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Newsletter

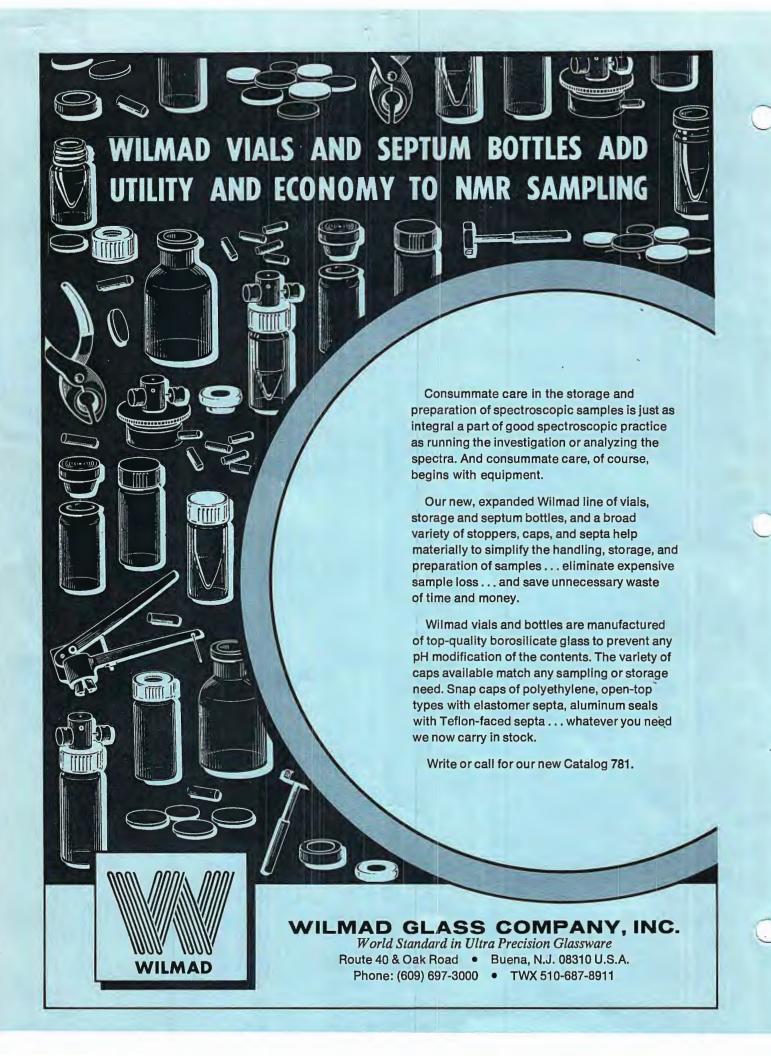
No. 263

August, 1980

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A monthly collection of informal private letters from Laboratories of NMR. Information contained herein is solely for the use of the reader. Quotation is <u>not</u> permitted, except by direct arrangement with the author of the letter, and the material quoted <u>must</u> be referred to as a "Private Communication". Reference to the TAMU NMR Newsletter by name in the open literature is strictly forbidden.

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All Newsletter Correspondence, Etc., Should be Addressed To:

Dr. Bernard L. Shapiro
Department of Chemistry
Texas A&M University
College Station, TX 77843 U.S.A.

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Professor B L Shapiro Texas A and M University College of Science College Station, Texas 77843

19 MAY 80

Dear Professor Shapiro

Investigation of Solutions of MnEDTA²⁻

Although ethylenediamine tetra acetate (EDTA) has found widespread use as a sequestrant, the structures of many of its complexes with metal ions in aqueous solution await confirmation. The MnEDTA2- complex is known to be seven co-ordinate in the solid state. We have carried out investigations using water proton relaxation times to ascertain the number of water molecules, n, co-ordinated to the metal ion in aqueous solution. Whilst this has been the major objective of this work, it has also been demonstrated that secondary sphere solvation i.e. water molecules hydrogen bound to complexed EDTA, plays an important part in determining the overall water relaxation times.

Contributions to overall relaxation rates from second solvation sphere water can be estimated experimentally by using larger, more flexible liquids than EDTA capable of octahedrally co-ordinating to $\mathrm{Mn^{2+}}$ or theoretically, using constants obtained from the crystal structure. Typical contributions to $\mathrm{T1^{-1}}$ due to secondary sphere water are $2\mathrm{s^{-1}}$ at 60 MH and 293K for an overall relaxation rate of $3.8\mathrm{s^{-1}}$.

Measurements of τ_r for the complexes have been obtained from frequency dependence of τ_1 -1,(FIG 1) a method which does not assume prior knowledge of D = $\frac{6}{15} \frac{S(s+1)}{r^6} \gamma_1 2g^2 \beta^2$.

Values of τ_r for Mn²⁺, MnEDTA²⁻, and MnEGTA²⁻ obtained are 4.2 x 10⁻¹¹s, 6.3 x 10⁻¹¹s, and 8.9 x 10⁻¹¹s, respectively. τ_s decreases from 1.5 x 10⁻⁸s to 1.67 x 10⁻⁹ at 60 MH_z upon complexation.

Values for n, calculated at various frequencies, range from 1.0 to 1.1, suggesting that the number of water molecules bound to Mn^{2+} in its complex with EDTA is one. It thus seems likely that the crystal structure of MnEDTA^{2-} retains its integrity in aqueous solution.

