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No. 412 January 1993

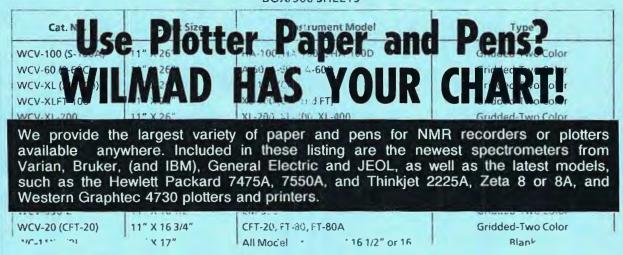
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FORTHCOMING NMR MEETINGS

International Symposium on Solid-State NMR Spectroscopy of Polymers, Keystone, Colorado, February 28 - March 3, 1993; Contact: Barbara D. Hogan, Du Pont, CR&D, P. O. Box 80356, Wilmington, Del. 19880-0356; Phone: (302) 695-4394; FAX: (302) 695-8207.

PITTCON'93, Atlanta, Georgia, March 8-12, 1993; Contact: Dept. 60, Suite 332, Penn Center Blvd., Pittsburgh, PA 15235-5503; Phone: (412) 825-3220, Toll-free: 1-800-825-3221; Fax: (412) 825-3224.

1993 Keystone Symposia on Molecular & Cellular Biology, Taos, New Mexico: March 8-14, 1993, Frontiers of NMR in Molecular Biology - III; Organizers: T. L. James, S. W. Fesik, and P. E. Wright; Information: Keystone Symposia, Drawer 1630, Silverthorne, CO 80498; Telephone: (303) 262-1230.

<u>In Vivo Magnetic Resonances Spectroscopy VI.</u> Symposium on <u>In Vivo Magnetic Resonances Spectroscopy: Focus on Spectroscopic Techniques</u>, St. Louis, Missouri, March 13-14, 1993; Contact: Radiology Postgraduate Education, Room C-324, University of California School of Medicine, San Francisco, CA 94143-0628; Phone: (415) 476-5808; Fax: (415) 476-9213.

34th ENC (Experimental NMR Conference), St. Louis, Missouri, March 14-18, 1993; Contact: ENC, 815 Don Gaspar, Santa Fe, New Mexico 87501; Phone: (505) 989-4573; Fax: (505) 989-5073. See TAMU NMR Newsletter 408, 45.

Advanced School on Magnetic Resonance and Protein Dynamics, Erice, Sicity, Italy, March 15-21, 1993; Contact: Prof. Oleg Jardetzky or Stanford Magnetic Resonance Lab., Stanford Univ., Stanford, CA 94305-5055; Fax: (415) 723-2253; See TAMU NMR Newsletter 411, 50.

High Resolution NMR Spectroscopy (a residential school), University of Sheffield, England, April 1993; Organizer: Dr. B. E. Mann (Sheffield); For information, contact Ms. L. Hart, The Royal Society of Chemistry, Burlington House, Piccadilly, London W1V 0BN, England; Tel.: 071-437-8656.

Gordon Research Conference: Magnetic Resonance, Wolfeboro, NH, July 11 - 16, 1993; Contact: Dr. A. M. Cruickshank, Gordon Research Center. University of Rhode Island, Kingston, RI, 02881-0801; (401) 783-4011 or -3372; FAX: (401) 783-7644.

35th Rocky Mountain Conference on Analytical Chemistry, Denver, Colorado: July 25-29, 1993; Contact: Patricia L. Sulik, RML, Inc., 456 S. Link Ln., Ft. Collins, CO 80524; Phone: (303) 530-1169.

12th Annual Scientific Meeting and Exhibition of the Society of Magnetic Resonance in Medicine, New York, NY, August 14-20, 1993; Contact: SMRM, 1918 University Ave., Suite 3C, Berkeley, CA 94704; Phone: (510) 841-1899; FAX: (510) 841-2340.

1993 FACSS Meeting, Detroit, Michigan, October 17-22, 1993; Contact: H. N. Cheng, Hercules, Inc., Research Center, 500 Hercules Road, Wilmington, DE 19808; Phone: (302) 995-3505; Fax:. (302) 995-4117. See TAMU NMR Newsletter 411, 10.

Additional listings of meetings, etc., are invited.

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December 7, 1992 (received 12/11/92)

Professor Bernard Shapiro Editor/Publisher TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

"Collective motions along the long side chain on polypeptide"

Dear Professor Shapiro,

As usual it takes your reminder notice to prompt us into submission. In our continuing studies of the side-chain dynamics of polypeptides, we have been finding the collective motions along the long side chain on polypeptides in the solid state.

The main chain of poly(γ -benzyl L-glutamate)(PBLG) forms an α -helical conformation, and this rod-like molecule packs into hexagonal in the crystalline state. On the other hand, the structure of the side chain of PBLG has not been clarified yet, and the glass-like transition is observed at above the room-temperature. We have measured T_1 and line-shape of the solid state 2 H-NMR(30.7 MHz) for three PBLGs deuterated in γ , ζ , and κ positions, respectively, in the side chain. T_1 values obtained are shown in Figure 1, with the side chain structure of PBLG. The values show almost same temperature dependence for three samples, indicating same correlation times. This finding implies that same molecular motions exist in the different positions along the side chain, for the deuteron T_1 is mainly dominated by the quadrupole interaction. The Pake powder pattern is destroyed out to a singlet, due to the fast and larger-amplitude reorientations in this temperature range.

Sincerely,

Toshifumi Hiraoki

So Kitazawa

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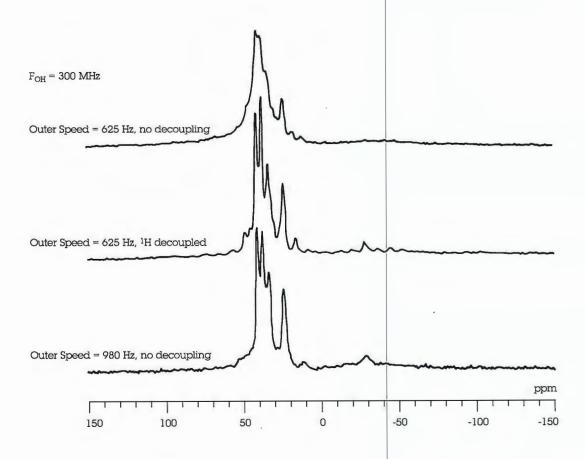
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December 8, 1992 (received 12/12/92)

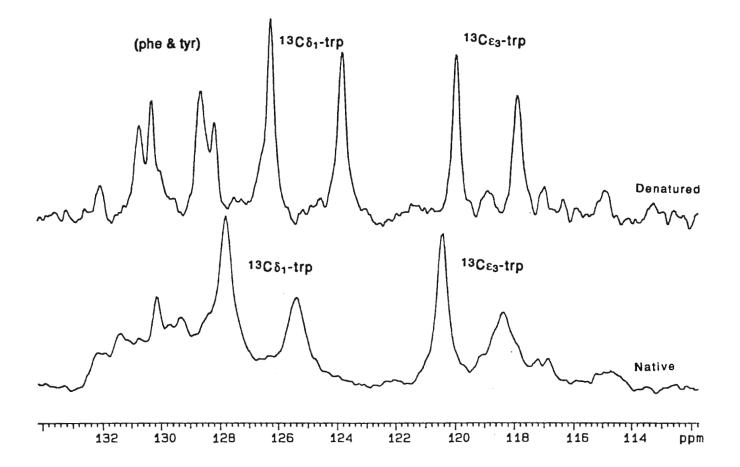
Dr. B. L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

¹³C Differential Broadening of Thioredoxin

Dear Barry,

For some time we have been using ¹³C NMR relaxation measurements to study internal motions in ¹³C-labeled peptides and proteins in liquid solution. Normally we measure T₁, T₂, and the steady state NOE of ¹³C nuclei that have attached protons. The relaxation rates are subsequently interpreted in terms of overall rotational motion and internal motion of the molecule. It is also true that useful dynamical information can be obtained from proton-coupled ¹³C spectral lines. Consider, for example, the case of a ¹³C nucleus with a single attached proton. Because of interference between the chemical shift anisotropy and the dipolar interactions, the individual components of the ¹³C doublet generally will have different widths provided the CSA of the particular ¹³C is sufficiently large. In fairly big molecules such as proteins, the more restricted the internal motion at the site of the ¹³C, the larger this so-called differential broadening will be. The requirement on the size of the CSA is readily met by aromatic ¹³C nuclei (for example in tryptophan) at typical frequencies of 75 MHz and above. Examples of differential broadening that we have observed in ¹³C-tryptophan in thioredoxin (TRX) are described below.

