is noted in the database when fully unambiguous authentication has been performed. The result is that the database currently has some 200 compounds, and a rather large number simply entered from literature data. Some of these compounds and data are in advance of the literature; for example, the latest cell-wall cross-linking structures that have been synthesized in these laboratories are already included.

The database format itself underwent considerable development. We wanted a computer-based interactive system with considerable flexibility and some chemical intelligence. At the time, databases were considerably more boring that some of the emerging ones, and our choice at the time was to write it in Apple's HyperCard, to which anyone with a Mac has access. That turned out to be an excellent choice from the point of view of flexibility and customizability. We now retain that as our favored format but have ported the data to Claris FileMaker Pro, a cross-platform application for both MS Windows and Mac OS.

Anyway, we are about to release the Database to the general public. Until we add a little more functionality to the FileMaker Pro version, and address a few of the cross-platform issues, hopefully by the end of this year, it will be available only in HyperCard format and on the Internet, but the price is right — it is free! Users wishing to access our machine via Gopher or FTP on the internet should E-Mail me at RALPHJ@MACC.WISC.EDU so that I can give them access to our server. Documentation is provided in a format which does not require the creating application to view, print, and even steal from (i.e. selection of text and graphics for copying to other applications is supported). In the future we will be able to provide copies on diskette for either platform, and will provide hard copy versions too as required, but there may be a cost associated with these distribution methods. The Internet method will provide access to the continually changing database and is the preferred distribution. There will also be instructions for researchers to add compounds to their own database and to submit those entries (via an export feature) to us for inclusion in the official database. Data in this database can be cited in publications — this is a repository for data that will be available in the long term. We expect this database to grow substantially and to evolve into a complete tool for researchers in the plant cell wall chemistry area.



connection with Chemistry drawing software; and there are many more features than can be described here. It should be quite easy to master, although a manual is provided and perhaps required for some of the less obvious features.

Sincerely, had stalk

John Ralph, William Landucci, Sally Ralph, and Lawrence Landucci.

POSITION AVAILABLE

I have further funding available for a good Ph.D. student with strengths or interests in Organic Synthesis, Plant Cell Wall Chemistry and, of course, NMR to do an exciting project in the area of plant cell wall cross-linking mechanisms. Please contact me at (608) 264-5407, FAX (608) 264-5275, or E-Mail at RALPHJ@MACC.WISC.EDU if you have an interest, want lots of easy-access NMR time, and enjoy cold winters. The US Dairy Forage Research Center is an equal opportunity employer.

Application Profile

Electrical



Description/Application

Radio-frequency coils for magnetic resonance imaging (MRI) and image-directed spectroscopy (IDS) equipment rely on TEFLON® PTFE fluoropolymer for support and insulation of conductors. MRI and IDS are valuable noninvasive techniques for showing soft tissure contrast in medical and biological examinations.

The coil's functions are twofold: to transmit pulses of radio-frequency energy to sample tissue and to pick up faint signals emitted by the tissue in response. It does not come into contact with the tissue.

The unique feature of the coil shown is the application of conductors by electroplating rather than adhesive bonding or lamination. The coil operates in the 100 to 600 MHz range with very high radio-frequency voltage, up to 2 kV. Current sizes range from about 3.50 to 5.25 in. (89 to 133 mm) high with flange diameters from 2.25 to 3.50 in. (57 to 89 mm).

Manufacturer

Polyflon Co., Div, of Crane Co., New Rochelle, NY Phone: (914) 636-7222. Fax: (914) 633-3664.

Co-developer

Centre for Magnetic Resonance, University of Queensland Brisbane, Australia. Phone: (61) (7) 365-4100. Fax: (61) (7) 365-3833.

Benefits Gained from this Development

Higher quality images. Compared with adhesive lamination, use of plated TEFLON reduces the potential for gaps and eliminates the need for adhesives, both of which would decrease reception sensitivity and increase background noise, according to Polyflon.

Faster imaging. Improved image quality reduces needs for image enhancement computations, saving time in examining patients.

Lower production costs. Plating and etching technology is less costly than alternative methods, Polyflon reports.