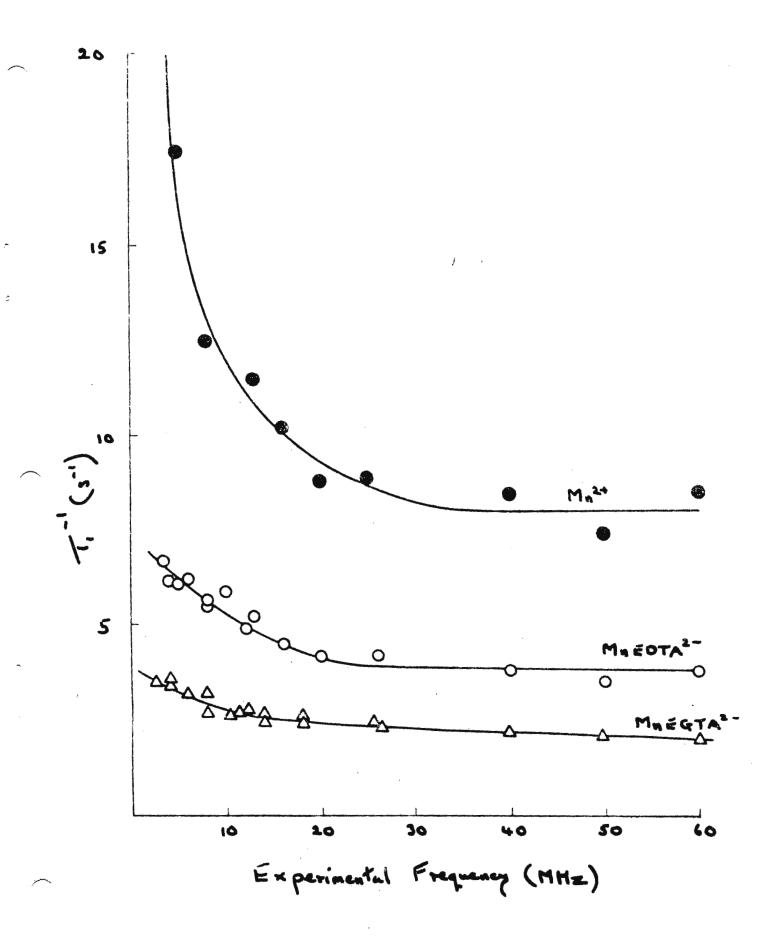
We apologize for the delay in sending this contribution.

Yours sincerely

E G SMITH

555mi4.

F13 1.



The University of Manitoba

Department of Chemistry Winnipeg, Manitoba Canada R3T 2N2



June 27, 1980

Dr. B. L. Shapiro
Department of Chemistry
Texas A & M University
College Station, Texas 77843
U.S.A.

Dear Dr. Shapiro:

Re: Tickling anywhere, particularly ¹⁹F.

We have recently made two modifications to our Bruker B-SV3 B decoupler that may be of interest to other owners of this unit.

The first modification removes the fixed frequency (31 P at 36.44 MHz in our case) 80 W power amplified and connects a jumper across the coax connectors in the "main frame". Because the 10 W driver amplifier is of broad-band design, the decoupler will now provide 10 to 12 W over the entire multinuclear range of our WH-90. The decoupling frequency is provided by the Shomandl synthesizer in the spectrometer. We have used this decoupler for a number of experiments requiring $^{19}{\rm F}$ decoupling and have found that 10 W is more than sufficient to decouple the one-bond carbonfluorine coupling in a CF3 group; provided that the decoupling frequency lies within about 100 Hz of the fluorine resonance. An external power amplifier may be added, if desired, but the effective range appears to be limited more by modulator bandwidth than by power available. The 90 MHz decoupler circuit in the Bruker probes may be tuned to 84.7 MHz with a slight adjustment of the "resonance" capacitor. To decouple $^{19}\mathrm{F}$ and $^{1}\mathrm{H}$ simultaneously, or to extend the range below 84.7 MHz, it would be necessary to build a matching network to double-tune the decoupler coil.

The other modification adds an external modulation jack and switch to the B-BMl modulator (figs. 1 and 2). This allows modulation of the decoupling frequency with an external signal or gating of the decoupler with a TTL logic pulse from the pulse programmer (NIC 293A).

con't

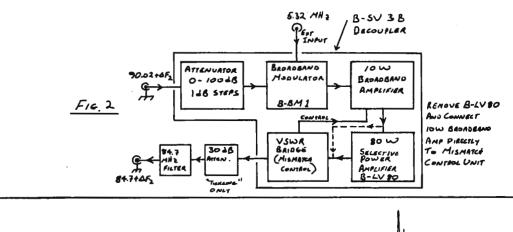
We have also used this input to mix the 90 + ΔF_2 MHz signal from the WH-90 proton decoupler with a 5.32 MHz signal from the Shomandl synthesizer to obtain 84.70 + ΔF_2 MHz, allowing computer-controlled-offset when decoupling or "tickling" fluorine. The advantage of this approach is that the offset-synthesizer has a much smaller step size than the Shomandl: 0.007 Hz compared to 1 Hz.