Escherichia coli TRX is a low molecular weight (11,700), almost spherical redox protein. The "active site" of E. coli TRX consists of cysteine residues at positions 32 and 35 while TRX has trp residues at positions 28 and 31. X-ray and NMR work have shown that trp-31 lies on the surface of the protein and that trp-28 is partially buried and spatially proximate to the pair of cysteines. We have studied two single trp variants of E. coli TRX, W28F and W31F, in which phenylalanine replaces trp-28 and trp-31 respectively by incorporating into those proteins two, different, isotopically labeled trp residues, one with ¹³C at the δ_1 (or 2) position of the indole ring and one with ¹³C at the ε_3 (or 4) position of the indole ring. From the relaxation measurements we found that the trp side chain motion is quite restricted in W28F and W31F. To see if the motion of the trp rings could be made less restricted, we performed relaxation measurements on urea denatured W28F and W31F. Indeed at 8-9 M urea, the internal motion of the trp side chains became substantially less restricted. As the urea concentration was titrated in W28F and W31F, two ¹³C NMR signals in slow exchange became apparent for each label, one from the native, folded forms and one from the denatured forms. Differential broadening measurements gave dramatic evidence of the contrast in the native and denatured forms as shown in the figure (next page) for reduced W28F. In the native protein the differential broadening is substantial for both ¹³Cδ₁ and ¹³Cε₃-trp while there was essentially no differential broadening in the denatured proteins. It is clear that measurements of differential broadening can be quite informative in the study of molecular dynamics.



Proton coupled ¹³C NMR spectrum at 75.4 MHz (referenced to internal dioxane at 67.37 ppm) of reduced W28F in 50 mM, pH 6.5 phosphate buffer (below) and in 8 M urea (above) at 20 ° C.

Please credit this contribution to the account of B. D. Nageswara Rao.

Best regards,

Marvin D. Kemple

Peng Yuan



NMR Instruments

Dr.B L Shapiro TAMU NMR Newletter 966 Elsinore Court Palo Alto CA 94303 USA



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Our Ref: DW0625

18 December 1992 (received 12/21/92)

Dear Dr Shapiro,

Progress Towards 750 MHz - Update

Since our last communication (11 November 1991) on our 750 MHz Superconducting Magnet Development Programme we have made excellent progress towards our goal of providing a 750 MHz magnet as the flagship of our product range.

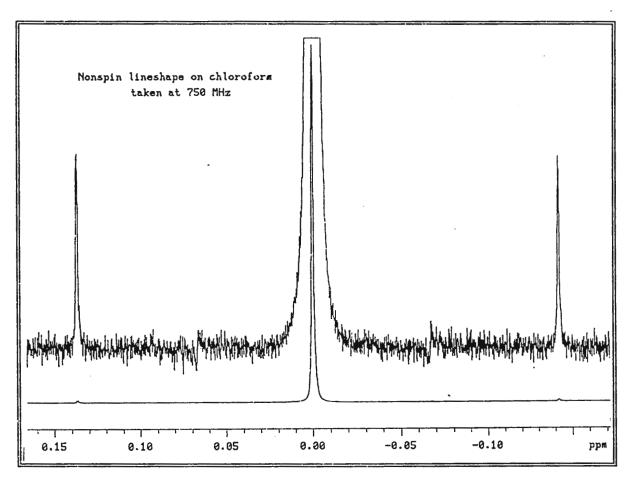
We previously reported results from Phase I of our programme which culminated in a 720 MHz prototype NMR magnet system. This system operates at a temperature of 2.2K and requires relatively complex cryogenic control. Although this was a milestone in high field NMR development we highlighted the problems associated with operating such a system as part of a routine high resolution NMR Instruments, specifically the need for highly skilled operators and the potential for NMR artefacts due to vibrational effects.

To overcome these inherent problems and offer the most user friendly system possible we have developed a 750 MHz magnet that operates at a temperature of 4.2K. Thus operating in the same convenient, well-understood way as all of Oxford's NMR magnets including the 600 MHz system.

We are pleased to announce that the prototype 750 MHz magnet has been successfully energised to field and is presently undergoing trials at Oxford's factory in the UK. Some optimisation may be required but the results obtained so far are extremely encouraging.

Although the magnet drift has not fully settled out, we have seen overnight drift rates as low as 10 proton Hz/hour and fully expect it to reduce further. The homogeneity analysis is encouraging, as can be seen from the nonspun lineshape on chloroform taken at 750 MHz shown overleaf, but some further work needs to be done to optimise the shimming. The operation of the cryostat is as simple as that of a 600 MHz cryostat, and the cryogenic performance is good, offering two months helium refill interval. The next step is to carry out some further integration and optimisation.

Continued overleaf.....



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This development could not have been carried out without the unique contributions of Bill Timms, Paul Noonan, Peter Daniels and their team. Partial financial support from the Department of Industry under the collaborative LINK scheme is acknowledged.

Yours sincerely

Dr Bill Proctor Managing Director

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Dr Neil Killoran Technical Manager

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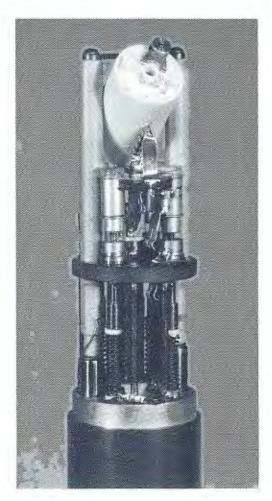
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9.4	¹³ C	100	10	250	6
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Measurement of Long-Range ¹³C - ¹H Coupling Constants in Carbohydrates

Rossitza Gitti and C. Allen Bush

Department of Chemistry and Biochemistry University of Maryland Baltimore County Baltimore, Maryland 21228

Valuable conformational information can be derived from measurments of 3-bond heteronuclear coupling constants between ¹H and ¹³C across the glycosidic linkage in oligosaccharides, (Rosvear et al., 1982). Although the measurements are technically difficult, efforts have continued to develop suitably parameterized trigonometric formulas for correlating the coupling constants with the dihedral angles of the glycosidic linkage, (Tvaroska, 1990). Among a number of new NMR experiments recently proposed for measurement of long-range heteronuclear coupling, one by Montelione et al. (1989) involves generation of 'E-COSY' type cross peak patterns in homonuclear 2-d spectra of samples enriched in ¹⁵N or ¹³C. While most of the applications reported for this method involve ¹⁵N, it should also work for ¹³C enriched carbohydrates. Although selective or partial enrichment might avoid some spectroscopic problems such as ¹³C - ¹³C coupling and ¹H line broadening by ¹³C relaxation, it should be easy to prepare uniform 100% enriched bacterial polysaccharides by growing bacteria in ¹³C enriched media. Selective or partial enrichement could be difficult for bacteria requiring multiple carbon sources.

Therefore we have applied the method of Montelione et al. (1989) to uniformly highly 13 C enriched glucose. In these experiments we expect each homonuclear crosspeak to be split into four components separated in both dimensions by the one-bond 13 C - 1 H coupling. A phase-sensitive 2-d 1 H TOCSY spectrum of uniform 98% labeled 13 C glucose was recorded on a GN-500 spectrometer with MLEV-17 mixing with trim pulses. The results in Fig. 1 show the cross peaks of the upfield satelites of H2 and H4 of the β -anomer with H1. Fig. 2 shows cross peaks of H1 of the α -anomer with H2 and H3. Values of the coupling constants can be estimated from the contour plots made with high digital resolution. Using Hare's FELIX program to 'zoom' sub-blocks of our spectrum to 0.3 Hz/pt resolution, we have estimated J values to approximately \pm 0.4 Hz. Measurements are in reasonable agreement with literature values for the β -anomer of 5.5 Hz for 2 J_{C1-H2} (Walker et al, 1976) and for the α -anomer of 5.3 Hz and 1.3 Hz for 3 J_{C3-H1} and 2 J_{C2-H1} respectively (Gidley and Bociek, 1985).