Material Chosen and Why

The excellent dielectric properties of TEFLON PTFE at radio frequencies and its nonmagnetic nature are critical to image quality.

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Electroplated PTFE also performs well at low temperatures, even under cryogenic conditions. While most substrate materials become brittle below -196°C, PTFE remains ductile at -268°C — nearly the temperature of liquid helium. As a result, elec-

troplated PTFE is suitable for many superconductive designs, as well as circuits and systems in harsh environments, including avionics and space-related systems.

Why Pure PTFE?

Pure PTFE exhibits extremely low loss, with a dielectric constant of 2.06. This value does not change with temperature or frequency, permitting the design of stable broadband circuits with high performance over wide temperature ranges.

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The combination of pure PTFE and Polyflon's proprietary electroplating process produces materials that handle a wide range of applications, from specially shaped antennas and tiny, non-magnetic high voltage trimmer capacitors to medical imaging coils for NMR/MRI systems. Whatever the requirement, Polyflon's electroplated PTFE can fit your needs for today's applications and tomorrow's opportunities.



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Dr. B.L. Shapiro 966 Elsinore Court Palo Alto, CA 94303

(received 8/11/93)
Billerica, August 6, 1993

RF-GRASPTM: Gradient Spectroscopy with B₁- Fields

Dear Barry,

We have recently developed an NMR probe which has in addition to a homogeneous RF coil, a coil which creates a gradient RF field. We are interested in using B_1 - gradients for coherence pathway selection in high resolution NMR spectroscopy. This interest is a reflection of the demonstrated power and versatility of B_0 - gradient methods, and the perceived promise of B_1 - gradient methods. B_1 - gradients share many of the capabilities of B_0 - gradients, and have some advantages including: (1) the gradient is **frequency selective** either for a given spectral line, or for a given nuclear spin, (2) B_1 - gradients have **no influence on the line shape**, (3) B_1 - gradients **do not disturb the lock channel**, (4) **fast switching** (< 10 μ s), (5) more degrees of freedom, variables include the amplitude, length, phase and frequency of the gradient pulse, (6) B_1 - gradients are non-secular, so they do not commute with internal Hamiltonians. Internal Hamiltonians can be selectively suppressed by B_1 - gradients.

In addition, the hardware necessary for RF-GRASPTM experiments is quite modest, requiring only the RF-GRASPTM probe and coil switching electronics (see Figure 1). No additional amplifiers, and no modifications to the console are required.

In the design of an RF-gradient probe, one has to choose the symmetry of the B₁-gradient field. The easiest spin response is obtained from a planar gradient field (see Figure 2), in which only the amplitude of the RF field is spatially dependent. In terms of RF efficiency a better solution is a quadrupolar coil, which creates a radial gradient field (Figure 2) and which has a higher filling factor. A quadrupolar RF gradient has some definite advantages over a planar gradient. The dephasing due to a planar gradient is in a plane perpendicular to the rotating frame axis along which the gradient RF field is applied. In the case of a quadrupolar RF gradient the dephasing is the result of both an amplitude and phase distribution, scattering the magnetization over the surface of a sphere rather than in a plane. Also due to the radial phase distribution the gradient phase does not have to be adjusted to the phase of the homogeneous coil. Finally, a radial RF gradient field, or any non-planar RF gradient field, can be converted into a planar spin response via multiple pulse cycles.

One of the earliest applications of B_0 --gradients was to select either the echo or anti-echo diagonal in COSY spectra¹. This is readily accomplished by introducing a spatially dependent phase shift prior to the mixing pulse so that the mixing pulse transforms both x and y components of the magnetization. A B_1 - version of this experiment² can be implemented by combining the B_1 - gradient into a composite z-pulse, which then influences the spin system exactly as would a B_0 - gradient. The gradient field generated in the RF-GRASP probe has