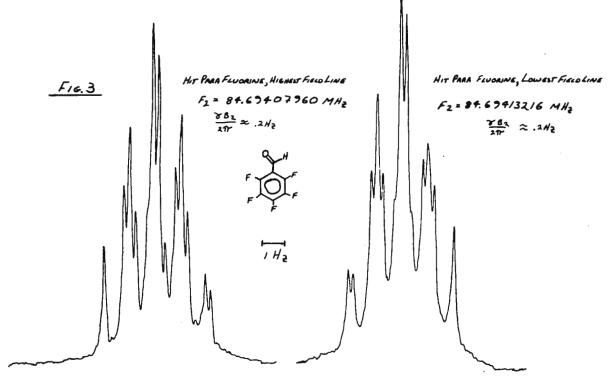
Figure 3 shows the results of a $^1{\rm H}$ - $\{^{19}{\rm F}\}$ double resonance experiment using this system.

Please credit this letter to Ted Schaefer's account.

Sincerely,

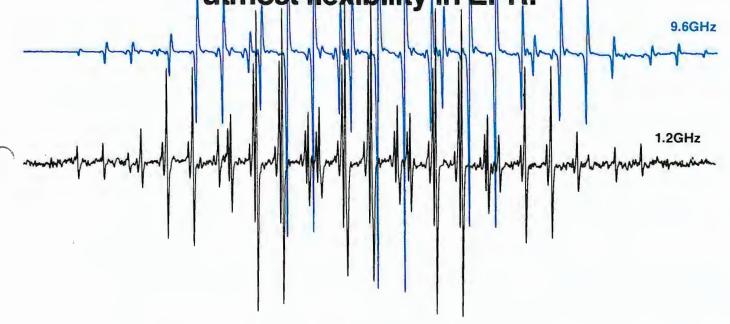
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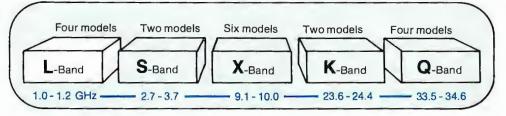


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Dr. A. Steigel Institut für Organische Chemie der Universität Düsseldorf

Direktor: Professor Dr. L. Birkofer

4 Düsseldorf, den June 24, 1979 Universitätsstraße 1 Fernsprecher 311-2298/99

Professor B.L. Shapiro Department of Chemistry Texas A and M University College Station, Texas 77843

Diastereotopicity as Probe to Lactam-Lactim Tautomerism

Dear Professor Shapiro,

Our previous 13 C NMR study (Newsletter 251-7) on the prototropic lactam-lactim tautomerism of $\underline{1}$ and $\underline{2}$ did not provide conclusive evidence for the preferred tautomeric structure. We now wish to show that information on the prototropic and silatropic tautomerism of $\underline{2}$ - $\underline{5}$ can be obtained by 1 H NMR using the methylene signals as structural probe.

The method is based on the finding that, in contrast to the fixed dilactam compound $\underline{6}$, the methylene protons are diastereotopic in the fixed lactam-lactim compounds $\underline{7} - \underline{9}$. The shift difference of 0.08 - 0.09 ppm in the latter compounds can be explained by the different anisotropy of the carbonyl and OR^1 groups.

In view of this effect, the observation of isochronous methylene protons in the potentially tautomeric compound $\underline{2}$ indicates the predominance of structure $\underline{2a}$. However, in the case of $\underline{1}$ the isochrony of the methylene protons does not allow a discrimination between the presence of structure $\underline{1a}$ and a fast tautomerization in structure $\underline{1b}$. On the other hand, the methylene protons of the trimethylsilyl derivatives $\underline{3}$ - $\underline{5}$ are diastereotopic (shift difference 0.08-0.10 ppm) showing the presence of the lactam-lactim structures 3b-5b.

Yours sincerely,

Alaia Staigal

Alois Steigel

UNIVERSITY OF CALIFORNIA, DAVIS

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SANTA BARBARA • SANTA CRUZ

DEPARTMENT OF CHEMISTRY

DAVIS, CALIFORNIA 95616 July 8, 1980

Dr. Bernard L. Shapiro Department of Chemistry Texas A&M University College Station, Texas 77843

Isotopomer Identification Through SPT Experiments

Dear Dr. Shapiro:

Selective population transfer (SPT) experiments in FT NMR are normally accomplished by selective inversion of a single transition followed by a non-selective observation pulse. Under appropriate conditions which include a small flip angle observation pulse, only transitions directly connected to the inverted transition are altered in intensity. Subtraction of this spectrum from an unperturbed spectrum results in a difference mode display bearing a close resemblance to an INDOR response in CW NMR. Utilization of a larger flip angle can result in intensity alterations of all transitions connected to the inverted transition. The use of SPT experiments for elucidation of carbon-proton as well as proton-proton spin-spin couplings has been aptly demonstrated 1,2 . Here we report the use of SPT experiments in ^{31}P NMR to make isotopomer identifications and enable determinations of ^{31}P - ^{31}P and ^{31}P - ^{195}P t couplings. The isotopomers of $Pt_2(\mu-S)(CO)(PPh_3)_3$, arising because of the 34% natural abundance of 195 Pt (the only Pt isotope possessing a magnetic moment), are shown at the top of Figure 1 (Pt* indicates 195 Pt). The resulting 31 P {H} NMR spectrum (spectrum A) shown in Figure 1 is complicated because of contributions from each of the isotopomers. However, selective inversion of one of the stronger lines gives rise to the difference mode spectrum (spectrum B) which displays just the (nearly) ABX splitting of the unlabeled isotopomer. Further SPT experiments were used to identify the spectra of all but the doubly labeled isotopomer. A more complete description of the experimental parameters along with the $^{31}P-^{31}P$ and $^{31}P-^{195}Pt$ couplings obtained by spectral simulation will be presented in a forthcoming publication. We note that the ability of the Nicolet NT-200 spectrometer (on which the spectra were obtained) to switch from high to low power under computer control made implementation of the experiment a trivial matter. Please credit this contribution to the account of Prof. G.N. La Mar.