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- P. R. Rosvear, H. A. Nunez and R. Barker (1982) Biochemistry, 21, 1421-1431.
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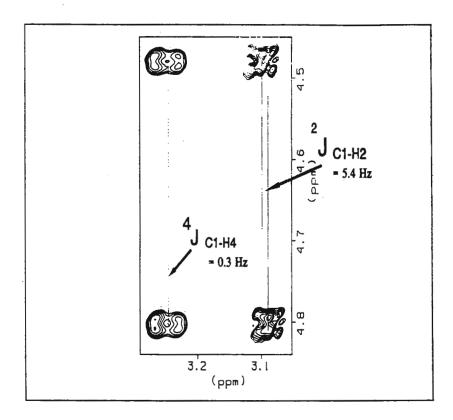


Figure 1. TOCSY cross peaks of β -H1 with H2 and H4 yield estimates of ${}^2J_{C1-H2}$ and of ${}^4J_{C1-H4}$.

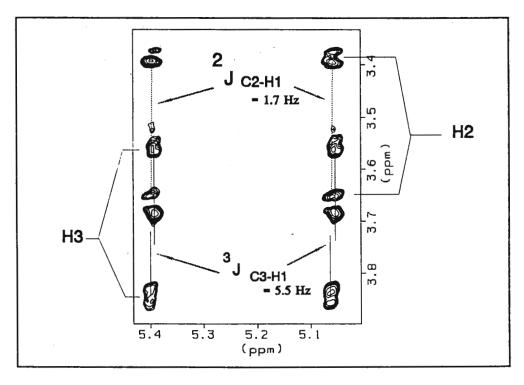


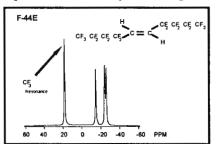
Figure 2. TOCSY cross peaks of α -H2 and H3 with H1 yield estimates of ${}^2\mathrm{J}_{C2-H1}$ and ${}^3\mathrm{J}_{C3-H1}$



Professor Bernard L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

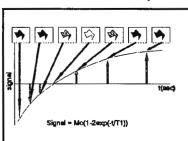
Dr. Shapiro:

Measuring tissue oxygenation non-invasively *in-vivo* is a daunting task. However, it is well known that the spin-lattice relaxation rate of ¹⁹F in perfluorocarbon (PFC) emulsions is linearly related to dissolved oxygen content. ¹ Spatial mapping of oxygen tension *in-vivo*, using sequestered PFC emulsions, has also been published. ² Producing pO₂ maps can take an extraordinary amount of time because of long spin-lattice time constants of ¹⁹F PFC emulsions at normal oxygen tensions. We propose a technique to map oxygen tension *in-vivo* by employing ¹⁹F echo planar inversion recovery imaging of sequestered PFC emulsions. An inversion recovery method is more accurate than a progressive saturation technique because it provides twice the dynamic range. Since echo planar



imaging also produces an entire image in less than 100 milliseconds, more images can be obtained or averaged at different inversion times making spin echo techniques time prohibitive.

Female, C3H mice are administered a 10g/kg dose of F-44E PFC emulsion (see spectrum in Figure 1) by tail vein injection. In three to five days most of the PFC emulsion is sequestered in the recticuloendothelial system of the liver and spleen. At



this time the animals are anesthetized with 1.5% isoflurane in air or pure oxygen. Measurements are performed on a 2.0T GE-CSI-II imaging spectrometer with 20g/cm self-shielded gradients. The

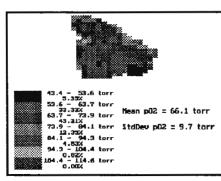
transmitter frequency is selectively set on the CF₃ resonance and a 64x64 ¹⁹F projection echo planar image, instead of a FID, is acquired at each inversion recovery time (Figure 2). The following TI intervals were used: (0.08,0.2,.5,1,2,4,8) seconds at a TR of 10 seconds. At a spectral width of 70KHz, the acquisition time was 28.6msec. With NA=2, the total acquisition time was approximately 2.5 minutes.

Biomedical Engineering Department

100 Institute Road Worcester, MA 01609-2280 (508) 831-5447 FAX (508) 831-5483

(received 11/30/92) November 18, 1992

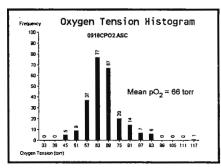
Oxygen Tension Mapping with ¹⁹F Echo Planar Inversion Recovery Imaging



The oxygen tension maps are then produced by calculating the relaxation rate, R₁, on a pixel by pixel basis, using a nonlinear least squares algorithm. Each pixel is then assigned a pO₂

value, in torr, from calibrations obtained from phantom experiments. A pO₂ map of the liver and spleen of a mouse breathing 21% O₂ is shown in Figure 3. The corresponding histogram with bin widths of 6 torr is shown in Figure 4. Oxygen tension maps were also produced for the same animal breathing 100% O₂ and post-mortem with mean pO₂'s of 115 and 8.7 torr, respectively. We hope to extend this technique to spatially map tumor oxygenation in-vivo to study the effects

of adjuvants on radiotherapy.



Parhami and B.M. Fung, J. Phys. Chem., 87, 1928(1983).
 S.K. Holland, R. Soto, C.C. Jaffe, E. Gordon, and J.C. Gore, Magn. Reson. Imaging, 6, (Supplement 1), 72(1988).

Bernard J. Dardzinski

Christopher H. Sotak



Oklahoma State University

STILLWATER, OKLAHOMA 74078-0447 PHYSICAL SCIENCES 106 TEL: (405) 744-5920 FAX: (405) 744-6007

DEPARTMENT OF CHEMISTRY COLLEGE OF ARTS AND SCIENCES

Dr. B. L. Shapiro 966 Elsinore Court Palo Alto, CALIFORNIA 94303 11/24/92 (received 11/27/92)

SUBJECT: Novel Heteroarotinoids

Dear Dr. Shapiro:

Our contribution to the Newsletter is concerned with some heteroarotinoids (peripherally related to certain natural retinoids). The examples prepared recently were interesting in that at 300 MHz most of the signals were visible. The proton signals are followed by the signals for C-13 in ppm (all from TMS). What

Ar-H (Ar-C and vinyl-C): δ 6.80-8.03 (116-153 ppm) C=O: 166.7 ppm

Ar-H (Ar-C and vinyl-C): δ 7.20-8.04 (123.7-141.7 ppm)

C=O: 166.6 ppm

is interesting is that the NMR signals for the oxygen compound (top) in DCCl₃ do not appear to correlate well with what is present in the solid state analysis. The two aryl rings are more twisted (they are tilted approximately 45° with respect to each other) in the solid state than anticipated from the NMR analysis. We are looking at several other examples in this novel family.

Sincerely yours,

K. Darrell Berlin Regents Pofessor

Water Suppression Inverse 'H/"N Triple - resonance NMR Pulse Shaping 3D NMR Gradient - DQF COSY

CP - MAS



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ARX		300	400	500	Ail
Probehead	Sample	_			
Resolution Test					
All 5 mm ¹ H 5+10 mm ¹³ C	ODCB/C ₆ H ₆	0.2	0.2	0.2	
Lineshape Test					SSB %
All 5 mm ¹ H	10% CHCl ₃	6/12	7/15	7/15	<1
All 5 (10) mm ¹³ C	80% C ₆ H ₆	3/7 (3/7)	3/7 (3/8)	4/8 (4/8)	< 0.5 (< 1)
Sensitivity Test					
5 mm ¹ H Selective	0.1% EB	175	250	450	<10
5 mm ¹ H Inverse Detection	0.1% EB	135	190	350	<15
5 mm ¹ H Dual, QNP, VSP	0.1% EB	100	140	200	<15
5/10 mm ¹³ C QNP, Dual	ASTM	100/320	160/450	180/600	<15/<20
5/10 mm ¹³ C QNP, Dual	10% EB	70/200	100/300	150/400	
5/10 mm ¹³ C VSP multinuc.	ASTM	100/320	160/450	180/600	<15/<20
5/10 mm ¹³ C VSP multinuc.	10% EB	70/200	100/260	150/320	
5/10 mm ¹⁵ N VSP multinuc.	90% Form.	10/35	15/55	20/70	<25/<30

EB = ethylbenzene (for ¹³C with ¹H-dec.); ASTM = $60\% C_6D_6$ in dioxane Form. = formamide ('IH-dec. without NOE) lineshape: 1H = CHCl₃ linewidth at ht. of 1 °C-satellites/at 20° this level 1 °C = 1 SSB = Spinning sidebands measured with 8 transients QNP: 5 or 10 mm ¹H, ³¹P, ¹³C, ¹⁵N VSP: 5 mm¹⁵N - ¹³P; 10 mm ¹⁰⁹Ag - ³¹P.

