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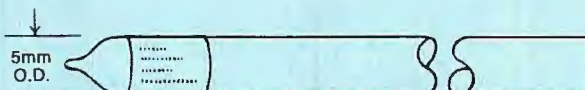
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All Newsletter Correspondence, Etc., Should be Addressed To:

Dr. Bernard L. Shapiro  
 Department of Chemistry  
 Texas A&M University  
 College Station, TX 77843 U.S.A.

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## Pharmaceutical Products Division

Abbott Laboratories  
North Chicago, Illinois 60064

August 24, 1982

Prof. Bernard L. Shapiro  
Department of Chemistry  
Texas A&M University  
College Station, TX 77843

Dear Barry,

Assignment of Some  $^{13}\text{C}$  NMR Chemical Shifts

A compound with beta-lysine was submitted for  $^{13}\text{C}$  NMR analysis. In an attempt to assign all carbons a sample of beta-lysine 1 was also titrated in  $\text{D}_2\text{O}$ . Table I shows the titration values of 1. The titration curve does not clearly assign C-4 and C-5 of 1 since they both have beta shifts. A literature search notes conflicting assignments. In<sup>1</sup> C-4 is assigned to the 23 ppm resonance and C-5 to the 29 ppm resonance. In<sup>2</sup> these assignments are reversed. Recently an acylated derivative of the original compound was submitted for analysis. Since  $^{13}\text{C}$  NMR chemical shifts are now available for the compound without beta-lysine, a titration curve of the sample showed that the acetyl group was attached to the amino group at C-3 of beta-lysine. The titration values for the 3-N-acetyl-beta-lysine group are shown in Table II. Since C-2 and C-4 showed no beta shifts, it is now clear that in beta-lysine C-5 is the resonance at 23 ppm and C-4 is the resonance at 29 ppm at an acid pD. Therefore the assignments in<sup>2</sup> are the correct ones.

Sincerely yours,

*Ruth*

Ruth Stanaszek

RS:dmh

<sup>1</sup>JOC 43, 1282 (1977).

<sup>2</sup>Chem. Pharm. Bull, 25, 280 (1977).

August 24, 1982

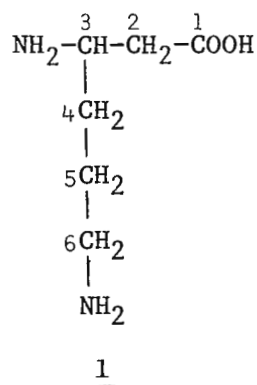


Table I

## Titration Values for Beta-Lysine

<u>Assignment</u>	<u>pD</u>	<u>9.8</u>	<u>9.6</u>	<u>8.9</u>	<u>4.4</u>	<u>3.8</u>	<u>Beta Shift</u>
C-2		43.2	42.1	40.4	38.9	38.8	-4.9
C-3		49.3	49.4	49.5	49.6	49.4	
C-4		32.3	31.6	30.7	29.9	29.9	-2.3
C-5		25.0	24.6	24.1	23.7	23.7	-1.3
C-6		40.3	40.2	39.9	39.8	39.8	

Table II

## Titration Values for 3-N-Acetyl-Beta-Lysine

<u>Assignment</u>	<u>pD</u>	<u>12.3</u>	<u>9.6</u>	<u>8.0</u>	<u>6.3</u>	<u>2.0</u>	<u>Beta Shift</u>
C-2		42.3	42.1	42.0	41.9	41.9	
C-3		48.1	47.7	47.7	47.8	47.9	
C-4		32.1	31.8	31.8	32.0	32.0	
C-5		28.3	24.7	24.2	25.4	25.6	-2.7
C-6		40.9	40.0	39.9	40.0	40.0	

14 September 1982

Professor B L Shapiro  
 Department of Chemistry  
 Texas A & M University  
 College Station  
 Texas 77843  
 U S A

Dear Professor Shapiro

A RABBIT for breeding NOE's

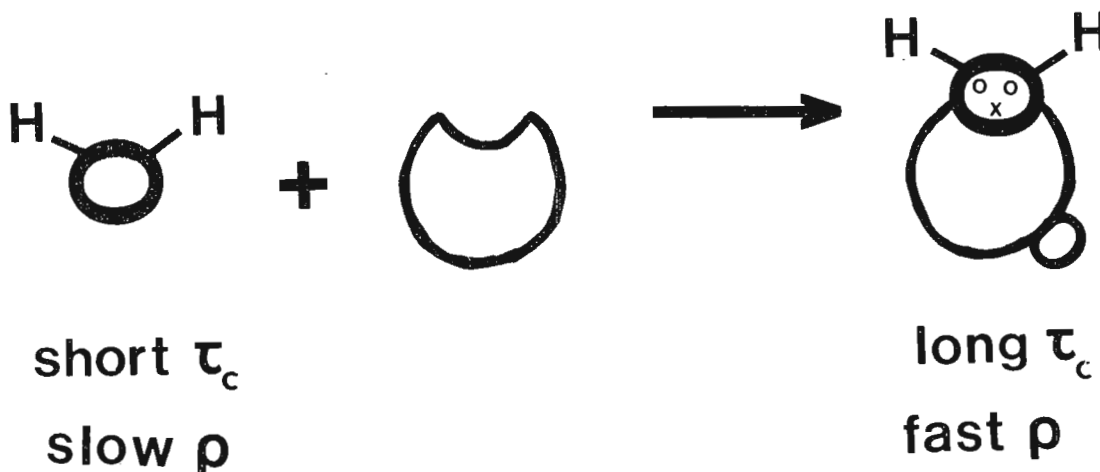
It is my privilege to announce a new advance in animal husbandry which will be of interest to all those who do nOe experiments on small molecules.

We have been using kinetic nOe's to measure interatomic distances and to probe conformational equilibria,<sup>1</sup> but the long  $T_1$  values and slow nOe growth rates ( $\rho_{dd}$ ) demanded painfully long relaxation delays. It took all night to do a simple kinetic nOe experiment. The reason is clear from eqn. 1 which shows that  $\rho$  is proportional to the rotational correlation time  $\tau_c$ :

$$\rho_{dd} \propto \tau_c \cdot r^{-6}$$

Small molecules have short  $\tau_c$  and so breed nOe's slowly.

Much impressed by the fecundity of rabbits we have attempted to emulate their example in the nmr tube. The necessary genetic engineering is indicated below.



In practice this transformation is most easily achieved by binding the molecule of interest to a diamagnetic reagent such as  $\text{La}(\text{FOD})_3$ , metalloporphyrin, cyclodextrin, etc.  $\tau_c$  in the adduct is longer,  $\rho$  is bigger and  $T_1$  is shorter. As a result the nOe's grow faster and sometimes end up larger because  $\rho$  competes better against  $\text{O}_2$  and rust particles. The figure below shows one example: the time saving for a given S/N for the nOe after 1 second irradiation is sixfold.

We hope to publish more examples soon. RABBIT is Relaxation Aided By Binding Tightly.

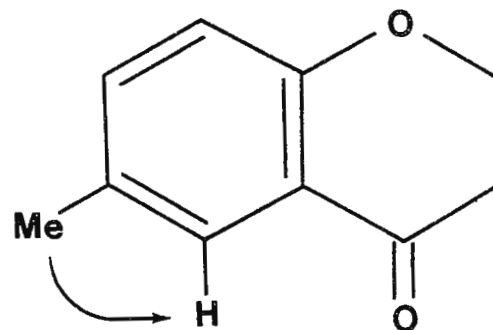
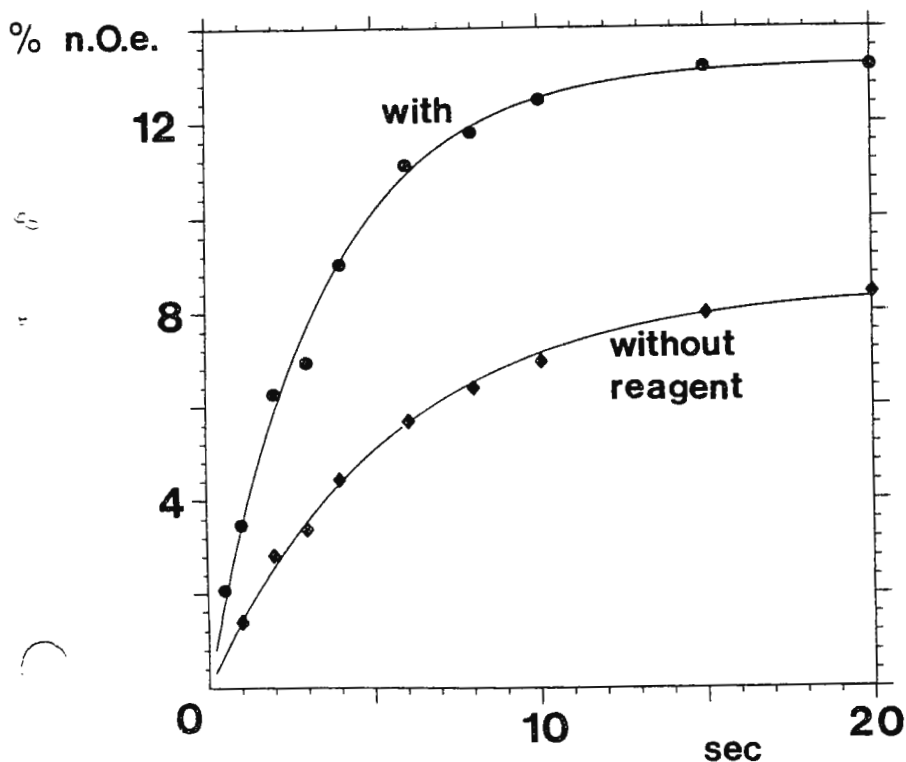
As usual,<sup>1-3</sup> the work was done by John Mersh.