Sincerely,

A.L. Balch Professor

Department of Chemistry

G.B. Matson

Operations Manager UCD NMR Facility

C.H. Lindsay

Research Assistant

References

- 1) R. Sorensen, R.S. Hansen and H.J. Jakobsen, J. Magn. Reson. <u>14</u>, 243 (1974); A.A. Chalmers, K.G.R. Pachler and P.L. Wessels, Org. Magn. Reson. 6, 445 (1974).
- 2) K. Kushida, K. Aoki and S. Satoh, J. Am. Chem. Soc. 97, 443 (1975); K. Bock, R. Burton and L.D. Hall, Can. J. Chem. <u>54</u>, 3526 (1976).

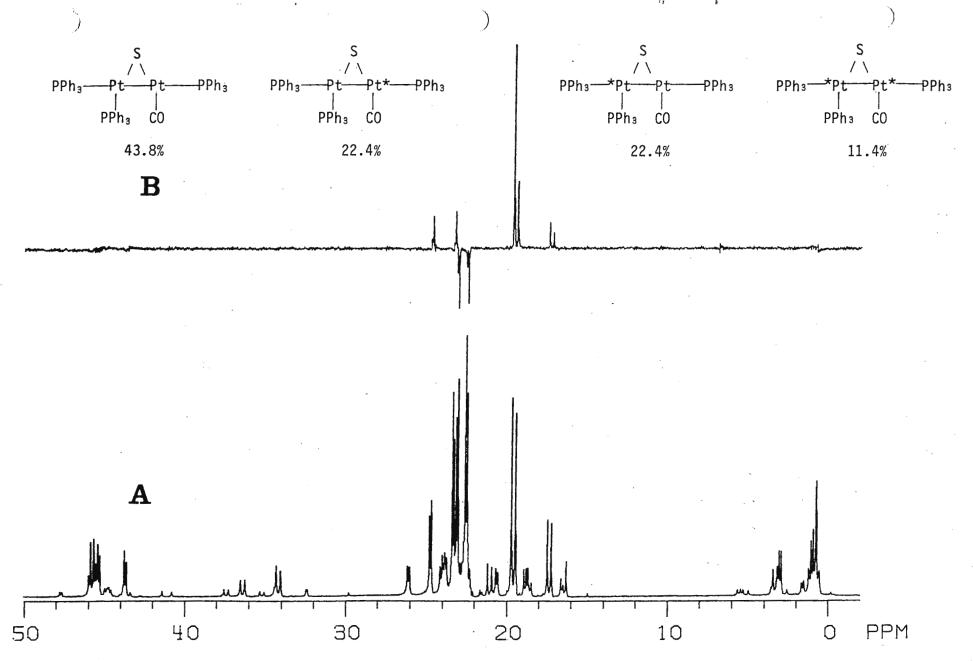


Figure 1. The isotopomers of $Pt_2(\mu-S)(CO)(PPh_3)_3$, arising because of the 34% natural abundance of ^{195}Pt , are shown at the top of the figure. Spectrum A shows the normal $^{31}P\{^1H\}$ spectrum, while spectrum B displays the difference mode spectrum resulting from inversion of the strong line just under 20 ppm. Spectrum B clearly identifies the (nearly) ABX pattern resulting from $^{31}P-^{31}P$ coupling in the unlabeled isotopomer.



The Ohio State University

Department of Chemistry

140 West 18th Avenue Columbus, Ohio 43210 Phone 614 422-2251

July 9, 1980

Dr. B.L. Shapiro
Department of Chemistry
Texas A and M University
College Station, Texas 77843

¹³C Shifts in Benzenium Ions

Dear Barry:

In response to the dreaded pink note, some thoughts on the alleged charge shift relationship for π carbons in carbenium ions. We recently had occasion to examine ^{13}C shifts in benzenium ions. The shifts vary widely with substituents solvent and temperature, yet a satisfying regularity ensues from the following treatment.

Let's assume these shifts have contributions from a) the carbon skeleton, b) a medium effect which acts uniformally at all carbons and what's left is c) due to charge. For a) we choose ^{13}C shifts in neutral model compounds in skeletal environments which most closely resemble those for carbons in the carbenium ions. Once the neutral model shifts, δ_{m} , have been subtracted from the carbenium shifts, δ^+ , the shifts which remain are internally almost the same for each R $^+$ but are displaced from one R $^+$ species to another. This displacement turns out to be equal to δ_3^+ - δ_{M_3} (neutral C₃). If we take the sum for all the ring carbons

$$\sum_{C_1}^{C_6} \left[\left(\delta_{i}^{+} - \delta_{mi} \right) - \left(\delta_{3}^{+} - \delta_{m_3} \right) \right]$$

it comes close to 166 ppm each time, almost the charge shift index. In other words, the introduction of a charge of +1 into the neutral model system results in a total 166 ppm downfield shift provided we subtract $\delta^+ - \delta$ from all the shifts. The latter must represent an environmental effect from solvent (for instance electric field) that acts at all carbons equally.