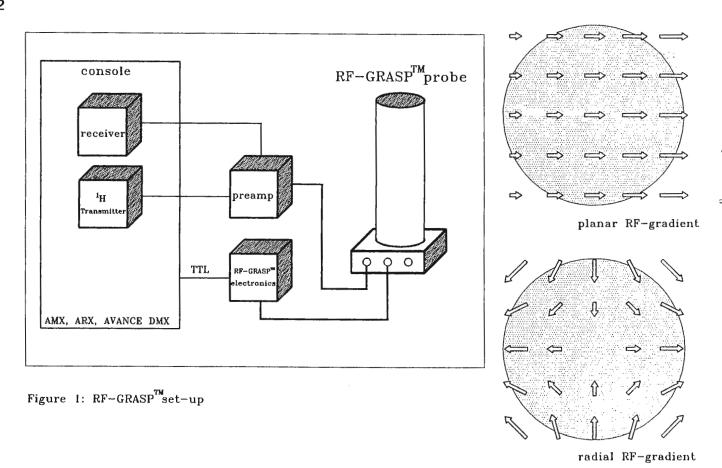


Figure 2: Symmetry of RF-Gradient fields

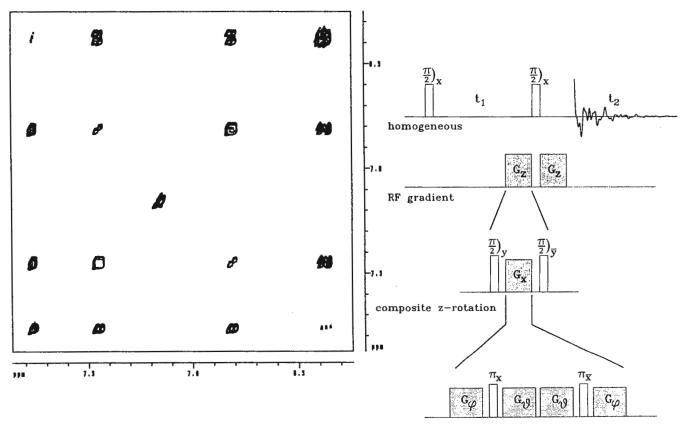


Figure 4: 400 MHz RF-GRASPTMN-type COSY spectrum of 1-chloro 3-nitrobenzene in deuterated benzene

Figure 3: $RF-GRASP^{TM}N-type$ COSY

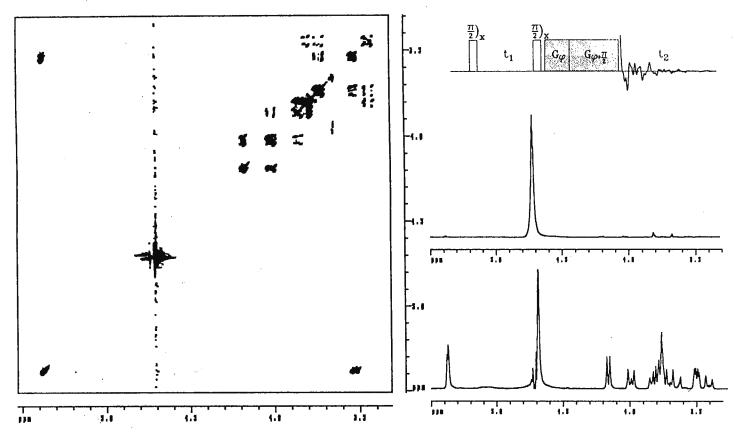


Figure 5: 400 MHZ RF-GRASP DQF-COSY of sucrose in water.

Figure 6: Pulse sequence of the RF-GRASP DQF-COSY experiment and the spectrum of sucrose in water compared to a projection of the DQF-COSY experiment.

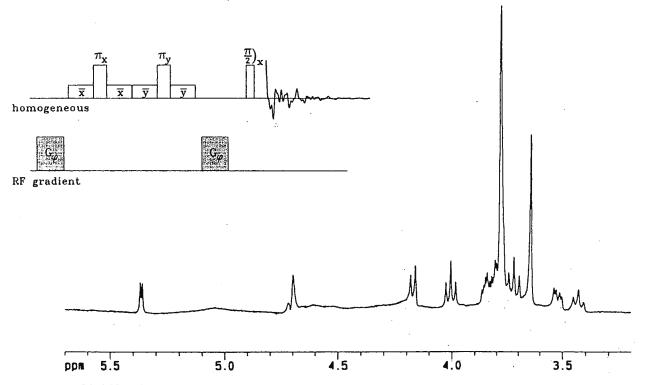


Figure 7: 400 MHz single scan proton spectrum of sucrose in water, obtained with the RF-GRASP water suppression sequence shown above.