Yours sincerely

*Terry Sanders*

J K M Sanders

1. J. D. Mersh & J. K. M. Sanders, Tetrahedron Letters, 1981, 4029
2. SUDSY : TAMUNMR, January 82 & J. Magn. Reson., Nov. 82
3. Snopake in 2D NMR : J. Magn. Reson. Oct. 82



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DEPARTMENT OF CHEMISTRY

DAVIS, CALIFORNIA 95616

13 September, 1982

Dr. Barry L. Shapiro  
Department of Chemistry  
Texas A and M  
College Station, TX 77843

## PARAMAGNETIC EFFECTS ON DEUTERIUM RELAXATION

Dear Barry,

Sorry about the delay, and the dreaded pink sheet.

We have been applying deuterium NMR to protein dynamics in solution, by studying motional narrowing of  $^2\text{H}$  resonances. Since we are primarily interested in paramagnetic proteins, nonquadrupolar  $^2\text{H}$  relaxation must be accounted for and excluded. Quadrupolar relaxation should be least effective for a deuterated methyl group, due to fast internal motion, and the quadrupolar spin-lattice relaxation should be much slower than the spin-spin relaxation as  $\omega_0\tau_m > 1$  (@ 55 MHz). We measured the  $^2\text{H}$   $T_1$  of the deuterated methyls in 1,3 -  $\text{d}_6$  sperm whale metaquo myoglobin to be 49 ms; the corresponding proton  $T_1$  is 3.8 ms. Upon addition of  $\text{CN}^-$  to yield the low spin form of the protein, the  $^2\text{H}$   $T_1$  was 73 ms; the proton  $T_1$  is 100 ms. Simply scaling the  $T_1$  rates by 1/42.4 and subtracting yields quadrupolar  $T_1$ 's that are within 4% of each other. Thus scaling proton relaxation rates by the ratio of the  $\gamma$ 's, to get the deuterium nonquadrupolar rates, seems to work well in this system.

We are interested in protein side chain dynamics in heme proteins; the deuterated hemins are synthesized by Professor Kevin Smith and his group.

Sincerely,

Robert D. Johnson

Gerd N. La Mar

## GX Report #2

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27th September, 1982.

Professor B.L. Shapiro,  
Department of Chemistry,  
TEXAS A and M UNIVERSITY,  
College Station,  
Texas,  
UNITED STATES OF AMERICA.

Dear Barry,

N.A.T.O. ADVANCED STUDY INSTITUTE ON N.M.R. OF LIQUID  
CRYSTALS

The above Advanced Study Institute will be held from 26th July to 7th August at San Miniato, a Tuscan village about 40 kms from Florence and Pisa. We aim to cover the whole range of the application of N.M.R. to studying oriented liquids including the determination of geometries of small molecules dissolved in liquid crystals, the characterization of order and structure in pure mesophases (thermotropics, lyotropics, discotics, membranes), and the study of dynamical behaviour.

The lectures will be given by N. Boden, J. Charvolin, P. Diehl, J.W. Doane, G.R. Luckhurst, Z. Luz, A. Pines, I.C.P. Smith, R.L. Vold, R.R. Vold, C. Zannoni and the two directors (myself and Carlo Veracini). Attendance at the Institute will be limited to about seventy students, who should preferably be engaged in the study of oriented systems. The cost of food and accommodation for the school will be of the order of \$450.

Intending participants from N.A.T.O. countries may apply for a grant to cover a part of the accommodation costs. Those interested in receiving further details of the Advanced Study Institute should write to me.

Best wishes,

Jim.

J.W. EMSLEY.

Dr. Bernard L. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station  
TX 77843 U.S.A.

J-modulated spin echo on chloralkanes

Dear Barry,

the well known spin echo experiment can be applied with various types of broad band decoupling. We used the following sequence:

C-13: Delay -  $\frac{\pi}{2}$  -  $\tau$  -  $\pi$  -  $\tau$  - ACQUISITION

H-1: BB(-15dB) | BB(-5dB)

The J-modulated spin echo (JMSE) transforms signal multiplicities due to heteronuclear coupling to intensity modulations of the entire BB-decoupled resonance signal. Many applications with  $^1J_{CH} = 125$  Hz have been reported. In special

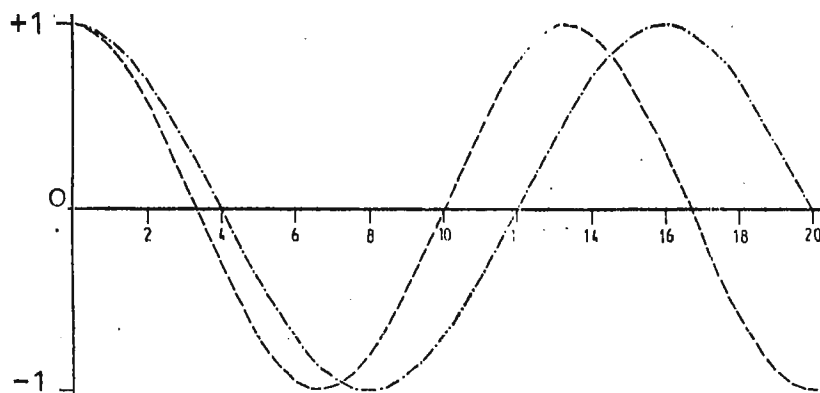


Fig. 1 Intensity modulations of resonance signals by JMSE

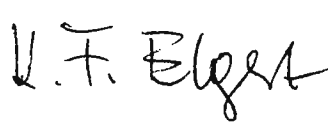
--- = -CH-      - - - = -CHCl-

systems e.g. chloroalkanes, with  $^1J_{CH} = 125$  Hz and  $^1J_{CHCl} = 150$  Hz intensity modulations of resonance signals can be calculated as shown in fig. 1 for -CH- and -CHCl-. JMSE enables selective assignments of structural moieties by resonance signals of zero intensity. This procedure is time saving in comparison to a

full 2D-experiment. However, it is limited to systems with definite coupling constants different by at least 5 Hz. Application is demonstrated in fig. 2 by JMSE spectra of 2,4-dichloro pentane taken at suitable  $\tau$ .

Best regards  
Your sincerely

  
(Prof. Dr. R. Kosfeld)



(Dr. K.-F. Elgert)

a)

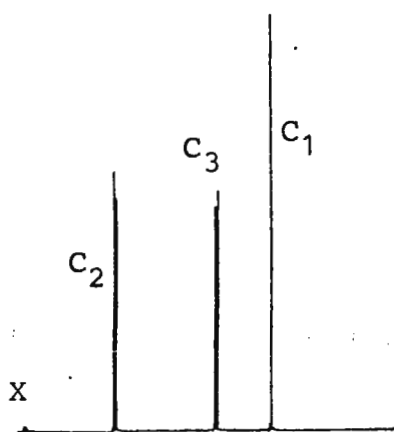
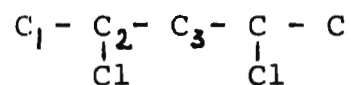


Fig. 2

C-13 nmr spectrum of 2,4-dichloro  
pentane



(mixture of diastereomers)

a) broad band decoupled

b) - e) JMSE spectra with  $\tau = 8,$   
10, 12, 20 msec, resp.