One could argue that second order differences cancel out all the interesting effects. However the treatment does lead to a common number \circ 166 ppm/e-. Also, it seems we have to include the shift of the saturated ring carbon.

Examples with model structures and results follow below.

Best wishes,

Gidem

Gideon Fraenkel Professor of Chemistry

Σ[(δ _i ⁺	$-\delta_{Mi}) - (\delta_{s}^{+} - \delta_{M3})]$
I CF ₃ CO ₂ in CF ₃ CO ₂ H	165.5 ppm
I AlCl4 in CH2Cl2	167.4
II FSO in SbF5	167.0
III + Al ₂ Cl ₇ in CH ₂ Cl ₂	165.2

LABORATOIRE DE CHIMIE THEORIQUE

Professor B.L. SHAPIRO TEXAS A&M University College of Sciences

COLLEGE STATION, Texas 77483 U.S.A.

Title: writing with an analog plotter and a Nicolet 1180 computer

Nancy, July 12, 1980

Dear Professor Shapiro

Your yellow remainder arrived during the examination period and since all undergraduate students are now on vacation, we have the feeling it is time to answer. Anyway, we are afraid we have lost at least one issue of your newsletters.

For meany reasons (practical as well as esthetic), we were eager to make our Nicolet 1180 computer draw axis or write comments on a spectrum sheet. This appeared, at first, impossible since we could not afford a digital plotter and since, regarding analog devices, the Nicolet Basic software just allows to plot a curve. We therefore decided to adapt the digital plotter software to our situation.

Basically, it consists in replacing each instruction specific to the digital plotter by a set of instructions which allows to drive the analog plotter (SEFRAM TGM 101 in our case) in a proper way. This is essentially achieve by controlling the speed of the pen according to direction changes. As an example, an horizontal straight line will be drawn quickly whereas the pen will move slowly when a curve is to be plotted. This is done in order to get rid of the inherent inaccuracy of an analogic plotter. All the required modifications are included in a dedicated routine (STDPL) which is simply called just vefore PLOTS (Nicolet initialization plotting routine). This routine has to be loaded by the linking loader at the basic program generation stage.

In order to obtain a good quality of presentation, some minor adjustments on the plotter may be necessary (increase of the sensitivity and decrease of the time constant). The joined figure has been obtained in less than 5 minutes. The program is available on request.

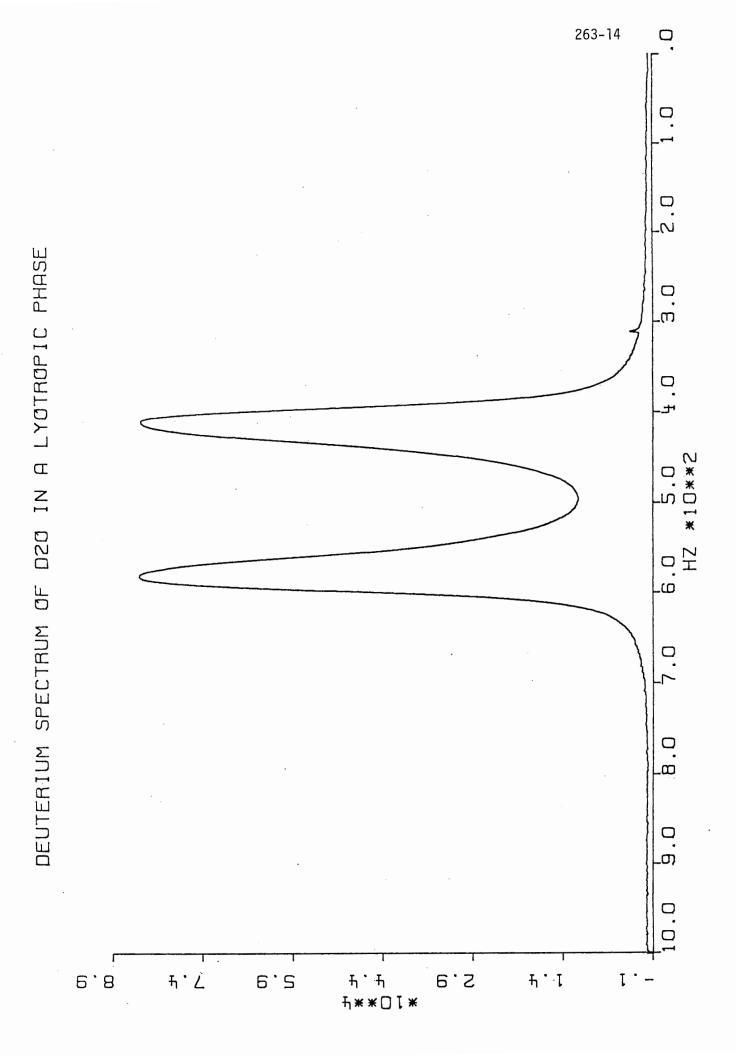
Sincerely yours

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July 17, 1980

NATIONAL INSTITUTE OF ENVIRONMENTAL HEALTH SCIENCES P.O. BOX 12233 RESEARCH TRIANGLE PARK, N.C. 27709

Professor B.L. Shapiro Department of Chemistry Texas A and M University College Station, Texas 77843

Dear Barry:

Glutathione Conjugates of Epoxides

One of the major biotransformation pathways for epoxides is conjugation with glutathione (GSH). Further metabolism of the GSH conjugates gives rise to the mercapturic acids (N-acetylcysteine conjugates). We have been interested in the stereochemical aspects of this important detoxication reaction because of the influence of stereochemistry on the metabolism and/or excretion of GSH conjugates.