Console	Dimensions and weights are approximate; voltage +10/ - 5% max. variation; other line freq. and voltage upon request
dimensions electronics cabinet	w 130 x d 0.75 x h 1.20 m
weight	450 kg
power (dissipation)	220 V/50 Hz/16 A (ca. 4 kW)



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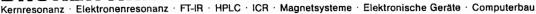
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Dr. B.L. Shapiro TAMU NMR NEWSLETTER 966 Elsinore Court Palo Alto, CA 94303, USA

December 14, 1992 (received 12/17/92)

First High Resolution Spectrum at 750 MHz

Dear Barry:

BRUKER is proud to present the first high resolution spectrum at 750 MHz. This spectrum was obtained within a few days after energizing our AMX 750 in its final dewar. We first successfully energized a 750 MHz persistent superconducting magnet, which BRUKER built in cooperation with the Kernforschungszentrum Karlsruhe, more than one year ago, and reported it in the TAMU NEWSLETTER in October 1991. We expected then, that the development of a complete 750 spectrometer would take several years, but progress has been more rapid.

The first 750 MHz spectrum shown in Fig. 1 was obtained from a cyclic tripeptide (Cyclo [L - Pro - L - Pro - L - Thz]), using a 5 mm proton selective probe (with lock) and BOSS[™] shims on an AMX 750 with SE-451[™] receiver. For comparison, the same spectrum is also shown using an AMX 600 spectrometer. Besides the 25 % higher dispersion, which is clearly evident, the absolute resolution of the 750 MHz spectrum is essentially the same as at 600 MHz. Due to the higher dispersion, the two spin systems which at 600 MHz are still partially overlapping, are now well resoved at 750 MHz.

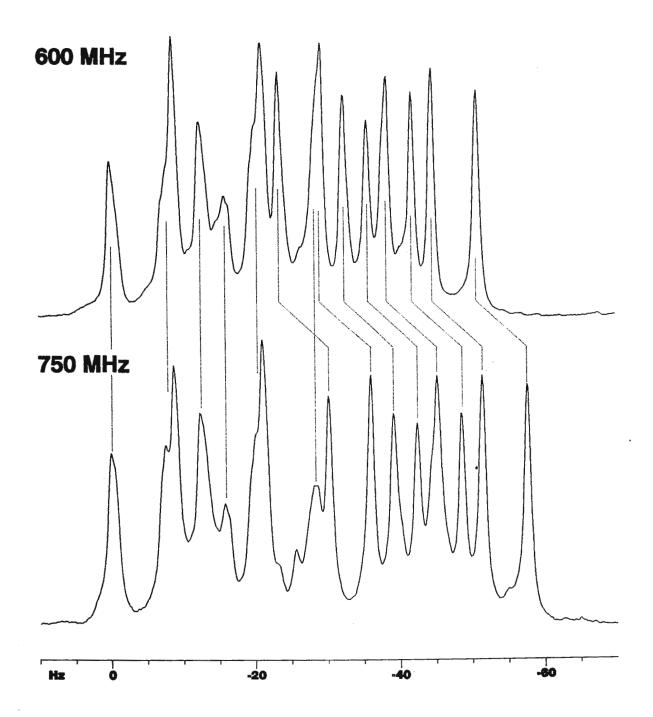
We hope that this first spectrum gives a good indication of the future potential of this new class of high-resolution, solid-state and microimaging NMR spectrometers. As progress continues, we will be happy to keep you informed about the latest developments with this new research spectrometer.

Sincerely,

Prof. Dr. Günther Laukien

pr. Gerhard Roth

Fig.1: 600 and 750 MHz spectra of cyclo[L-Pro-L-Pro-L-Thz] in CDCl₃. The expansion shows the signals of the cis-δ proton of proline 1 and 2. Both spectra are plotted onthe same Hz/cm scale.



We acknowledge Dipl. Phys. R. Ruin for quickly shimming the magnet and Dr. W. Bermel for recording this first spectrum.

UMEÅ UNIVERSITET Avdelningen för organisk kemi

Telefon 090-16 50 00



UNIVERSITY OF UMEÅ

Department of Organic Chemistry

Telephone + 46 + 90 + 16 50 00

Dr B. L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, California 94303

Dec 1 1992 (received 12/10/92)

Dear Barry,

OCC and Asymmetry Parameters of Phenyllithiums Using Solid State ⁷Li NMR.

For many organolithium compounds it is possible to obtain a detailed information of the state of aggregation in solution by measuring lithium relaxation, provided that there exists a reliable probe for the effective rotational correlation time of the studied complex. However, this latter requirement is quite critical and the obtained quadrupolar splitting constants (QSC) may be far from reliable.

To circumwent this problem and to compare the solution and solid state data, we have studied microcrystalline samples of variously complexed phenylithiums using static and MAS ⁷Li NMR. A representative experimental and simulated powder spectrum is shown below for the PMDTA complex yielding a QCC of 260 kHz as expected for a monomer. For the TMEDA adduct, being a dimer according to X-ray, we obtained a QCC of 147 kHz, which affords a QSC of 161 kHz as compared to the solution value of 162 kHz. Similar agreement between solid and solution data was noted for tetrameric complexes. So solid state studies can be quite useful to characterize reactive organolithium species and to connect X-ray and solution experiments. However, keep your polymeric rotors away from these species. Organolithium compounds eat them.

160 80 0 -80 -160 kHz

All solid state studies so far have been performed on our old Bruker MSL-100 instrument. We will soon have solid facilities on our 500 MHz system also together with an upgrade from AM to a rack version of the new "X32-free" AMX.

Best regards

Ul Edlund

Dan Johnels



Universität Basel

Institut für Physik Kondensierte Materie

Institut für Physik Klingelbergstrasse 82 CH-4056 Basel

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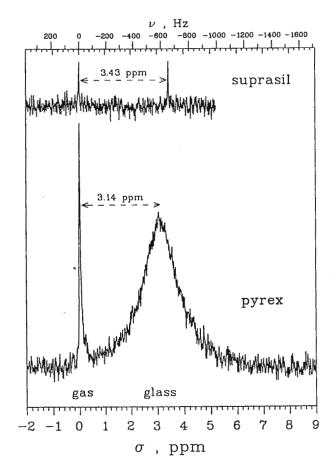
Prof. B.L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303 USA

December 14, 1992 (received 12/18/92)

NMR of ³He in vitreous silica

Dear Barry,

We have lately been interested in measuring solvent effects on the chemical shift of ³He in liquids. During these studies we observed that ³He can also be detected if it has penetrated into the glass of the sample wall (see the Fig.). So far we have been able to obtain signals of ³He in quartz (suprasil) (one FID) and in pyrex (several thousand FID's). In suprasil the diffusion is rather fast (half time of ≈2 days); in pyrex it is slower. We have studied the ³He relaxation and have been able to attribute it to the dipole-dipole interaction in pyrex with Fe3+ impurity ions and in suprasil with unpaired electrons of defects. Presently we are studying ³He in various solids.



With best regards

Prof. P.Diehl,

Dr. R. Mazitov,

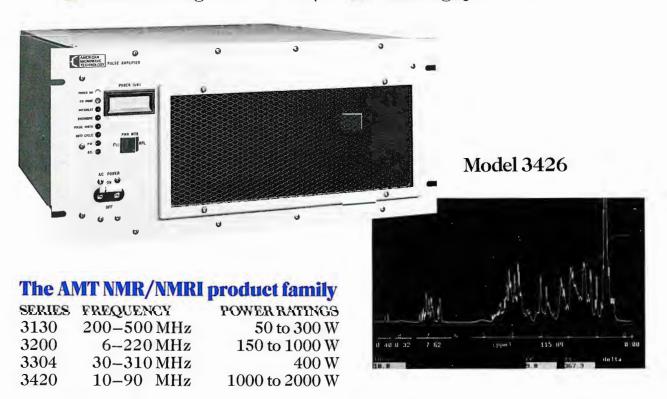
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Amplitude droop

Phase change/output power

Phase error overpulse

Noise figure

Output noise (blanked)

Blanking delay

Protection

 $10 - 90 \, \text{MHz}$ $1000 \, \mathrm{W}$ $100 \, \mathrm{W}$ $0 - 900 \, W$

65 dB

 $\pm 2 dB$ 50 ohms

< 2:1 $20\,\mathrm{ms}$

Up to 10% 250 ns typ.