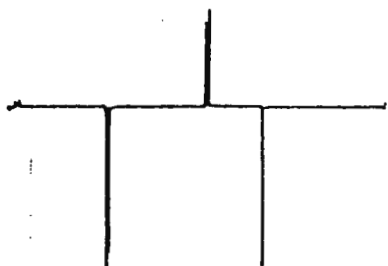
X = solvent,  $CDCl_3$

BRUKER WM 300, pulse repetition:

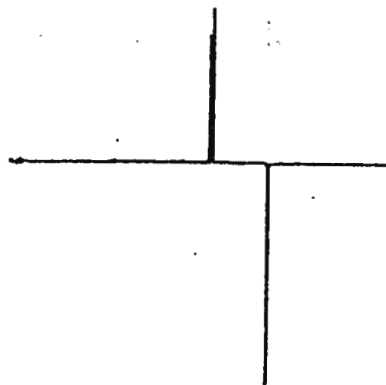
60 sec, pulse width: 17.3  $\mu$ sec,

pulse sequence generated by multi-  
pulser unit

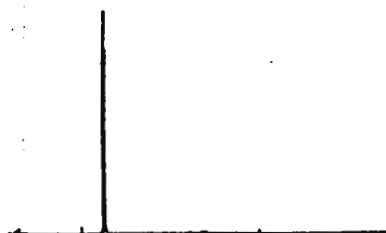
b)



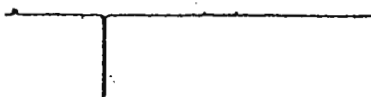
c)



d)



e)



DÉPARTEMENT DE CHIMIE ORGANIQUE

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PROFESSEUR B. P. ROQUES

PARIS, LE September, 17th, 1982

Professor B. L. SHAPIRO  
Department of Chemistry  
Texas A & M University  
College Station  
Texas 77843 - U.S.A.

$^{15}\text{N}$  antisymmetric  $T_1$  measurements by the INEPT method.

Dear Professor Shapiro,

The great sensitivity gain of the INEPT sequence (1) is now well established and looks very attractive for the relaxation measurements of insensitive nuclei such as  $^{15}\text{N}$ . We have developed a method derived from INEPT (2) leading to the "antisymmetric"  $^{15}\text{N}T_1$  a new parameter useful for peptides conformational analysis. The models chosen are  $^{15}\text{N}$  enriched (90%) enkephalin derivatives : Tyr- $^{15}\text{N}$ -Gly- $^{15}\text{N}$ -Gly- $^{15}\text{N}$ -Phe, I, Boc-Tyr- $^{15}\text{N}$ -Gly- $^{15}\text{N}$ -Gly- $^{15}\text{N}$ -Phe- $\text{OCH}_3$ , II, and Tyr- $^{15}\text{N}$ -Gly- $^{15}\text{N}$ -Gly- $^{15}\text{N}$ -Phe- $^{15}\text{N}$ -Leu, III. If one consider in the peptide backbone each  $^{15}\text{N}$  and its  $^1\text{H}$  linked nucleus as independant AX spin-systems ( $A = ^{15}\text{N}$ ,  $X = ^1\text{H}$ ), three observable parameters can be measured for each one : the total magnetization related to the symmetric  $T_1^S$  for both nitrogen,  $M_N$ , and proton,  $M_H$ , and the intensity difference within each doublet  $M_{NH}$ , related to the antisymmetric  $T_1^a$  (3).

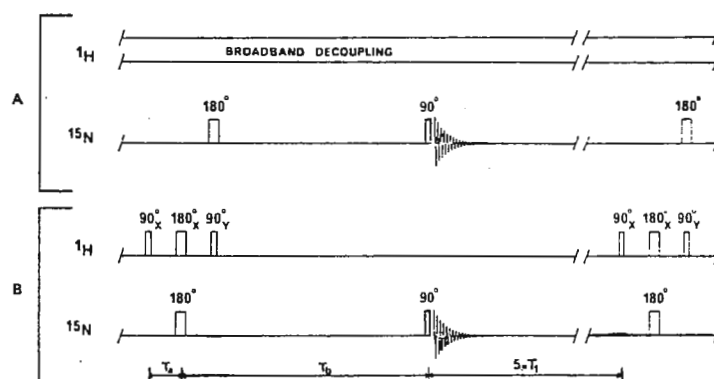


Figure 1.

Pulse sequence used for relaxation measurements.

A : Symmetric  $T_1$  (Freeman-Hill method).

B : Antisymmetric  $T_1$  (slightly modified INEPT sequence).

-  $^{15}\text{N}$   $180^\circ$  pulse = 42  $\mu\text{s}$  ; -  $^1\text{H}$   $180^\circ$  pulse = 400  $\mu\text{s}$ .

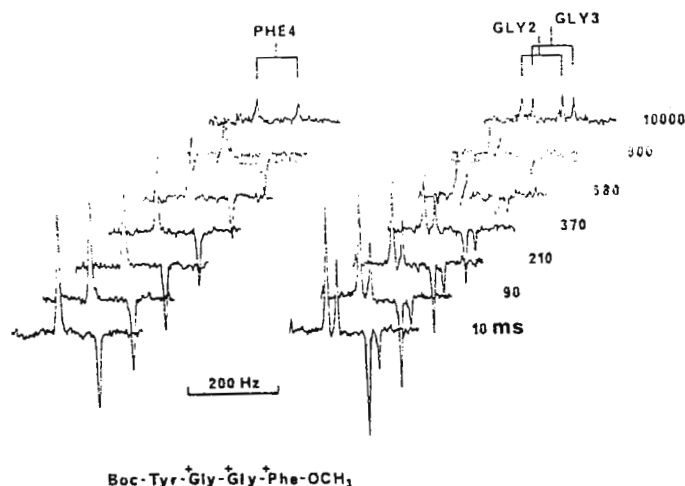


Figure 2.

Antisymmetric  $T_1$  measurement for  
Boc-Tyr-Gly-Gly-Phe-OCH<sub>3</sub>  
in Me<sub>2</sub>SO (0.1 M) at 310°K (50 scans).

A sequence excitation (B) is proposed to measure  $T_1^a$  and compared with the sequence for measurement of the conventional  $T_1^s$  (A). The sequence A is characterized by  $^1\text{H}$  broad-band irradiation during the evolution period  $\tau_b$  whereas such irradiation does not occur in the sequence B. By contrast the  $T_1^a$  are obtained by using an excitation sequence based on the spin-echo polarization transfer method (INEPT) firstly described by Morris and Freeman. The original INEPT sequence which creates a  $^{15}\text{N}$  magnetization in the x-y plane by a  $^{15}\text{N}$  modulated proton spin-echo is just altered by suppressing the second  $^{15}\text{N}$  90° pulse (Figure 1.).

After the INEPT excitation the two components of the  $^{15}\text{N}$  doublet are enhanced but in the opposite direction. Therefore the modified pulse excitation allows to observe the recovery (along the x-axis) of the magnetization-difference between the two components of the doublet for  $\tau_b$  (Figure 2.). As expected, in each peptide models  $T_1^s$  and  $T_1^a$  values differ one from each other as shown in Table 1. Moreover a large variation range in the  $T_1^a$  appears in free peptides what is not the case of the protected one according to large differences in conformational flexibility occurring in these compounds (4).

Table 1.  $^{15}\text{N}$  symmetric  $T_1^s$  and antisymmetric  $T_1^a$  spin-lattice relaxation times (in sec) at 310 K for compounds : Tyr-\*Gly-\*Gly-\*Phe, I, Boc-Tyr-\*Gly-\*Gly-\*Phe-OCH<sub>3</sub>, II, and Tyr-\*Gly-\*Gly-\*Phe-\*Leu, III.

Compound	Residue	$T_1^s$	$T_1^a$
<u>I</u>	Gly <sup>2</sup>	0.95	0.028
	Gly <sup>3</sup>	0.95	0.34
	Phe <sup>4</sup>	0.85	0.22
<u>II</u>	Gly <sup>2</sup>	1.35	0.54
	Gly <sup>3</sup>	1.40	0.61
	Phe <sup>4</sup>	1.60	0.78
<u>III</u>	Gly <sup>2</sup>	1.15	0.028
	Gly <sup>3</sup>	1.15	0.52
	Phe <sup>4</sup>	0.90	0.47
	Leu <sup>5</sup>	0.90	0.41

a)  $T_1$  have internal estimated error less than 10% (90% confidence interval), except for the Gly<sup>2</sup> $T_1$ 's in the free peptides (~20%).

Sincerely yours.