Studies of the ^{13}C NMR spectra of the glutathione and mercapturic acid conjugates of styrene oxide, phenanthrene 9,10-oxide and pyrene 4,5-oxide show that the peak separation (0.1-0.4 ppm) between diastereoisomers for the carbons which were formerly the epoxide ring (CHOHCHSR-) is sufficient to allow quantitation of the mixture of isomers. We have recently assigned the relative stereochemistry of the GSH and mercapturic acid conjugates of styrene oxide (four isomers each, two positional isomers as a mixture of diastereoisomers) by preparing the conjugates from (+)- and (-)-styrene oxide.

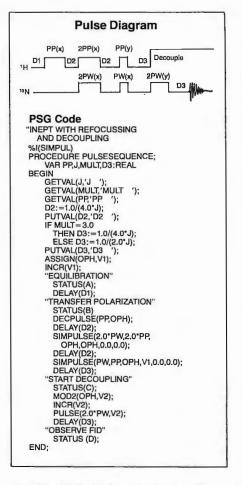
In related work, ¹³C NMR analysis of the GSH conjugates formed from (+) benzo(a)pyrene 4,5-oxide-4,5-¹³C by a purified glutathione transferase from little skate (<u>Raja erinacea</u>) liver demonstrated that equivalent amounts of the positional isomers (4-hydroxy-5-glutathionyl and 5-hydroxy 4-glutathionyl) were formed as single diastereoisomers. This was confirmed by comparison with the spectrum of the synthetically produced isomers (mixture of four stereoisomers) and HPLC. Whether this stereochemical behavior is restricted to the purified fish liver enzyme, the substrate, or this particular combination is actively being pursued by the use of other arene oxide substrates and with purified glutathione transferases from other sources.

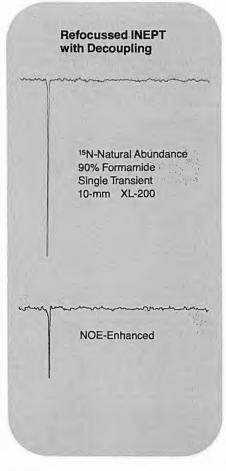
Manuscripts of this work are in preparation and I will be happy to send preprints to anyone interested.

Sincerely,

Richard H. Cox, Ph.D.

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Illustrated here is a simple implementation of these ideas.

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- **ASSIGN** INCR RND MOD4 · Ability to specify and vary phase and receiver off-times dynamically
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- · Use of math statements for sequence timing calculations

- Complete separation of sequence code from parameter sets
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- Use of indirect parameter labels in sequence code
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Example sequences

Standard two-pulse Carr-Purcell-Meiboom-Gill T2

Quadrupole echo Cross-polarization

Miltiple-contact cross-polarization

Selective excitation Quadrature selective excitation

INEPT with refocussing and decoupling

J-Cross polarization

Refocussed J-cross polarization Noise off-resonance spin echo Inversion-recovery spin echo Multiple quantum 2D

Proton-carbon correlated 2D Heteronuclear enhanced 2D Double quantum 13C-13C spectroscopy

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- Flicker-free TV display with graphics capability
- 13-bit ADC

Accessories

- 19F transmitter
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- Maintenance kit
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Data System

- PASCAL language
- State-of-the-art operating system
- Disk-based using modular design software concept
- Concurrent and sequential PASCAL
- Floating-point data and math format
- Multitasking-simultaneous acquire, plot, print, display parameter entry
- Queuing of acquisitions, plots, prints and calculations
- Spooling of plots and prints
- Disk-resident data tables
- Separate FID and spectral storage
- System resident PASCAL compiler for user programming
- User access to data files

- · Expandable user-defined command and parameter architecture
- Floating-point or integer transform
- Convolution difference/gaussian apodization functions
- Parameter set libraries
- 2D transform
- Plot graphics
- T1, T2, 3-parameter least-squares-fit analysis programs
- Spin simulation
- LAOCOON with magnetic equivalence
- User-definable disk libraries
- PASCAL system source code availability
- NOE calculation
- Add-subtract-convolution spectral manipulation



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DEPARTMENT OF HEALTH, EDUCATION, AND WELFARE

PUBLIC HEALTH SERVICE
NATIONAL INSTITUTES OF HEALTH
BETHESDA, MARYLAND 20205
July 22, 1980

Professor Barry Shapiro Department of Chemistry Texas A & M University College Station, Texas 77843

Dear Barry:

We have recently been studying molecular motions in a series of hormone peptides and trying to understand the relationship between conformational constraints and biological activity. In the course of this work we have carried out a temperature and magnetic field strength dependence of the ^{13}C T_1 values of a Bradykinin. The data, are summarized below for each of the backbone \upsigma -carbon atoms.