a radial symmetry, and so the composite z-pulse is also used to convert this radial field into a planar field. Figure 3 shows the pulse sequence for the P- and N-type selected COSY experiment and the N-type 400 MHz proton cosy spectrum of 1-chloro 3-nitrobenzene in deuterated benzene is depicted in Figure 4. A single scan per t₁ increment is acquired. The RF gradient strength is 15 G/cm, and the total duration of the composite z-pulse is 4 ms.

One of the most exciting applications of GRASP has been in multiple quantum filtering of which an important homo-nuclear example is the double quantum filtered COSY experiment. An RF-GRASP DQF COSY experiment was introduced by Canet and coworkers³ for use with a planar RF gradient. In the case of a radial RF gradient field a small modification of the sequence is used⁴. In Figure 5 a phase-sensitive RF-GRASP DQF COSY spectrum is shown below for a sample of sucrose in water. The spectrum is acquired with a single scan per t_1 increment. The radial RF gradient pulse duration is 700 μ s, at a gradient strength of 15 G/cm. The degree of water suppression is best seen by comparing the one-pulse spectrum with a projection from the 2D dataset (Figure 6).

Recently a successful solvent suppression method was introduced, which combines pulsed field gradients with selective excitation⁵. We have adapted this so-called Watergate sequence to allow the use of B_1 - gradients⁶. The sequence consists of an RF gradient spin echo sequence in which the formation of an echo from the solvent resonance is prevented by applying a selective π -pulse in addition to a hard π -pulse. The suppression sequence can be performed with either planar B_1 - gradients or with radial B_1 - gradients (or indeed with any non-planar RF gradient). In the general case two orthogonal inversions are needed to recover the full echo intensity. The magnetization is refocussed along the z-axis and can be observed after a read-out pulse; alternatively this sequence can be incorporated in multi-dimensional experiments such as during the mixing time of a NOESY experiment. Figure 7 shows the suppression pulse sequence and a single scan 400 MHz proton NMR spectrum of sucrose in water. The RF gradient pulse is 500 μ s at a gradient strength of 15 G/cm. The selective $\pi/2$ durations are 2.8 ms.

These are a few examples of what we have accomplished with B₁- gradients. We hope to report to you soon on new homonuclear RF-GRASPTM experiments, and also on the implementation of B₁- gradients in heteronuclear spectroscopy.

Best Wishes,

Werner E. Maas

Frank H. Laukien

David G. Cory

Dept. of Nuclear Engineering

Massachusetts Institute of Technology

¹ P. Barker and R. Freeman, J.Magn.Reson, **64**, 334, 1985

² W.E.Maas, F.H. Laukien and D.G. Cory, J.Magn.Reson, A103, 115, 1993

³ J. Brondeau, D. Boudot, P. Mutzenhardt and D. Canet, J. Magn. Reson. 100, 611, 1992

⁴ D.G. Cory, F.H. Laukien and W.E. Maas, J. Magn.Reson. in press

⁵ M. Piotto, V. Saudek and V. Sklenár, J. Biomol. NMR, 2, 661, 1992

⁶ W.E. Maas and D.G. Cory, submitted to J. Magn. Reson.



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PE Zeeman 5000 PE Zeeman 3030 PE Zeeman 4100ZL PE HGA-400

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ULTRA-PRECISION NMR SAMPLE TUBES





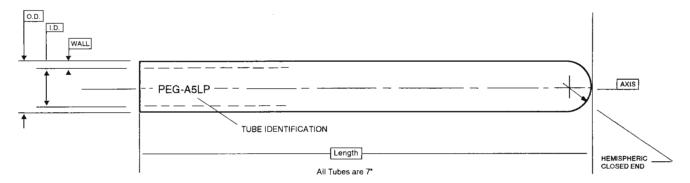
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TUBE SPECIFICATIONS