5% to 20 ms typ.

10° to rated power, typ.

4° to 20 ms duration, typ.

11 dB typ.

< 20 dB over thermal < 2 µs on/off, TTL signal

1. VSWR: infinite VSWR

2. Input overdrive: up to 10 dB

3. Over duty cycle/pulse width

4. Over temperature

Supplemental characteristics:

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1. RF input: BNC (F)

2. RF output: Type N (F) 3. Noise blanking: BNC (F)

4. Interface: 25 pin D(F), EMI filtered

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1. AC power on

5. Over temperature

2. Peak power meter

6. Over drive

3. Over pulse width

7. CW mode

4. Over duty cycle

System monitors

1. Forward/Reflected RF power

2. Over pulse width/duty cycle

3. DC power supply fault

4. Thermal fault

Front panel controls

1. AC power

3. Duty cycle

2. Pulse width

Cooling

Operating temperature

AC line voltage

AC power requirements

Package

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Professor B.L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303 CB# 3290, Venable and Kenan Laboratories The University of North Carolina at Chapel Hill Chapel Hill, N.C. 27599-3290

(received 12/15/92)

"Squeezing more out of FT-PGSE self-diffusion experiments - blood from a turnip (?)"

Dear Barry,

Thank you for the final ultimatum to TAMU NMR Newsletter subscriber PS (normal address RIT, Stockholm, but presently having a short sabbatical in the laboratory of Charles S. Johnson Jr., at UNC). We have done some independent work along the lines of improved data evaluation in the case of FT-based pulsed-gradient NMR spin-echo experiments for the determination of self-diffusion coefficients on multi-component or polydisperse samples in solution. Self-diffusion coefficients contain a wealth of highly relevant and easily conceived physico-chemical information, especially on colloidal or macromolecular systems in solution, and NMR is undoubtedly the method of choice for accessing this type of information, as compared to radioactive tracer methods, or optical techniques.

The traditional approach of measuring NMR spin-echo intensities or peak integrals for subsequent fitting to the Stejskal-Tanner equation is normally sufficient for the majority of needs. However, if there is extensive peak overlap between components, or if the sample contains some polydisperse polymer, one must turn to more advanced data processing. With present-day instrumentation, it is natural then to try to use the complete experimental information available in bandshapes already stored on the spectrometer disk, and to utilize the really enormous computing power available on the on-line or network-connected workstations typically available.

Details of full FT-PGSE spectral data processing through deconvoluting routines like CONTIN, SPLMOD or DISCRETE (see e.g S.W. Provencher, Comput. Phys. Commun., 27 (1982) 213 and 229), adding some extra "decision rules" for the deconvoluting process was recently submitted for publication (Morris and Johnson, see also previous communications in JACS 114 (1992) 3139 and 776). Figure 1 illustrates the power of that approach, and the information content that can be achieved. The 2D variant of the techniques is named DOSY (Diffusion Ordered NMR Spectroscopy). Together with former Stockholm postdoc Dirk Schulze (now at Shell Laboratories, Amsterdam), PS developed a complementary approach, that enables one to directly deconvolute spectral bandhapes, contributing to severely overlapping bands in FT-PGSE data sets, even in the case of very similar self-diffusion coefficients (Figure 2; the paper will be submitted when this appears in print).

We believe that procedures like these will be of significant value in future FT-PGSE work on more complex systems than previously considered feasible. Please credit this contribution to Peter Stilbs subscription.

Best regards,

Kevin Morris

Charles S. Johnson, Jr

Charle

Sincerely Yours

Peter Stilbs

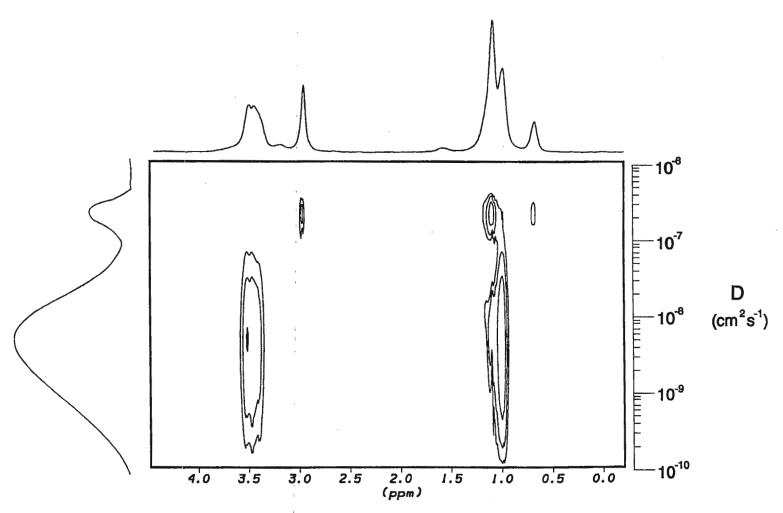
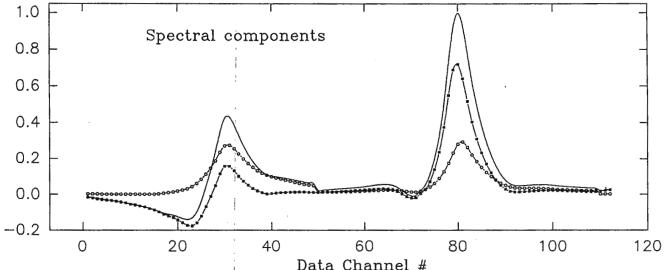


Figure 1: A 250 MHz Proton DOSY spectrum of 1% Ethyl(hydroxyethyl)cellulose (EHEC) and 0.34% cetyl trimethylammonium bromide (CTAB) in heavy water. Such systems exhibit highly unusual rheologic behaviour. Note the very wide (polydisperse) EHEC diffusion distribution, and the very much narrower one for CTAB.



Data Channel #
Figure 2: A deconvolution of the highly overlapping FT-PGSE spectra of a mixture of n-octane (open circles) and n-heptanol (filled squares); as based on differential signal attenuation effects on the combined bandshape (solid line) during FT-PGSE experiments.



CENTRO DE INVESTIGACION Y DE ESTUDIOS AVANZADOS DEL I.P.N.

APARTADO POSTAL 14-740 MEXICO 07000, D.F.

November 6, 1992. (received 11/21/92)

Dr. Bernard L. Shapiro Editor/Publisher TAMU NMR Newsletter 966 Elsinore Ct. Palo Alto, CA 94303 USA

¹H NMR Studies of E and Z 2-hydroxyindolenines

Dear Professor Shapiro:

The proton signals of 2-hydroxyindolenines $\underline{1}$ - $\underline{4}$ and of the 2-methoxy derivative $\underline{5}$ were assigned after first-order analysis of the 300 MHz spectra measured in CDC $\overline{1}_3$ and DMSO-d₆ solutions from mixtures of E and Z isomers. The data are listed in the Table.

In the chloroform spectra, the proton signals from position 2 owing to the E and Z isomers (E/Z ratio \underline{ca} 1:10) appear as broad doublets resulting from the interaction with the 2-OH group. For the Z isomer the H-2 signal appears at lower fields (0.41-0.47 ppm), compared with that of the E isomer, as a result of a selective anisotropic effect exerted by the carbonyl moiety of the ester group at C-8. Accordingly, the relative shieldings of the H-4 signals in the E,Z pairs vary in opposite directions, with H-4 in the Z isomer resonating at higher fields (0.30-0.41 ppm) than the E isomer. In the later form, H-4 is coplanar with the 8-carbonyl moiety^{1,2}. Consequently the chemical shifts owing to H-2 and H-4 can be used as key arguments for E,Z assignments. Although all other aromatic protons are less affected by the configuration of the C/3 = C/8 double bond, they were easily discerned from the proton-proton coupling patterns.

On changing the solvent from CDCl $_3$ to DMSO-d $_6$ some chemical shift variations were observed. As expected, the most severe change occurs in the 2-OH chemical shift, which was considerably more shielded in CDCl $_3$ (by \underline{ca} 3 ppm) than in DMSO-d $_6$.

Contrarly to observations for the related 2-oxoindolenines 3 , the E/Z ratio for $\frac{1}{2}$ - $\frac{5}{2}$ were found to be solvent independent.