	T =	14 ⁰	T = 3	38 ⁰
	15.1MHz	67.9MHz	15.1MHz	67.9MHz
Arg	.102	.212	.255	.344
Pro	.095	.183	.199	.301
Pro	.082	.195	.203	.283
G1y	.106	.214	.254	.340
Phe	.080	.167	.185	.250
Ser	.084	.176	.190	.249
Pro	.082	.173	.203	.255
pNO ₂ Phe	.078	.169	.180	.242
pNO ₂ Phe Arg	.083	.207	.210	.327

It is necessary to deterime the field strength dependence of the $\rm T_1$ values in order to correctly characterize the motional behavior in terms of the relevant rotational correlation times. We have analyzed these data using a model which assumes isotropic overall rotation with independent internal correlation times. The overall correlation time ranges from 4×10^{-8} sec at $14^{\circ}\mathrm{C}$ to 7×10^{-10} sec at 38° whereas the internal correlation times are around 1×10^{-10} and change by no more than a factor of two over the same temperature range. This analysis yields computed $\rm T_1$ values within experimental error of the measured values and also correctly predicts the observed NOE factors.

Please credit this contribution to the subscription of Dr. R. J. Highet.

Sincerely yours,

James A Ferretti



University of Edinburgh

Department of Chemistry

West Mains Road, Edinburgh, EH9 3JJ Scotland.

Yr. ref.: Our ref.: Telex 727442 UNIVED G Telephone 031 - 667 1081 Ext. 3416

30.6.80

Professor B.L. Shapiro, Department of Chemistry, Texas A & M University, College Station, Texas 77843, USA

Dear Professor Shapiro,

XL-100 Program Patch

Here is yet another modification to the old XL-100 programme, 994100-E and -D. It enables a plotter speed change while plotting, by typing S for slower and F for faster. The PT command is eliminated. The DC command is also rationalised. Typing DC now clears the drift correction and jumps to the scope routine without needing any more input.

With apologies for the brevity of this note.

Yours sincerely,

B

Alan Boyd

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University of Illinois at Urbana-Champaign

School of Chemical Sciences

July 14, 1980

Professor B. L. Shapiro Department of Chemistry Texas A & M University College Station, Texas 77843

RE: 1) NMR and ESR Studies of Thylakoid Membranes

2) Postdoctoral Opening

Dear Barry:

This is a note to update the research my group (S. Rajan and I) have been doing with some of the local biophysicists (Govindjee and Rita Khanna) on photosystem II, the oxygen-evolving part of photosynthesis in green plants.

It has been proposed that manganese is an integral part of photosystem II. In an effort to better understand the nature and role of the manganese we have made parallel NMR and ESR measurements of thylakoid membranes in aqueous suspension under a variety of conditions. Also, total Mn content was determined by neutron activation and oxygen evolution monitored. In other earlier work two pools of bound Mn have been identified, a loosely-bound pool related to 0_2 evolution and a tightly-bound pool. Our experiments indicate that the loosely-bound pool has at least two components of which the most loosely-bound is not related to 0_2 evolution. Also, it appears that the tightly-bound pool is associated with the light harvesting complex.

The aqueous proton relaxation rate $(R_2 = 1/T_2)$ is known to be sensitive to the amount and oxidation state of Mn present and to the extent and nature of its binding in a complex. Also, changes in binding can affect the correlation times that govern R_2 or change the accessibility of Mn to the aqueous protons. In ESR, free aqueous Mn(II) gives a distinctive six-line spectrum while other oxidation states and bound Mn(II) give at most very broad, weak absorption. The two types of measurement give confirmatory and supplementary evidence of changes in oxidation state and binding of Mn.

Among our findings for pea thylakoids are the following:

- (a) Aging of thylakoids at 35 C causes a parallel decrease in θ_2 evolution activity, in R_2 and in Mn content, confirming that R_2 monitors bound Mn related to θ_2 evolution.
- (b) Addition of 1 to 20 mM MgCl $_2$ causes a decrease in R $_2$ and an increase in the six-line ESR spectrum for free Mn(II), without any effect on 0 $_2$ evolution activity, indicating the presence of a pool of non-functional, loosely-bound Mn.
- (c) The decrease of R_2 by MgCl₂ is probably due mainly to the smaller molar relaxivity of free compared with bound Mn(II). Mg ion is known to cause structural changes in membranes, (e.g., grana stacking) but not in trypsinated membranes. However, MgCl₂ has the same effects upon the R_2 and ESR of trypsinated membranes.
- (d) The isolated light harvesting Chl <u>a</u>/Chl <u>b</u> preparation (LHC) contains $^{\circ}$ 1/3 of the bound Mn in the thylakoids; this may be the tightly-bound

Professor B. L. Shapiro

Mn pool.

- (e) Treatment of thylakoids with NH₂OH increases R₂ by up to nearly two-fold, presumably by the reduction of higher oxidation states of Mn to Mn(II). Also, the progressive release of bound Mn with [NH₂OH] is shown by the growth of the six-line Mn(II) ESR spectrum, by the progressive loss of O₂ evolution and by the decrease in R₂ at [NH₂OH] \gtrsim 1 mM. However, the release of Mn is not complete even at 100 mM NH₂OH, confirming the presence of a tightly-bound pool. H₂O₂ also increases R₂.
- (f) Reduction of thylakoids with tetraphenyl boron (TPB) causes R_2 to nearly double. The titration curve exhibits three sharp endpoints, the first at [TPB] ~ 2.5 mM corresponding to complete inhibition of 0_2 evolution. The endpoints may correspond to the reduction of Mn to Mn(II) in each of three pools.

Further work is continuing along these lines. In particular, we hope to characterize in more detail the species involved in the titration curves, for example, by removing some of the Mn and seeing the effect upon the titration.

I have a postdoctoral opening for someone interested in such studies. At least general familiarity with pulsed-NMR instrumentation would be helpful. If you're interested or know of someone who might be, please give me a call (217/333 0710) or send me a note, preferably with a curriculum vitae.