D. Marion

Ch. Garbay

B. Roques

Dominique MARION

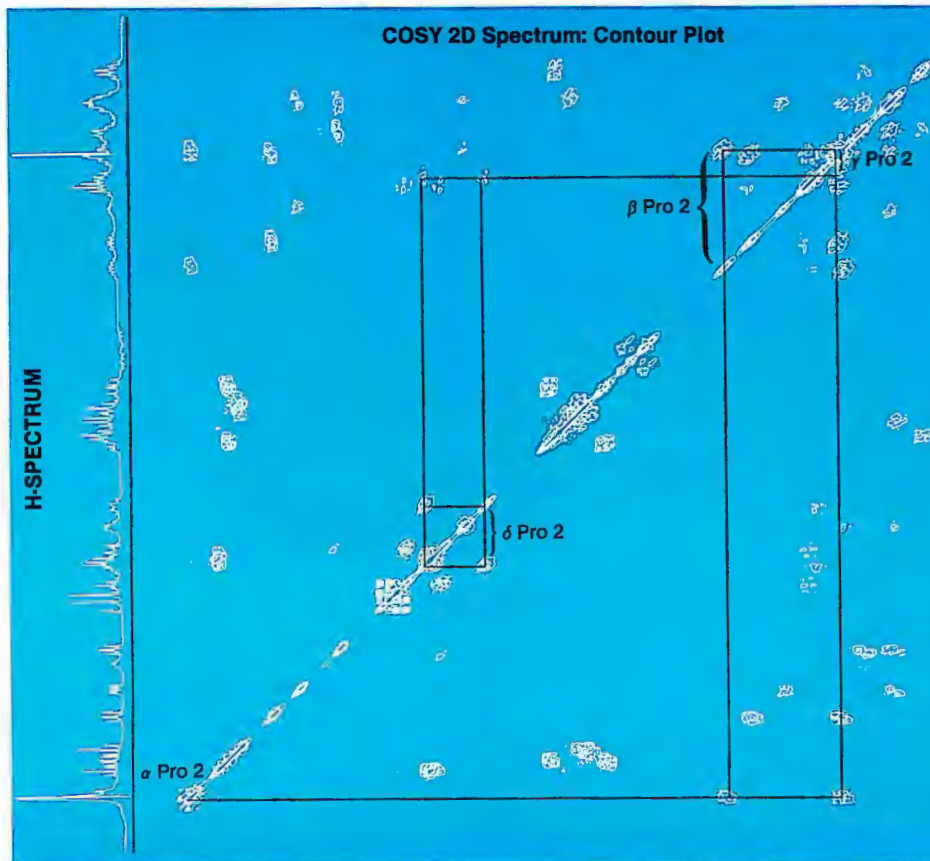
Christiane GARBAY-JAUREGUIBERRY

Bernard P. ROQUES

- (1) G.A. Morris, R. Freeman, J. Amer. Chem. Soc. (1979), 101, 560-563.
- (2) D. Marion, C. Garbay-Jaureguiberry and B.P. Roques, J. Amer. Chem. Soc. (1982) in press.
- (3) L.G. Werbelow, D.M. Grant, Adv. Magn. Reson. (1979), 9, 189-299.
- (4) D. Marion, C. Garbay-Jaureguiberry, B.P. Roques, J. Magn. Res. (1982) in press.

# $C_{50}H_{73}N_{15}O_{11}$ by 2-D NMR

When complex molecules are submitted to analysis—e.g. the oligopeptide Bradykinin sample in this experiment—two-dimensional spectra such as proton correlated spectroscopy (COSY) can often produce dramatic simplification of seemingly intractable spectra, even at high magnetic field. Here is evidence:



## Chemical Shift Assignments Using 2D Proton Correlated Spectroscopy (COSY).

The pulse sequence for COSY is given by:

$$90^\circ_x - t_1 - 90^\circ \pm x - \text{Acq}(t_2) \pm y \quad (1)$$

The chemical shifts of mutually coupled protons can be extracted from over-lapping multiplets by means of a contour plot obtained from the COSY Experiment Data. The normal Proton Spectrum is represented along the diagonal axis, and the symmetrical off-diagonal cross peaks provide the clue to any coupled protons.

The COSY Contour Plot of Bradykinin triacetate (Arg-Pro\*-Pro-Gly-Phe-Ser-Pro-Phe-Arg), illustrates the technique on the aliphatic moiety. With the known position of the H $\alpha$ -Pro 2 (\*), the chemical shifts of all the six remaining protons (2 $\beta$ ; 2 $\gamma$ ; 2 $\delta$ ) can easily be located as shown by the connecting lines.

(1) Ad Bax and Ray Freeman, JMR 44, p. 542-561 (1981)

**Q.E.D.** Both spectrum and contour plot of this COSY experiment were produced on a WM 400 at the Bruker Applications Laboratory. The WM Series of high-field NMR spectrometer systems comes with an extensive software system, including programs for 2-D processing display and plotting. A full-color graphic display processor further facilitates speed and clarity of stack and contour plots.

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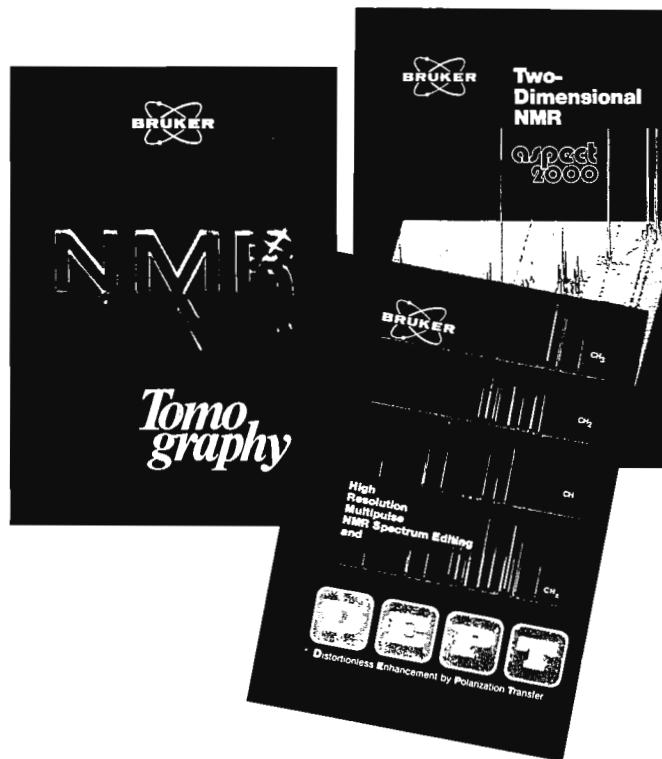
## NMR-Tomography

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In a short survey it is shown that for the last twenty years the instrumental development in the pulsed NMR field has been synonymous with the name of BRUKER and it is pointed out that the first commercially available Fourier Transformation (FT) spectrometers were developed by BRUKER in 1969. Since NMR tomography is based on both "pulsed" and "FT"-NMR, the unique experience of BRUKER in these fields represents the ideal basis for the recently developed imaging systems.

After a short introduction, the principles of NMR are described in the brochure followed by a short representation of the "Projection-Reconstruction-Technique". Due to the expected extraordinary importance of NMR tomography in the field of diagnostic medicine a comparison of the average X-ray tissue contrast with NMR data is given as well as some remarks about theoretically possible risks for patients. At the end of this brochure an "outlook" is given into new applications and of the expected development of NMR tomography.



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September 20, 1982

Professor B.L. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station, TX 77843

$^{13}\text{C}$  NMR Spectra of Polyethylene Oxide Derivatives

Dear Professor Shapiro,

We have been studying ionophores which can be "switched on" and "switched off" by closure and cleavage of disulfide bonds. We have found  $^{13}\text{C}$  spectra of great utility in making structural assignments and following reactions. In the course of our work we have made a number of spectral assignments on final ionophores and intermediates, some of which are given in the table. Spectra were measured on an FX-60 spectrometer at 15 MHz. Of special interest are the shifts which follow closure of the disulfide: + 15 ppm and -3.5 ppm for the  $\alpha$  +  $\beta$  carbons, respectively, due to the  $\beta$  +  $\gamma$  effects of the second sulfur atom.