- 1.- G. Jones and W.J. Rae, <u>Tetrahedron</u>, <u>22</u>, 3021 (1966).
- 2.- R.L. Autrey and F.C. Tahk, <u>Tetrahedron</u>, <u>23</u>, 901 (1967).
- 3.- M.S. Morales-Ríos, Y. Mora-Pérez and P. Joseph-Nathan, Magn. Reson. Chem., in press.

			Χ	Υ	R^1	R^2	R^3
		Ε	CO ₂ Me	CN	CO ₂ Me	Н	Н
	1	Z	CN	CO ₂ Me	CO ₂ Me	Н	Н
	12	Е	CO ₂ Et	CN	CO ₂ Et	Н	Н
\mathbb{R}^2	2	Z	CN	CO ₂ Et	CO ₂ Et	Н	Н
3 3 8 Y	3	Ε	CO ₂ Me	CN	CO ₂ Me	$N0_2$	Н
$^{\circ}$. <u>3</u>	Z	CN	CO ₂ Me	CO ₂ Me	$N0_2$	Н
R ₁	.4	Ε	CO ₂ Et	CN	CO ₂ Et	$N0_2$	Н
		Z	CN	CO ₂ Et	CO ₂ Et	NO_2	Н
		Ε	CO ₂ Me	CN	CO ₂ Me	Н	Me
	<u>5</u>	Z	CN	CO ₂ Me	CO ₂ Me	Н	Me

Table.- Chemical shifts in ${\rm CDCl}_3$ (and ${\rm DMSO-d}_6$).

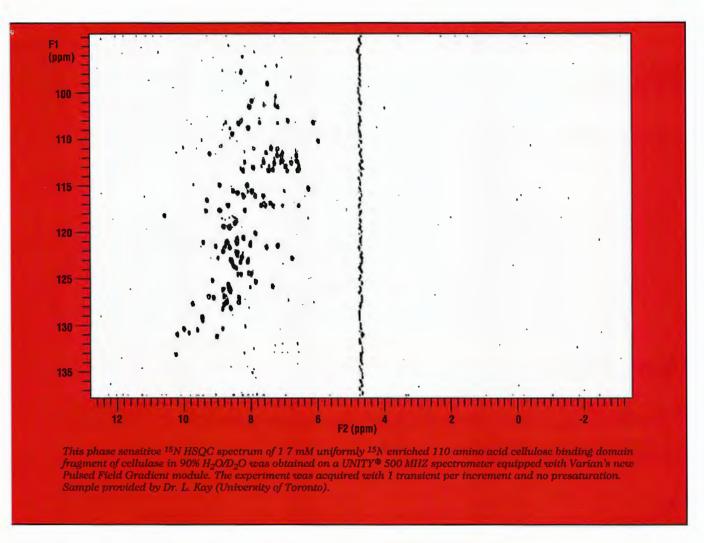
				•	•	
	Н2	H4	Н5	Н6	Н7	ОН
<u>1</u>	E 6.33(6.16)	8.76(8.57)	7.11(7.24)	7.53(7.65)	7.98(7.88)	4.65(7.69)
	Z 6.80(6.71)	8.46(8.29)	7.18(7.27)	7.60(7.69)	7.98(7.88)	4,50(7.40)
2	E 6.39(6.13)	8.75(8.57)	7.12(7.15)	7.52(7.63)	7.95(7.88)	4.68(7.66)
	Z 6.80(6.69)	8.46(8.27)	7.16(7.23)	7.59(7.66)	7.95(7.88)	4.52(7.32)
3	E 6.50(6.25)	9.76(9.56)	-	8.44(8.49)	8.13(8.01)	4.74(8.05)
	Z 6.92(6.76)	9.33(9.05)	-	8.48(8.52)	8.13(8.01)	4.48(7.77)
<u>4</u>	E 6.51(6.24)	9.74(9.51)	-	8.43(8.46)	8.12(8.00)	4.69(8.03)
	Z 6.92(6.72)	9.33(9.03)		8.47(8.50)	8.12(7.98)	4.54(7.70)
_	E 6.19(6.15)	8.72(8.57)	7.12(7.20)	7.55(7.66)	7.94(7.86)	, -
<u>5</u>	Z 6.88(6.66)	8.46(8.27)	7.18(7.28)	7.58(7.70)	7.94(7.86)	-

Sincerely yours,

Martha S. Morales-Ríos

Pedro Jøseph-Nathan

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December 16, 1992 (received 12/18/92)

Varian Obtains NMR Results at 750 MHz

Dr. B. L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303

Dear Barry,

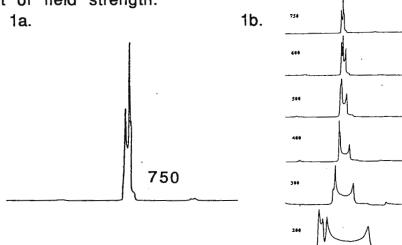
We are currently acquiring data on our new Varian UNITYplus™ NMR console connected to a prototype Oxford 750 MHz superconducting magnet! We believe these are the first reported high resolution NMR results at 750 MHz for chemical and biomolecular applications which further demonstrates Varian's continuing commitment to high field NMR.

Varian has been developing probes and spectrometer consoles at 750 MHz in anticipation of the Oxford 750 MHz magnet. The spectra below demonstrate that today's 750 MHz NMR systems provide excellent results in a variety of experiments with the resolution and sensitivity enhancements that are expected.

Results

1.a. The ²³Na MAS spectrum of Na₂MoO₄ continues to narrow as the data is acquired at higher field strengths.

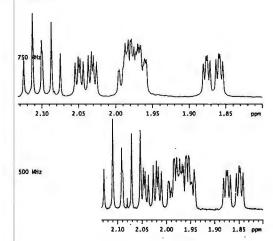
b. The series of spectra plotted from 200 to 750 MHz shows the effect of field strength.



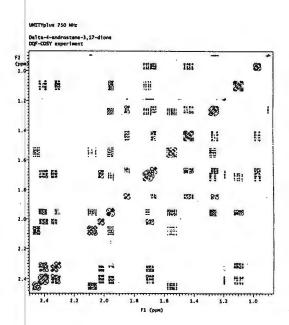


- 2.a. The second result is a comparison of resolution on a 750 MHz spectrometer with a 500 MHz spectrometer. The sample is delta-4-androstene-3,17-dione. The gain in resolution due to chemical shift dispersion is quite significant.
- b. The DQCOSY on the same sample demonstrates that the magnet and spectrometer technology available today is capable of providing long term two dimensional results.

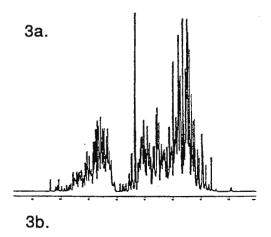
2a.

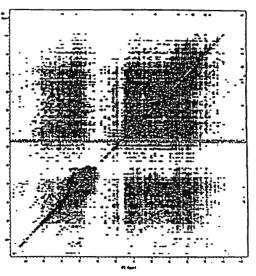


2b.



- The third result is a water presaturation experiment on 5mM Lysozyme in 90% H₂O/D₂O. The result shows that excellent lineshape can be obtained on the 750 MHz system and biomolecular studies are feasible today. The peaks next to the residual water in the 1D spectrum are not suppressed.
- b. The NOESY on 5mM Lysozyme in 90% H₂O/D₂O has a narrow water stripe with resolution and sensitivity superior to a 600 MHz spectrum.





We are extremely excited about these results and are looking foreward to working with scientists worldwide on additional applications at 750 MHz.

Sincerely,

Tinothe Davien Timothy Saarinen

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Mark van Criekinge George Gray-

R&D Engineer

Peter Sandor

PETER

Applications Chemist

Stracke

Applications Chemist

Ali Behbin & Behbon
Probe Engineer

Ron Haner Probe Engineer





Extending spectrometer technology beyond current limits is a major goal at Varian NMR Instruments. Today, our course of innovation continues to provide professionals with the resources to step outside traditional boundaries. By encouraging professionalism that dares to be different, Varian remains first and foremost with opportunities that keep you on the industry's cutting edge.

Senior Technical Staff Member

Reporting to the Manager of Research and Development, you will be responsible for extending the application boundaries of high-resolution NMR by designing novel experiments to perform with existing instrumentation, and by defining the operating parameters of new equipment. The successful candidate should also be capable of leading an engineering design team in the development of new NMR products.

This position requires an advanced degree in Applied Physics or Chemistry and 5 years' experience in biomolecular NMR techniques and applications. A thorough understanding of present day high-resolution NMR methods is also required. Familiarity with modern NMR instrumentation is desirable and good communication skills are essential.