5 and 10mm x 7" length



ACE Cat. No.	Megahertz (MHz) Inst. Frequency	O.D. inches	O.D. Tolerance	I.D. inches	I.D. Tolerance	Concentricity I.D.to O.D.	Camber (Over 7" Lgth.)
16XXA5LP-07	>500	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0005"	≤ .00025"
160XA5LP-07	360	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0010"	≤ .0005"
1600A5LP-07	200	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0015"	≤ .0010"
1600B5LP-07	150	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0020"	≤ .0015"
1600C5LP-07	100	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0020"	≤ .0020"
16XA10LP-07	360	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0015"	≤ .0005"
160A10LP-07	150	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0020"	≤ .0010"
160B10LP-07	80	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0030"	≤ .0015"
160C10LP-07	60	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0050"	≤ .0020"
	DISPOSABLE NMR TUBES						
1600A5LS-07	80	.1955	+.0000"/0005"	.165	±.005	≤ .003	≤ .002
1600A5RS-07	60	.196	±.003	.165	±.005	≤ .005	≤.002

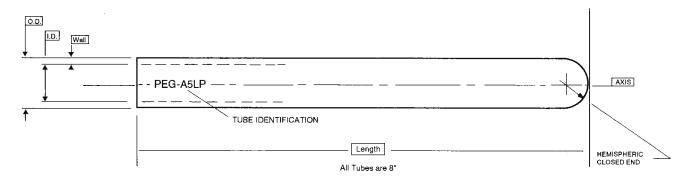
All Tubes are 7" long. Other sizes are available, phone for information.

All Tubes supplied in Package Quantity of 5 or 25 pieces.

ACE	Tube Size	Per	Per
Catalog No.	O.D./Length	Pkg./5	Pkg./25
16XXA5LP-07	5mm/7"	★58.75	★293.75
160XA5LP-07	5mm/7"	★43.25	★216.25
1600A5LP-07	5mm/7"	★37.00	★185.00
1600B5LP-07	5mm/7"	★27.50	★137.50
1600C5LP-07	5mm/7"	★22.50	★112.50
16XA10LP-07	10mm/7"	★93.75	★468.75
160A10LP-07	10mm/7"	★78.75	★393.75
160B10LP-07	10mm/7"	★68.00	★340.00
160C10LP-07	10mm/7"	★51.25	★ 256.25
1600A5LS-07	5mm/7"	★17.50	★ 87.50
1600A5RS-07	5mm/7"	★ 8.00	★ 40.00

TUBE SPECIFICATIONS

5 and 10mm x 8" length



ACE Cat. No.	Megahertz (MHz) Inst. Frequency	O.D. inches	O.D. Tolerance	I.D. inches	I.D. Tolerance	Concentricity I.D.to O.D.	Camber (Over 8" Lgth.)
16XXA5LP-08	>500	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0005"	≤ .00025"
160XA5LP-08	360	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0010"	≤ .0005"
1600A5LP-08	200	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0015"	≤ .0010"
1600B5LP-08	150	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .00 20 ¹¹	≤ .0015"
1600C5LP-08	100	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0020"	≤ .0020"
16XA10LP-08	360	.3937	+,0000"/-,0005"	.3569	±.0005"	≤ .0015"	≤ .0005"
160A10LP-08	150	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0020"	≤ .0010"
160B10LP-08	80	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0030 ^a	≤ .0015"
160C10LP-08	60	.3937	+.0000"/-,0005"	.3569	±.0005"	≤ .0050"	≤ .0020"
DISPOSABLE NMR TUBES							
1600A5LS-08	80	.1955	+.0000"/0005"	.165	±.005	≤ .003	≤ .002
1600A5RS-08	60	.196	±.003	.165	±.005	≤ .005	≤ .002

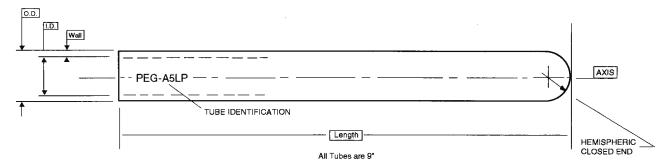
All Tubes are 8" long. Other sizes are available, phone for information.

All Tubes supplied in Package Quantity of 5 or 25 pieces.