	$\text{C}_1$	$\text{C}_2$	$\text{C}_4$	$\text{C}_5$	$\text{C}_7$	$\text{C}_8$
$\text{HO}(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_2\text{CH}_2\text{OH}$	61.4	72.7	70.0*	70.5*		
$\text{HO}(\text{CH}_2\text{CH}_2\text{O})_4\text{CH}_2\text{CH}_2\text{OH}$	61.4	72.5	70.1	70.4	70.4	
$\text{HO}(\text{CH}_2\text{CH}_2\text{O})_5\text{CH}_2\text{CH}_2\text{OH}$	61.6	72.5	70.5	70.5	70.5	70.5
$\text{Br}(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_2\text{CH}_2\text{Br}$	30.3	71.1	70.5	70.5		
$\text{Br}(\text{CH}_2\text{CH}_2\text{O})_4\text{CH}_2\text{CH}_2\text{Br}$	30.3	71.2	70.6	70.6	70.6	
$\text{Br}(\text{CH}_2\text{CH}_2\text{O})_5\text{CH}_2\text{CH}_2\text{Br}$	30.3	71.2	70.6	70.6	70.6	70.6
$\text{HS}(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_2\text{CH}_2\text{SH}$	24.2	72.8	70.2*	70.6*		
$\text{HS}(\text{CH}_2\text{CH}_2\text{O})_4\text{CH}_2\text{CH}_2\text{SH}$	24.3	72.9	70.2	70.6	70.6	
$\text{HS}(\text{CH}_2\text{CH}_2\text{O})_5\text{CH}_2\text{CH}_2\text{SH}$	24.2	72.8	70.2	70.6	70.6	70.6
$\text{S}(\text{CH}_2\text{CH}_2\text{O})_3\text{CH}_2\text{CH}_2\text{S}$	39.1	69.3	70.9*	71.0*		
$\text{S}(\text{CH}_2\text{CH}_2\text{O})_4\text{CH}_2\text{CH}_2\text{S}$	39.1	69.5	70.3	70.6	70.6	70.6
$\text{S}(\text{CH}_2\text{CH}_2\text{O})_5\text{CH}_2\text{CH}_2\text{S}$	39.2	69.2	70.2	70.8	70.8	70.8

\*assignments may be reversed

Sincerely

Morton Raban  
Professor of Chemistry

Farouk Kandil  
Visiting Professor of Chemistry  
(on sabbatical leave from the  
University of Aleppo, Syria)



U.S. Department of Energy  
Laramie Energy Technology Center  
P.O. Box 3395, University Station  
Laramie, Wyoming 82071

September 23, 1982

Professor B. L. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station, TX 77843

Dear Barry:

RE: Hydrocarbon Type Analysis of Fossil Fuels Using Spectral Editing Techniques.

We have been examining the feasibility of utilizing  $^{13}\text{C}$  NMR spectral editing techniques to quantitate hydrocarbon types found in fossil fuel. Several methods for spectral editing are given in the literature (1-4), but only the Gated Spin-Echo Technique (GASPE)<sup>3</sup> and the Distortionless Enhancement by Polarization Transfer (DEPT)<sup>1</sup> are applicable to quantitative hydrocarbon type analysis.

In order to compare the GASPE and DEPT methods, a test mixture was prepared. The composition of this mixture and its method of preparation are given in Table I (attachment).

The GASPE, Conventional Spin/Echo (CSE) and DEPT sequences were written with modifications of the literature for the JEOL FX-270 NMR spectrometer. By appropriate combination of addition and subtraction of GASPE and CSE spectra at  $\tau$  values of  $1/J$ ,  $1/2J$ ,  $1/4J$ , and  $3/4J$ , spectra containing only C, CH,  $\text{CH}_2$ , and  $\text{CH}_3$  were obtained. From the integration of this spectra, the percent carbon types were calculated for the mixture. Table II lists the percent of aliphatic carbon types using the GASPE method. An examination of the data in Table II indicates that quantitation of carbon types present in minor amounts is not accurate.

We are now completing the work using the DEPT sequence. Hopefully a full paper discussing the pros and cons of the two methods will be published in the near future.

Sincerely,

Daniel A. Netzel and  
LETC

Ed Clennan  
Chemistry Dept.  
Univ. Wyoming

Attach.

TABLE I: TEST MIXTURE<sup>a</sup>

COMPOUND	Moles used	#C	#CH	#CH <sub>2</sub>	#CH <sub>3</sub>
toluene	.01	1	5	0	1
2,2,4-trimethylpentane	.01	1	1	1	5
o-ethyltoluene	.01	2	4	1	2
acenaphthene	.01	4	6	2	0
2,3-dimethylnaphthalene	.01	4	6	0	3
1-methylnaphthalene	.01	3	7	0	1
1,2,3,4-tetrahydro- naphthalene	.01	2	4	4	0
hexane	.01	0	0	6	0
heptane	.01	0	0	5	2
tetradecane	.01	0	0	12	2
TOTAL		17	33	31	15
% TOTAL		17.71	34.38	32.29	15.63

<sup>a</sup>The samples were prepared by taking this mixture and diluting it to 25 ml with COCl<sub>3</sub>. The samples were sealed in 5 and 10 mm NMR tubes. Some of the samples were also .04 M in Cr(AcAc)<sub>3</sub>.

TABLE II: QUANTITATION OF ALIPHATIC CARBONS WITH THE GASPE METHOD

	EXPERIMENTAL				CALCULATED	
	CARBONS		% CARBON		CARBONS	% CARBON
CH <sub>3</sub>	14.06 <sup>a</sup>	(15.52) <sup>b</sup>	29.31	(32.34)	15	31.25
CH <sub>2</sub>	31.96	(31.93)	66.58	(66.53)	31	64.58
CH	10.48	( 3.98)	21.84	( 8.30)	1	2.08
C	1.24	( 5.01)	2.6	(10.44)	1	2.08
	57.74	(56.44)	120.33	(117.61)	48	99.99

<sup>a</sup>First Attempt

<sup>b</sup>Second Attempt

## REFERENCES

1. Bendall, M. R., D. M. Doddrell, D. T. Pegg, and W. E. Hall. Brochure by Bruker, 1982.
2. Patt, S. L. and J. N. Shoolery. J. Magn. Resonance **46**, 535 (1982).
3. Cookson, D. J. and B. E. Smith. Org. Magn. Resonance **16**, 111 (1981).
4. Bendall, M. R., D. M. Doddrell, and D. T. Pegg. J. Am. Chem. Soc. **103**, 4603 (1981).

Poul Erik Hansen, Institut I  
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Denmark

Professor B.L.Shapiro  
Department of Chemistry  
Texas A&M University  
College Station  
Texas 77843  
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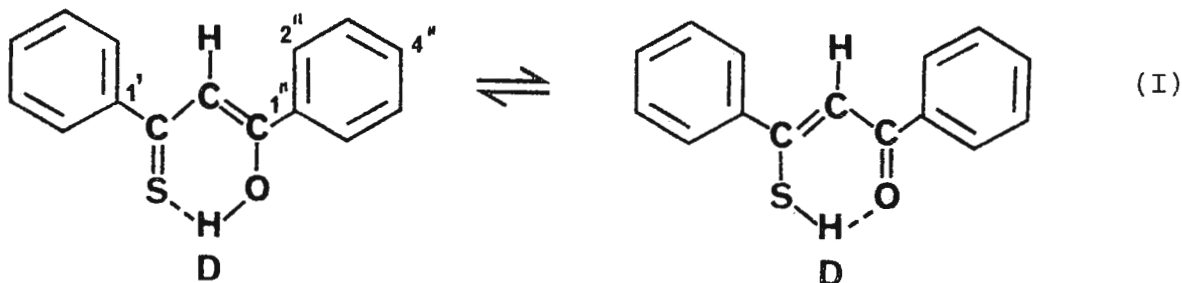
September 16 1982

Dear Professor Shapiro

DOES THE BORN-OPPENHEIMER APPROXIMATION BREAK DOWN IN THE  
CASE OF LONG-RANGE ISOTOPE EFFECTS?

In response to your colorfull reminders a short description  
of some results from my latest visit to Harald Günther at  
Siegen.

From our results in a previous paper<sup>1</sup> it is clear that sub-  
stituents at one of the phenyl rings in the  $\beta$ -thioxoketonesy-  
stem may change the enol-enethiol equilibrium, but also that  
deuterium substitution at the S or O may perturb the equili-  
brium (I) giving rise to long-range equilibrium isotope effects  
on  $^{13}\text{C}$  as shown in Table 1.



What we did this time was to look at the pentadeuteroderivative  
as shown in II. As seen from table 1, did this derivative  
also show small, but significant isotope effects( large for

long-range isotope effects ). As the effects furthermore are proportional to those observed when the equilibrium is perturbed by other means (D on S or O ) we do not hesitate to claim that the pentadeuteration leads to a change in the equilibrium.

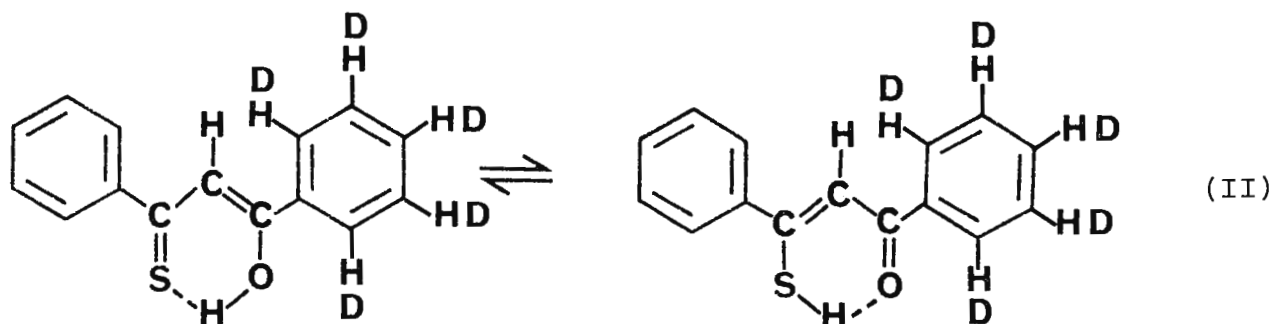


Table 1. Deuterium isotope effects on  $^{13}\text{C}$  nuclear shielding in ppm

	CS	CO	CH	C-1'
I	-4.69	+1.69	+0.32	-0.59
II	-0.11	+0.04	+0.01	-0.01

The change in the equilibrium shows that the  $\beta$ -thioxoketo-system is a very sensitive gauge of substituent effects. However, the remaining question is why does the equilibrium change.