Senior Applications Chemist

You will be responsible for conducting demonstrations of NMR equipment, running customer samples, presenting workshops, and evaluating new equipment. You'll also write manuals, assist in product development, participate in research programs, and provide post-sales customer support and training.

We require a PhD in Chemistry or Physics, or equivalent, and 3-5 years' experience with state-of-theart biomolecular NMR. You'll also need a background developing and/or implementing NMR pulse sequences, an understanding of NMR phenomenon sufficient to implement and test new experiments, and familiarity with UNIX*. Excellent communication and presentation skills essential. NMR hardware design or troubleshooting experience desirable.

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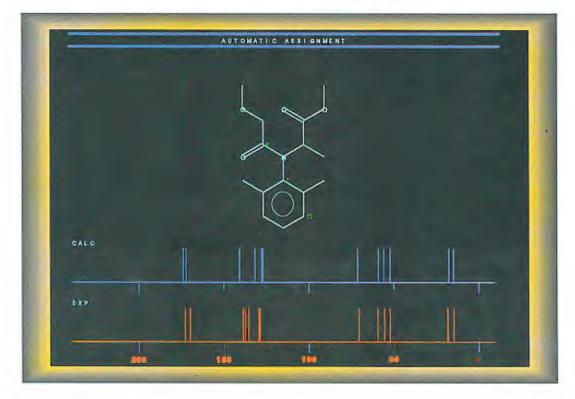
Structure Elucidation

CSEARCH

Offers identical presentation on Sun, Silicon Graphics, or VMS Reads ¹⁵C NMR peak tables directly Confirms calculated with measured spectrum Finds structurally related compounds by spectral matching Calculates spectra for compounds not in the database

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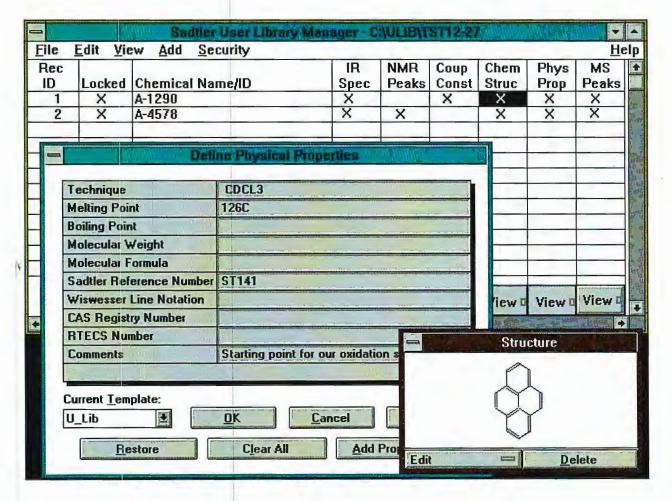
Comparison of the calculated spectrum of a proposed structure with its measured spectrum. The intelligence of CSEARCH is demonstrated by favorable comparison for a structure not contained within the database of 52,000 spectra.





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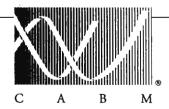
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November 23,1992 (received 11/28/92)

Triple Resonance Experiments for Determining Resonance Assignments in Small Proteins

Dear Dr. Shapiro,

During the last several months we have developed an improved strategy for determining sequence-specific NMR assignments in small isotope-enriched proteins. In our approach, the ¹H-¹H planes from a set of ¹⁵N- resolved 3D-NMR experiments are analyzed to provide both spin system identification and sequential connectivity information. These pulse sequences are extensions of the basic HCNH triple resonance^{1,2} experiment, modified to provide HCCH TOCSY transfer from sidechain protons, through sidechain and backbone carbon nuclei, to backbone amide nitrogen and protons. Multidimensional H_C_CA_N_H TOCSY³ experiments are first used to connect together the resonances of each amino acid spin system and to provide unique identification of many spin system types, including those of asparagine and glutamine which exhibit cross peaks between sidechain aliphatic and sidechain amide nuclei. Next, sequential connections between amino acid spin systems are established using multidimensional H_C_CO_N_H TOCSY4. The H_C_CA_N_H TOCSY3, or "CA-TOCSY", spectra provide an extensive set of intraresidue connections between proton, carbon, and nitrogen resonances of each amino acid in the sequence, while the complementary H_C_CO_N_H TOCSY⁴, or "CO-TOCSY", spectra provide sequential connections between aliphatic α , β , γ , and δ proton and carbon resonances of residue i and the amide proton and nitrogen resonances of residue i+1. These experiments are related to CBCANH and CBCA(CO)NH triple experiments recently described by Grzesiek & Bax^{5,6}.

Representative data from triple-resonance TOCSY spectra are shown in Figs. 1 and 2. Together, these multidimensional CA- and CO-TOCSY experiments provide the information needed to determine most of the sequence-specific backbone and sidechain resonance assignments in small proteins. Details of these pulse sequences and their application in determining sequence specific resonance assignments in proteins will be presented elsewhere^{3,4}.

bare Se Vigo Mitsuru Tashiro

Gaetano T. Montelione

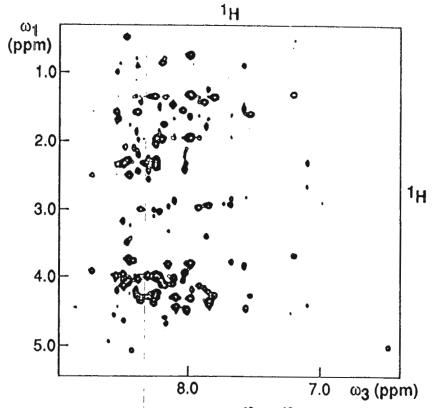


Fig. 1. 2D CO-TOCSY spectrum of a 100% uniformly ¹³C- and ¹⁵N-enriched sample of a modified 8.2 kDa domain (called 2-Domain²) of the immunoglobulin-binding Protein A from <u>Staphylococcus</u> aureus at protein concentration of 2 mM. These data were obtained at pH 6.5 and temperature of 30°C. in 24 hours using a Varian Unity 500 NMR spectrometer. Detailed analysis of the spectrum reveals that most of the cross peaks are sequential correlations between the sidechain aliphatic protons of residue i and backbone amide protons of residue i+1. In addition, for asparagine and glutamine residues intraresidue cross peaks are observed between aliphatic and sidechain amide protons. Connections between sidechain aliphatic and amide protons of asparagine and glutamine are also observed in CA-TOCSY spectra.

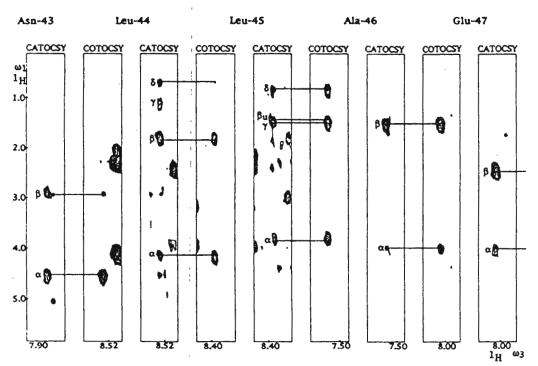


Fig. 2 Illustration of our strategy for correlating intraresidue cross peaks of residue i from the 30 CA-TOCSY spectrum with the corresponding sequential cross peaks at the amide frequency of i+1 in the 30 CO-TOCSY spectrum. This panel shows $\omega 2 = ^{15}N$ strips from 3D data sets establishing connections within the polypeptide segment Asn-43 to Glu-47 of the Z-Domain of Protein A⁷. Sample conditions are the same as in Fig. 1. Although this Z-Domain is highly ω -helical and exhibits highly degenerate ^{1}H and ^{13}C spectra, the 2D and 3D CA- and CO-TOCSY experiments have enabled us to determine an extensive set of sequence-specific ^{1}H , ^{15}N and ^{13}C resonance assignments without recourse to NOESY.

⁽¹⁾ Montelione & Wagner, 1989 J. Am. Chem. Soc. 111, 5474. (2) Montelione & Wagner, 1990 J. Magn. Reson. 87, 183. (3) Lyons & Montelione, submitted. (4) Montelione, et al. J. Am. Chem. Soc. (in press). (5) Grzesiek & Bax, 1992 J. Am. Chem. Soc. 114, 6291. (6) Grzesiek & Bax, 1992 J. Magn. Reson. 99, 201. (7) Nilsson et al., 1987 Protein Engineering 1, 107.