ACE	Tube Size	Per	Per
Catalog No.	O.D./Length	Pkg./5	Pkg./25
16XXA5LP-08	5mm/8"	★ 64.75	★323.75
160XA5LP-08	5mm/8"	★ 47.75	★238.75
1600A5LP-08	5mm/8"	★ 40.75	★203.75
1600B5LP-08	5mm/8"	★ 30.25	★151.25
1600C5LP-08	5mm/8"	★ 24.75	★123.75
16XA10LP-08	10mm/8"	★97.50	★487.50
160A10LP-08	10mm/8"	★83.75	★418.75
160B10LP-08	10mm/8"	★72.25	★361.25
160C10LP-08	10mm/8"	★56.25	★281.25
1600A5LS-08	5mm/8"	★19.25	★ 96.25
1600A5RS-08	5mm/8"	★ 8.50	★ 42.50

TUBE SPECIFICATIONS

5 and 10mm x 9" length



ACE Cat. No.	Megahertz (MHz) Inst. Frequency	O.D. inches	O.D. Tolerance	I.D. inches	I.D. Tolerance	Concentricity I.D.to O.D.	Camber (Over 9" Lgth.)
16XXA5LP-09	>500	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0005"	≤ .00025"
160XA5LP-09	360	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0010"	≤ .0005"
1600A5LP-09	200	.1955	+.0000"/0005"	1655	+.0005"/0000"	≤ .0015"	≤ .0010"
1600B5LP-09	150	.1955	+.0000"/0005"	1655	+.0005"/0000"	≤ .0020"	≤ .0015"
1600C5LP-09	100	.1955	+.0000"/0005"	.1655	+.0005"/0000"	≤ .0020"	≤ .0020"
16XA10LP-09	360	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0015"	≤ .0005"
160A10LP-09	150	.3937	+.0000"/0005"	.3569	±.0005°	≤ .0020"	≤ .0010"
160B10LP-09	80	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0030"	≤ .0015"
160C10LP-09	60	.3937	+.0000"/0005"	.3569	±.0005"	≤ .0050"	≤ .0020"
			DISPOSABLE NI	IR TUBES			
1600A5LS-09	80	.1955	+.0000"/0005"	.165	±.005	≤ .003	≤ .002
1600A5RS-09	60	.196	±.003	.165	±.005	≤ .005	≤ .002

All Tubes are 9" long. Other sizes are available, phone for information. All Tubes supplied in Package Quantity of 5 or 25 pieces.

ACE	Tube Size	Per	Per
Catalog No.	O.D./Length	Pkg./5	Pkg./25
16XXA5LP-09	5mm/9"	 ★ 71.25 ★ 52.50 ★ 45.00 ★ 33.25 ★ 27.25 	★356.25
160XA5LP-09	5mm/9"		★262.50
1600A5LP-09	5mm/9"		★225.00
1600B5LP-09	5mm/9"		★166.25
1600C5LP-09	5mm/9"		★136.25
16XA10LP-09	10mm/9"	★ 101.75	★506.25
160A10LP-09	10mm/9"	★ 88.75	★443.75
160B10LP-09	10mm/9"	★ 76.50	★382.50
160C10LP-09	10mm/9"	★ 62.00	★310.00
1600A5LS-09	5mm/9"	★ 22.25	★ 111.25
1600A5RS-09	5mm/9"	★ 9.00	★ 45.00

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August 10, 1993 (received 8/17/93)

Dr. Bernard L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303 U.S.A.

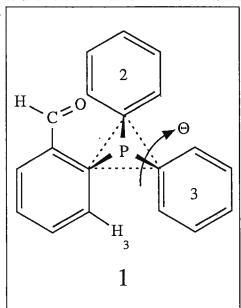
Dear Barry,

Some Properties of Triphenylphosphine (TPP) and 2-(Diphenylphosphino) benzaldehyde (DPPB).

Some time ago the very old news, that TPP and its relatives are ubiquitous as ligands, and so forth, penetrated even my cloister. Some desultory reading implied that a precise analysis of the ¹Hnmr spectrum was not available, other than some early ones in benzene solution (ASIS!), and that an intramolecular rotational potential was also unknown or, at least, unpublished for TPP.