The magnitude of the isotope effects in case II does not depend upon the ration of H and D compound in the mixture and it is not changed upon a ten fold dilution.

We consider two possibilities. The first is that the  $\text{COH-C}_6\text{H}_5$  part not is planar and as the five H are exchanged with five D the steric interaction between the COH group and the phenyl ring is reduced. The other possibility is that the  $\text{C}_6\text{D}_5$  radical is slightly more electronegative than the  $\text{C}_6\text{H}_5$  radical. This however means that the Born-Oppenheimer approximation not is valid in this case. A heretic thought among theoreticians. Nevertheless, many other data in the litterature point towards a break down at least for the very small effects observed in long range isotope effects.

I hope some theoretician would like to comment on why they believe so strongly in the B-O app. and what experimental evidence on the same level as long range isotope effects support the B-O approximation.

1. P.E.Hansen, F.Duus and P.Schmitt, OMR 18,58(1982)

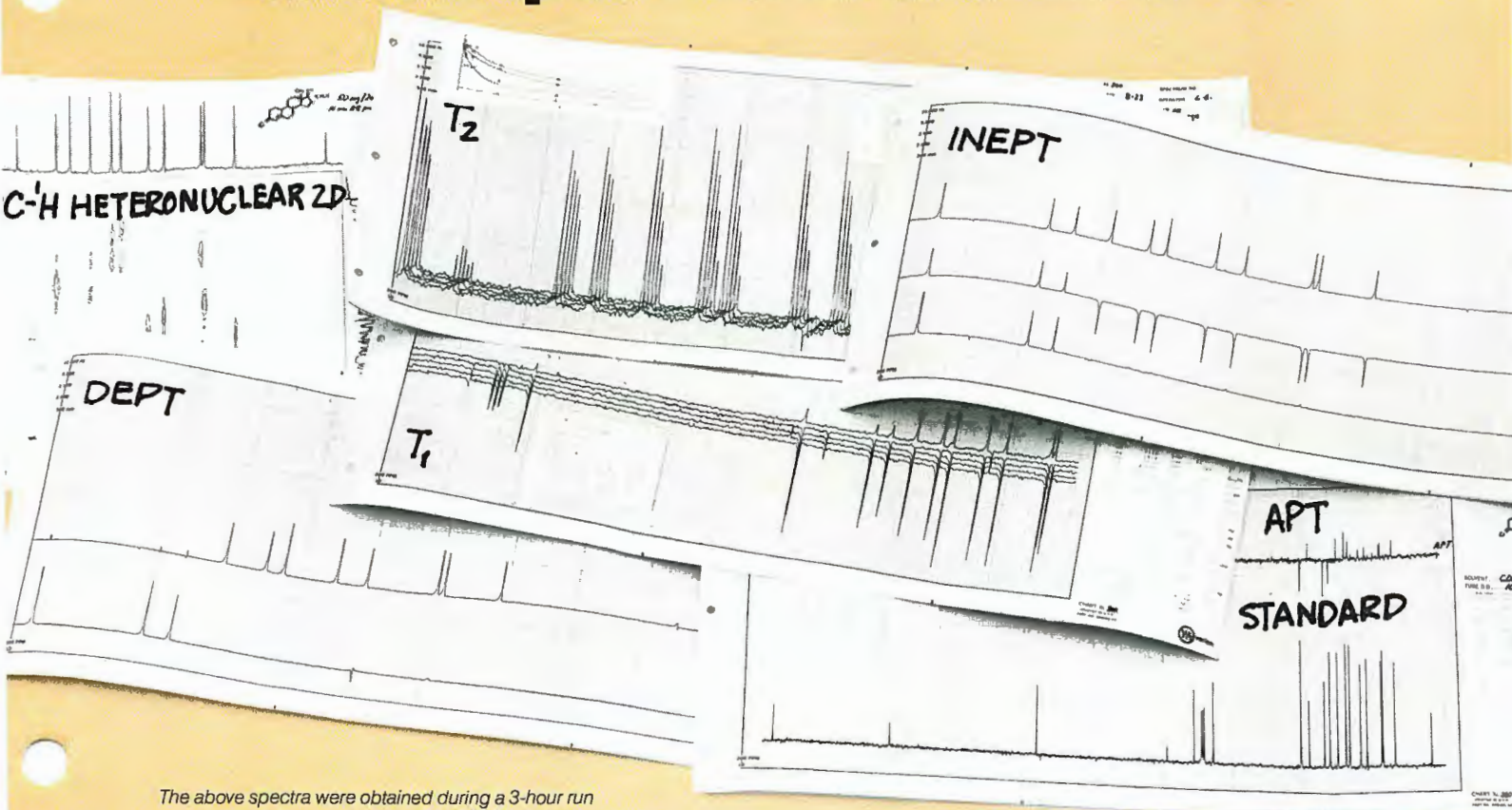
Yours sincerely



Poul Erik Hansen

PE! The isotope effect is defined as  $\Delta^n = \delta_{C(H)} - \delta_{C(D)}$

# Some manufacturers claim these experiments are difficult



The above spectra were obtained during a 3-hour run on an XL-300 Superconducting FT NMR Spectrometer System.

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Dr. Peter Rinaldi is a chemist at the *Major Analytical Instruments Facility*, Greenwood, Ohio. MAIF is a research and testing facility serving Case Western Reserve University and scientists throughout the Ohio Valley region. All quotes are from the MAIF NEWSLETTER, Vol. 1, Issue 3, March 1982, reprinted courtesy of MAIF.

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Professor B.B. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station  
Texas 77843  
U.S.A.

Ihr Zeichen

Unser Zeichen  
I.u./Ja

Ihre Nachricht vom

### Shielding Constants of Copper and Gallium

Dear Barry,

as addition to the fruitful discussions on "shielding" and "screening" during the summer school at Stirling, I would like to direct the attention on further available shielding constants in the free atomic shielding scale ( $\sigma^*$ ):

1) For copper  $\sigma^*$  is given in the following paper:

O. Lutz, H. Oehler, and P. Kroneck,  
Z. Physik A 288, 17 (1978); in the meanwhile further  
shielding data are available on copper compounds:  
O. Lutz, H. Oehler, P. Kroneck, Z. Naturforsch. 33a,  
1021 (1978), P. Kroneck, O. Lutz, A. Nolle, H. Oehler,  
Z. Naturforsch. 35a, 221 (1979); K.O. Becker, H.P. Schäfgen,  
Sol. State Comm. 32, 1107 (1979)

2) For Gallium  $\sigma^*(^{71}\text{Ga}^{3+} \text{ in } \text{H}_2\text{O}) = -880(45) \cdot 10^{-6}$  was found:

J. Kodweiß, O. Lutz, W. Messner, K.R. Mohn, A. Nolle,  
B. Stütz, D. Zepf, J. Magn. Res. 43, 495 (1981)

From the figure given there, the influence of the different  
halides ligands is very obvious.

Sincerely yours



(Otto Lutz)

# CONCORDIA UNIVERSITY



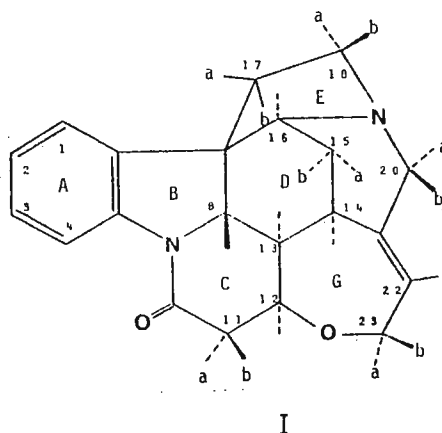
September 23, 1982

Dr. B.L. Shapiro,  
Department of Chemistry,  
Texas A & M University,  
College Station, Texas 77843,  
U.S.A.

Dear Barry,

## APPLICATION OF NOED MEASUREMENTS TO STRYCHNINE

In the course of some work with Jack Edward, of McGill University, on the application of  $^1\text{H}$  spin-lattice relaxation measurements to the structure and stereochemistry of some strychnine sulfonic acids, we turned up some apparently anomalous relaxation rates. These anomalies seemed to disappear if some of the previously reported (1)  $^1\text{H}$  chemical shifts for strychnine (I) were re-assigned. We therefore undertook a re-examination of the assignments, using nuclear Overhauser effect difference (nOed) measurements at 400 MHz.