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Professor Barry Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303 U.S.A.

(received 12/2/92) 26 November 1992

Accurate Multiexponential Analysis of Relaxation Decays Based on Linear Prediction and Singular Value Decomposition

Dear Prof. Shapiro,

An important element in relaxation-time analysis is the fitting of a decay curve to the sum of a series of exponential functions. An examination of past literature clearly reveals that such multi-exponential decays function is particularly difficult to analyse using nonlinear least squares fitting. The difficulty lies in the fact that a decay of this type is particularly insensitive to changes in the parameters defining the exponential components, or even to change in the number of components assumed to represent the decay. Besides, in such a fitting process, the experimental error has a strong influence on the accuracy of the determined amplitudes and damping factors (inverse time constants). Thus conventional nonlinear least squares analysis is sensitive to noise and gives poor estimates when the signal-to-noise ratio is low. Hence, a very accurate analysis of multiexponential decays requires exceedingly, and sometimes impossibly, accurate data extending over a very large part of the decay curve.

Recently, we proposed a novel method based on linear prediction and singular value decomposition for the parameters of superimposed exponentials corrupted by random noise. By invoking the principle of linear prediction the fitting can be carried out by a linear least squares procedure, and therefore needs no subjective initial guess. The least squares procedure is based on singular value decomposition, which enables one to distinguish between signal and noise subspaces. This method yields a list comprising the damping factor and amplitude of each distinct exponential decay. The mathematical method used in this work was originally proposed by Kumaresan and Tufts (KT)² for estimating the parameters of damped exponential sinusoids in additive white noise. While KT's method performs well for spectrum analysis applications, it is shown here that it provides poor estimates of damping factors when directly applied to the multiexponential decay cases without any modification. This failure is because the bias in the damping factor and amplitude become significant at low signal-to-noise ratio. This type of bias is common in linear prediction methods and is due to the noise variance contributing significantly to the signal subspace. To overcome this problem, we present an empirical formula to substract the noise contamination from the signal singular values. In addition to the normal effective singular values, we have also found it useful for further enhancing the accuracy to include the two sequential smaller singular values and their corresponding singular vectors such that the smaller signal components remain present in the signal subspace.

Intensive Monte Carlo simulations for Kumaresan and Tufts's two-exponential test sample² and Clayden and Hesler's three-exponential test sample¹ are used to exhibit the improved robustness of this method compared to conventional nonlinear least squares analysis. A typical result of Kumaresan and Tufts's two-exponential test sample compared to nonlinear least square analysis is shown in Fig. A and B respectively with the true values of (damping factor and amplitude) located at (0.1,1) and

(0.2,1). Experimental data of ⁷Li longitudinal and transverse relaxations are also used which contain only 14 data points. Since ⁷Li is an I=3/2 quadrupolar nucleus, the relaxations exhibit two-exponential behavior when the extreme narrowing condition is not satisfied. The longitudinal relaxation has the function form $\left(\frac{1}{5}e^{-r_1t} + \frac{4}{5}e^{-r_2t}\right)$, and the transverse $\left(\frac{3}{5}e^{-s_1t} + \frac{2}{5}e^{-s_2t}\right)$, respectively. The damping factors of both longitudinal and transverse relaxations estimated by our new method, our previously proposed null point spectrum analysis³, and the conventional nonlinear least squares analysis are listed in the following table. It should be noted that our new results are consistent with the null point spectrum analysis and need no subjective initial guess.

Method	$r_1, r_2 \ (\mathrm{s}^{-1})$	$s_1, s_2 \; (\mathrm{s}^{-1})$
our work	2.560, 4.848	3.661, 6.150
null point spectrum analysis	2.173, 4.824	3.498, 6.479
nonlinear least squares analysis	2.339, 4.446	3.144, 5.390

We are now trying to extend this method to more accurate quantification of NMR spctral parameters. Further modification and application shall be reserved for a later occasion.

References

- 1. N. J. Clayden and B. D. Hesler, J. Magn. Reson. 98, 271 (1992). (and references therein)
- 2. R. Kumaresan and D. W. Tufts, IEEE Trans. ASSP-30, 833 (1982).
- 3. (a) W. S. Price, N. H. Ge, L. Z. Hong, and L. P. Hwang, J. Am. Chem. Soc. (in press).
 - (b) N. H. Ge, W. S. Price, L. Z. Hong, and L. P. Hwang, J. Magn. Reson., 97, 656 (1992).
 - (c) W. S. Price, N. H. Ge, and L. P. Hwang, J. Magn. Reson., 98, 134 (1992).
 - (d) T. S. Lee and L. P. Hwang, J. Magn. Reson., 89, 51 (1990).

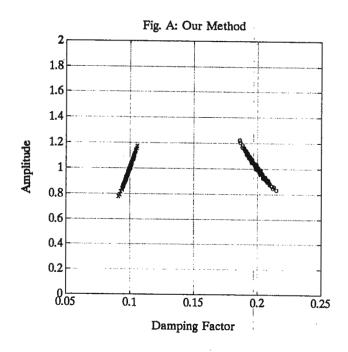
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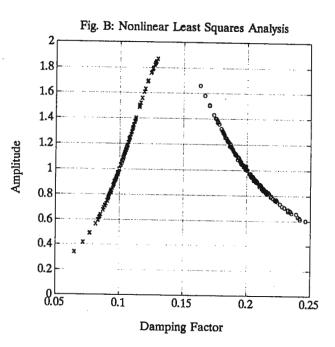
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The Department of Biomedical Engineering at Worcester Polytechnic Institute is seeking a postdoctoral research associate to participate in NMR spectroscopy and imaging studies characterizing porous media using diffusion methods with particular emphasis on oil cores (the candidate will work closely with scientists from Schlumberger-Doll Research). The successful candidate will spend approximately two-thirds of their time on this project. For the remaining portion of the appointment, the candidate will participate in a project evaluating the efficacy of pharmaceutical interventions in the treatment of stroke using diffusion-weighted imaging and multiple-quantum NMR spectroscopy. The individual will also help supervise graduate students. Qualifications for the position include a doctoral degree in a basic science and hands-on experience with NMR spectroscopy and imaging. Salary is commensurate with experience.

The NMR laboratory is located in the Massachusetts Biotechnology Research Park, adjacent to the campus of the University of Massachusetts Medical School and a five minute drive from Worcester Polytechnic Institute. Equipment includes a 2.0T/45 cm GE CSI-II imaging spectrometer, exclusively for research, and two, 1.5T GE Signa clinical instruments (one of which is equipped for clinical spectroscopy), available for both clinical and basic research.

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All Newsletter Correspondence

Should Be Addressed To:

Dr. Bernard L. Shapiro TAMU NMR Newsletter 966 Elsinore Court Palo Alto, CA 94303, U.S.A.

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If the mailing label on your envelope of this issue is adorned with a large <u>red dot</u> or circle: this decoration means that you will not be mailed any more issues until a technical contribution has been received by me.



PLEASE Nage Length Instruction

Attention overseas subscribers: If you must use paper which is longer that 11", <u>please</u> take care that all material (including signatures, addresses - everything!) ends no more than 10" from the top of each of your pages. It is costly to make reductions, and henceforth I reserve the right to chop the excess length off any page, no matter what the result. Beware of the dreaded guillotine! Your cooperation in this matter will be greatly appreciated.

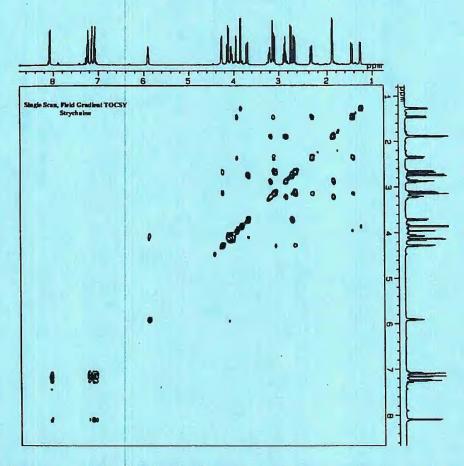
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FROM JEOL USA

Gradient Probes



The above data is a 512 by 512 *TOCSY* spectrum and was obtained on an Alpha-500 in less than 5 minutes with **JEOL**'s gradient probe. This same probe is also designed to do reverse and water suppression experiments. Pulse shaping, three channel operation, dynamic Rf control, matrix shims, as well as unsurpassed field reliability are all standard with the Alpha.

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