Our efforts in this matter led to the following conclusions (Can. J. Chem. 71, 639 (1993): a) the shift range of the phenyl protons is only 1.7 Hz at 300 MHz in CS_2/C_6D_{12} solution at 300 K; b) the rotation about the P-C bonds is hindered by a potential comparable to kT at 300 K; c) the diamagnetic anisotropy of the two phenyl groups causes a negligible shift of the para proton in the third phenyl group; d) the lone-pair on phosphorus will conjugate with a single phenyl group about 30% as well as a lone-pair on nitrogen under optimum conditions, that is, when it lies in a plane perpendicular to the phenyl group e) in TPP the conjugation involves three phenyl groups which are not oriented on average for optimal conjugation - the model and experiment agree as to the observed shifts; f) yes, a reliable analysis is possible if your linewidths are small enough.

Encouraged by these 'successes', Rudy Sebastian managed a full ¹H nmr analysis of DPPB in CS₂ and in acetone solutions and Rob Schurko ran the ¹³C{¹H}spectra as well. Many relative signs followed from partial decoupling experiments. For each solution one then has 55 chemical shifts and coupling constants, from which a number of useful conclusions can be drawn (see



Can. J. Chem. much later this year). Among them are: a) in the crystal, the C=O bond lies cis to P, whereas in solution the trans conformer predominates and reaction-field theory in first-order cannot account for the relative proportions of cis and trans in the two solvents; b) ⁵J(H, P) can change its sign from ring to ring; c) ⁴J(CHO, P) is -7.1(2) Hz in the trans conformer and ³J(CHO, P) is +29.4(1.3)Hz, a result of interest in the computation of couplings via the O=C-H···l.p. P interaction; d) P is electrically polarized such that the C=O bond is attracted towards a 'positive' region around P, while the C-H bond of the formyl group prefers the lone-pair region of P -all quite sensible in electrostatic terms; e) ¹J(C,P) in TPP is solvent dependent (10%).

Best wishes from

Ted Schaefer

TS/tl

P.S. What is that color of your "Reminder" sheet? It's not pansy-purple, Perkin's violet or madder-violet. Is it, possibly, pontiff-purple?

Ted - The official name of the color is, believe or not, "orchid". I chose the color for the R/U notices because it was the ugliest shade available. I though this might lower the residence time on the desks of the almost delinquent. Hope this satisfies your insatiable thirst for useless knowledge! With best regards, fare.

Table of Contents, cont'd.

Release of a Plant Cell Wall Compound Database	Ralph, J., La	nducci, W., R	alph, S., a	nd Landucci, L.	37
Position Available				. Ralph, J.	38
RF-GRASP TM : Gradient Spectroscopy with B ₁ -Fig	elds Maas ,	W. E., Lauki	en, F. H.,	and Cory, D. G.	41
Some Properties of Triphenylphosphine and 2-(Dipl	henylphosphino)	-benzaldehyde		. Schaefer, T.	51

.

All Newsletter correspondence should be addressed to

Dr. B. L. Shapiro 966 Elsinore Court Palo Alto, CA 94303 U.S.A.

(415) 493-5971 - Please call only between 8:00 am and 10:00 pm, Pacific Coast time.

Deadline Dates

No. 422 (November) 22 October 1993

No. 423 (December) 19 November 1993

No. 424 (Jan. 1994) 17 December 1993

No. 425 (February) 21 January 1994

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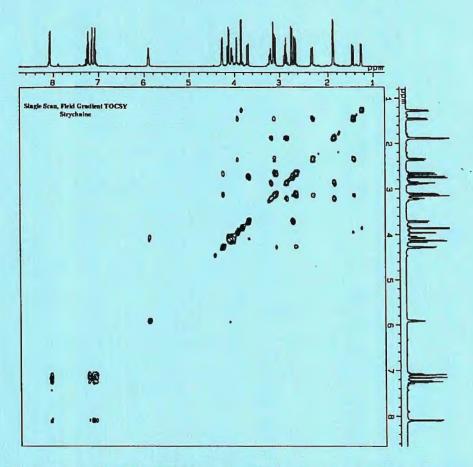
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