Those who attended the 1982 ENC meeting in Madison will recall that Dr. Suzanne Wehrli displayed in the Bruker suite two examples of applications of 2D techniques to strychnine -  $^1\text{H}$ - $^{13}\text{C}$  and  $^1\text{H}$ - $^1\text{H}$  shift correlation (COSY). We are indebted to Dr. Wehrli for copies of her plots. Her work provides an elegant verification of the assignment of the  $^1\text{H}$  signals to specific sites through scalar coupling connectivities. Our nOed work has determined the dipolar relaxation connectivities which establish the stereochemical relationships among the protons.

Strychnine is a good molecule on which to apply nOed measurements, since the spectra at 400 MHz (0.1 M in  $\text{CDCl}_3$  or  $\text{DMSO-d}_6$ ) are well dispersed. We used a slight modification of

the nOed procedure reported by Hall and Sanders (2). Degassing was unnecessary, since intramolecular relaxation is efficient in a molecule of the size and rigidity of strychnine. The data were interpreted with the aid of molecular models and calculated enhancements.

From irradiation of the H-8, 13, 15a, 15b, 18a, 18b, 20a, 20b, and 22 transitions we established that the assignments for the geminal pairs of protons at C-15, C-18, and C-20 should be reversed. The use of both CDCl<sub>3</sub> and DMSO-d<sub>6</sub> solutions to modify chemical shifts, and computer simulations using LAOCOON III led to a revised set of coupling constants for the four spin system H-17a, 17b, 18a, and 18b. The revised chemical shifts and coupling constants are listed in the Table.

Best regards,

Yours sincerely,

*Walter*

Walter J. Chazin

*Laurie*

Laurie D. Colebrook

LDC/dg

- 
1. J.C. Carter, G.W. Luther, and T.C. Long, J. Magn. Reson., 15, 122 (1974).
  2. L.D. Hall and J.K.M. Sanders, J. Am. Chem. Soc., 102, 5703 (1980).

Table. Revised <sup>1</sup>H chemical shifts and coupling constants for strychnine<sup>a</sup>

Proton	$\delta^b$	Coupling constants, J (Hz) <sup>c</sup>			
15a	2.36	15a,15b = -14.5	15a,16 = 4.9	15a,14 = 4.0	
15b	1.46	15b,16 = 2.0	15b,14 = 1.0		
17a	1.88	17a,17b = -15.5	17a,18a = 8.4	17a,18b = 11.7	
17b	1.89	17b,18a = 0.1	17b,18b = 6.9		
18a	3.19	18a,18b = -10.2			
18b	2.87				
20a	3.70	20a,20b = -14.7	20a,22 = 0.9		
20b	2.72				

<sup>a</sup>The remaining assignments were in agreement with those in Ref. 1.

<sup>b</sup>Ppm from TMS, CDCl<sub>3</sub> solutions.

<sup>c</sup>Negative signs were assumed for geminal coupling constants.



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CENTRAL RESEARCH & DEVELOPMENT DEPARTMENT  
EXPERIMENTAL STATION

1982 September 27

Professor Bernard L. Shapiro  
Department of Chemistry  
Texas A and M University  
College Station, Texas 77843

Dear Professor Shapiro,

Selenium-77 NMR of Some Fluorinated  
Alkenylphenyl and Alkylphenylselenides

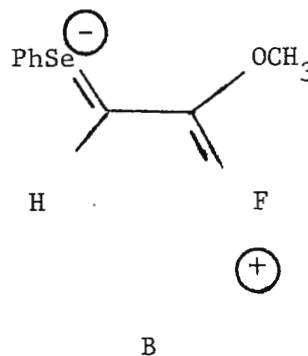
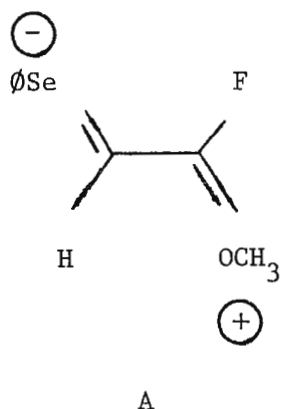
Organoselenium compounds have received much attention as reagents in organic synthesis.<sup>1</sup> Despite this interest, relatively little has been reported on the use of selenium-77 NMR as a tool for structural elucidation of organoselenium compounds<sup>2</sup> and only one article appeared in the area of fluoro-organoselenides where the fluorines are on the hydrocarbon portion.<sup>3</sup> The selenium-77 isotope has a natural abundance of 7.58% and has approximately 3 times the receptivity of carbon-13. In this note we wish to report the <sup>77</sup>Se chemical shifts and <sup>77</sup>Se-<sup>19</sup>F coupling constants for a few fluorinated alkylphenyl and alkylphenylselenides.<sup>4</sup>

The proton decoupled <sup>77</sup>Se spectra were recorded on a Varian FT80A spectrometer at 15.167 MHz in CDCl<sub>3</sub>. The <sup>77</sup>Se chemical shifts, measured relative to phenylvinylselenide (i.e. 395.5 ppm downfield from dimethylselenide) and the <sup>77</sup>Se-<sup>19</sup>F coupling constants for the alkenylphenyl and alkylphenyl selenides studied are listed in Tables I and II.

From Tables I and II we see the following:

1. Trans Se-F coupling constants are larger than cis in the fluorinated alkenylphenyl selenides.

2. A selenium trans to a fluorine, as in compound 2, is 20 ppm upfield relative to the cis fluorine compound. However, when a resonance electron donor is trans to a selenium, such as methoxy, then the selenium trans to the fluorine is downfield relative to the cis, due possibly to greater contribution of resonance structure A than resonance structure B.



3. An alkyl group geminal to a selenium (compd 7) deshields the selenium and also reduces the trans  $^3J_{\text{Se-F}}$  coupling constant while increasing the cis.

4. A chlorine atom  $\gamma$  to a selenium in the case of alkyl selenides causes a 24 ppm downfield shift relative to a fluorine and methoxy group. This may be due to some conformational or electronic effect.

#### References

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Compound 9 was prepared by addition of PhSeCl to vinyl fluoride; treatment of 9 with methanolic KOH gave compounds 2 and 3 which were separated by careful spinning band distillation (A. E. Feiring, unpublished results).

We hope this limited amount of data can generate further study of selenium-77 NMR by your readers.

Sincerely yours, ,

*F Davidson*  
F. Davidson

*A E Feiring*  
A. E. Feiring

FD:AEF/dew

Please credit this contribution to the account of D. D. Bly

Table I

Selenium-77 Chemical Shifts and  $^{77}\text{Se}$ - $^{19}\text{F}$  Coupling Constants  
for Fluorinated Alkenylphenyl Selenides

Compounds	$\delta^a$	$^3J_{\text{SeF}}$
1. $\text{PhSeCH=CH}_2$	0	
2. $\text{PhSe} \begin{array}{c} \text{H} \\ \diagup \\ \text{C}=\text{C} \\ \diagdown \\ \text{H} \end{array} \begin{array}{c} \text{H} \\ \diagdown \\ \text{C} \\ \diagup \\ \text{F} \end{array}$	-125.4	51.1
3. $\text{PhSe} \begin{array}{c} \text{F} \\ \diagup \\ \text{C}=\text{C} \\ \diagdown \\ \text{H} \end{array} \begin{array}{c} \text{H} \\ \diagdown \\ \text{C} \\ \diagup \\ \text{H} \end{array}$	-105.4	7.1
4. $\text{PhSe} \begin{array}{c} \text{OCH}_3 \\ \diagup \\ \text{C}=\text{C} \\ \diagdown \\ \text{H} \end{array} \begin{array}{c} \text{F} \\ \diagdown \\ \text{C} \\ \diagup \\ \text{F} \end{array}$	-143.7	40.0
5. $\text{PhSe} \begin{array}{c} \text{F} \\ \diagup \\ \text{C}=\text{C} \\ \diagdown \\ \text{H} \end{array} \begin{array}{c} \text{OCH}_3 \\ \diagdown \\ \text{C} \\ \diagup \\ \text{OCH}_3 \end{array}$	-154.0	<1
6. $\text{PhSe} \begin{array}{c} \text{F} \\ \diagup \\ \text{C}=\text{C} \\ \diagdown \\ \text{H} \end{array} \begin{array}{c} \text{F} \\ \diagdown \\ \text{C} \\ \diagup \\ \text{F} \end{array}$	-141.7	3.3 (cis) 41.0 (trans)
7. $\text{PhSe} \begin{array}{c} \text{F} \\ \diagup \\ \text{C}=\text{C} \\ \diagdown \\ \text{H} \end{array} \begin{array}{c} \text{F} \\ \diagdown \\ \text{C} \\ \diagup \\ \text{F} \end{array}$ ( $\text{EtO}_2\text{C}$ ) <sub>2</sub> CCH <sub>2</sub>	-95.5	5.9 (cis) 19.5 (trans)

a - negative sign is upfield from phenylvinyl selenide.

b - absolute value of coupling constants are given.

Table II

$^{77}\text{Se}$  Chemical Shifts and  $^{77}\text{Se}$ - $^{19}\text{F}$  Coupling Constants for  
Fluorinated Alkylphenyl Selenides

Compounds	$\delta^a$ ppm	$^3J_{\text{Se-F}}(\text{Hz})$
8. $\text{PhSeCH}_2\text{CF}_2\text{Cl}$	-101.5	3.9
9. $\text{PhSeCH}_2\text{CHFCl}$	-102.0	7.6
10. $\text{PhSeCH}_2\text{CF}_2\text{OCH}_3$	-125.1	3.2
11. $\text{PhSeCH}_2\text{CF}_3$	-125.0	3.9
12. $\text{PhSeCH}(\text{CF}_3)\text{CH}_2\text{Cl}$	-25	3.5
13. $\text{PhSeCH}(\text{CF}_3)\text{CH}_2\text{OCH}_3$	-49.2	1.7

a. negative sign is upfield from phenylvinyl selenide.

b. absolute value of coupling constants are given.

WASHINGTON  UNIVERSITY  
ST. LOUIS, MISSOURI 63130

September 20, 1982

DEPARTMENT OF CHEMISTRY  
Campus Box 1134

Dr. Bernard L. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station, TX 77843

Dear Barry:

We currently have a position available within the Departments of Chemistry and Medicine at Washington University for a postdoctoral research associate in the field of NMR of intact biological systems. I am soliciting applications and ask that you make this letter available to any qualified individuals whom you feel might be interested.

The primary focus of this position will be the hormonal regulation of liver metabolism as studied by H-1, C-13 and P-31 NMR experiments with rat liver in vivo and in vitro. The studies in vivo will make use of surface coil and chemical shift spin-imaging techniques while studies in vitro will utilize both perfused liver and isolated hepatocytes (liver cells). Time will also be available for independently motivated research within the context of our group interests.

The applicant should have a Ph.D. in chemistry, physics, biochemistry or other related field with extensive hands-on experience in magnetic resonance and a keen interest in biomedical applications. The initial appointment will be for a period of one year with additional extensions if mutually agreeable.

The NMR instrumentation available within our group for this project includes two state of the art Bruker NMR spectrometers, the WH-360 and CXP-200. Both spectrometers are fully multinuclear and employ widebore superconducting magnets of 72 mm and 85 mm inner diameter bores (inside room temperature shims) for the WH-360 and CXP-200 respectively. Extensive radio frequency design and test equipment is also available for NMR probe development and construction.

This position offers an exciting opportunity to gain extensive experience in the rapidly expanding field of intact tissue NMR while also becoming acquainted with the biomedical techniques needed to support such experiments. All interested applicants are invited to submit a letter detailing their background and interests along with a curriculum vitae; two letters of professional recommendation should also be submitted. Washington University is an Equal Opportunity, Affirmative Action Employer.

Sincerely,



Joseph J. H. Ackerman, Ph.D.  
Assistant Professor of Chemistry  
Research Assistant Professor of Medicine

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South San Francisco, California 94080  
(415) 952-1366

September 27, 1982

Professor Bernard L. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station, Texas 77843

University-Industry Collaboration; Positions Open

Dear Dr. Shapiro,

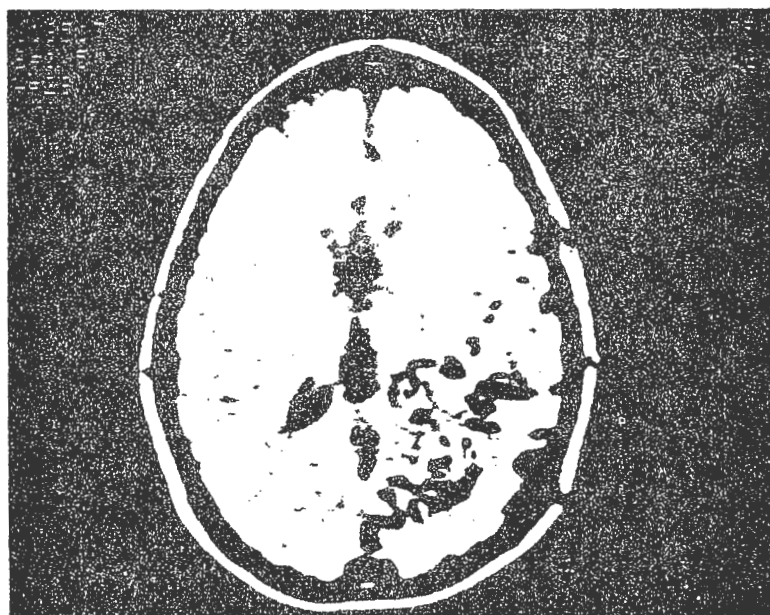
Since 1975 the Department of Radiology has had an NMR imaging program that has resulted in two imagers operating at 3.5 KGauss, one using a Varian 12-in pole tip magnet having a useful aperture of 6.5cm and a second incorporating an Oxford superconducting magnet and a useful aperture of 55cm. The small unit has been used to image rats, mice, gerbils, guinea pigs and phantoms that model disease conditions in humans (1,2) and the large unit has been used to evaluate the technology in volunteers and patients (3,4). Two examples of such images are shown below. An upgrading of this unit, as well as construction of additional ones, is now in progress. The University R & D program is supported by Disonics, a medical imaging company based in South San Francisco. The UCSF effort is a continuing program designed to improve hardware, develop imaging techniques, study their clinical effectiveness, and understand the reasons behind the tissue characteristics observed in images. The Disonics program involves the commercialization of this technology. We are interested in hearing from people interested in R & D or production responsibilities, either at the University or in Disonics. Positions are open for NMR spectroscopists, digital, RF or software engineers, or technologists. Resumes should be sent to our attention.

Larry E. Crooks, Ph.D.  
Associate Professor of Electrical  
Engineering  
Assistant Director, UCSF Radiologic  
Imaging Laboratory

Leon Kaufman, Ph.D.  
Professor of Physics and  
Director, UCSF Radiologic  
Imaging Laboratory

1. Herfkens R, Davis PL, Crooks LE, Kaufman L, Price DC, Miller RT, Margulis AR, Watts J, Hoenninger JC, Arakawa M and McRee R. NMR Imaging of the Abnormal Live Rat and Correlation with Tissue Characteristics. Radiology 141:211, 1981.

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NMR images of patients with a large liver tumor (bright central region in the top image), and an arteriovenous malformation (bottom image). Vasculature is seen dark because of flow. In the head image the dark regions of the brain demonstrate the abnormal vasculature.



STANFORD MAGNETIC RESONANCE LABORATORY  
STANFORD UNIVERSITY  
STANFORD, CALIFORNIA 94305

*Director: Oleg Jardetzky, M.D., Ph.D.  
Professor of Pharmacology*

(415) 497-6153  
(415) 497-4062

September 27, 1982

Professor B. L. Shapiro  
Department of Chemistry  
Texas A & M University  
College of Science  
College Station, TX 77843

Dear Barry,

Two positions at the Research Associate or Postdoctoral Fellow level will become available at SMRL in early 1983 as the laboratory inaugurates a new program with the installation of a 600 MHz spectrometer, which is currently on order. We are looking for highly competent, highly motivated individuals with an active interest in biological applications of NMR, willing to learn and to function as members of an interdisciplinary team in this rapidly growing field. Proficiency in NMR electronics, hardware and software and familiarity with probe design and construction are essential. Readiness to accept challenges and spearhead new developments highly desirable. Experience with in vivo NMR and imaging technology helpful, but not mandatory. Rank and salary dependent on qualifications. All inquiries should be addressed to me at the above address.

With best regards,

Yours sincerely,

  
Oleg Jardetzky

OJ:reh

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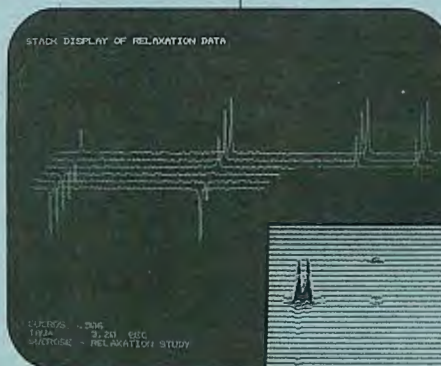
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