Sincerely yours,

Professor of Chemistry

akh

HOWARD UNIVERSITY



WASHINGTON, D. C. 20059

DEPARTMENT OF CHEMISTRY

July 16, 1980

Professor Bernard L. Shapiro Department of Chemistry Texas A & M University College Station, TX 77842

Dear Barry:

Scatchard plots are often used in the analysis of NMR data (1, 2). We have been examining the exchange dynamics of tetraphenylporphyrinato Zn(II) with pyridine and N-methylimidazole at 200 MHz and have found a version of a Scatchard plot most useful in assessing the values of several parameters needed in the analysis of the DNMR. The reaction considered is ZnTPP + L = ZnTPPL, which is in rapid exchange at room temperature with $\boldsymbol{v}_{o} = X_{f} \boldsymbol{v}_{f} + X_{b} \boldsymbol{v}_{b}$. If one makes up a sample of ZnTPP and ligand in CDCl3 and records the spectrum at several different dilutions the ligand frequencies will shift down field due to progressive dissociation of the complex. A plot of $\Delta_i/(ZnTPP)_T$ ($\Delta_i = \delta_f - \delta_0$) vs. Δ_i gives a straight line with the extrapolation to the Δ_i axis giving γ_0 for the fully formed complex in rapid exchange at that particular (ZnTPP)/L ratio (Fig. 1). From this one can easily determine the mole fraction free and bound and the equilibrium constant under a variety of conditions. A plot of these intercepts (Δ_i^0) vs. (ZnTPP)/L ratio gives a straight line with a sharp break at (ZnTPP)/L = 1 (Fig. 2). For pyridine the $Z_nTPP/L > 1$ still has a slope, for N-methylimidazole it appears to have a slope of zero. We believe that the low slope for pyridine in the (ZnTPP)/L > 1 region is due to π complexing and we use that slope to correct our other observed values for the T complexing. The point of the break at (ZnTPP)/L = 1 permits an accurate assesment of 8f - 8b.

1. H. Stamm, W. Lamberty and J. Stafe, J. Am. Chem. Soc., <u>102</u>, 1529 (1980).

2. N. K. Wilson, J. Phys. Chem., 83, 2649 (1979).

A. H. Turner

C. B. Storm

Sincerely yours,

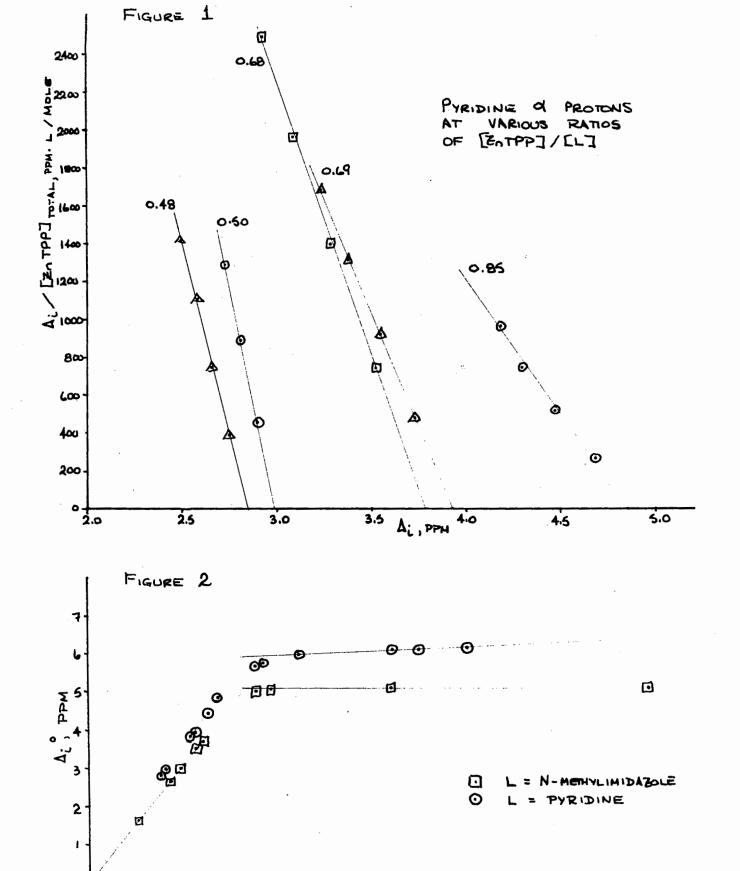
M. B. Swann

3.5

3,0

2.5

4.0



1.5 2.0 [EnTPP]/[L]

2.0

0,5

1.0



ABO AKADEMI FINLAND

July 17, 1980

Relativity, lone pairs and spin-spin coupling

Dear Professor Shapiro,

We have recently written a program which calculates nuclear spin-spin coupling tensors using the "Relativistically Parameterized Extended Hückel Method (REX)" /1/ and the relativistic equivalent of Ramsey's non-relativistic theory /2/. All the parameters are taken from ab initio atomic calculations, relativistic or non-relativistic, thus permitting an analysis of relativistic effects. Any element can be treated. The largest molecule we have looked at so far is $Pb_2(CH_3)_6$.

The trends when descending down a column of the periodic system are shown on the following page. These trends are quite typical for the cases with and without lone pairs, respectively, as discussed qualitatively earlier by several workers (Kennedy&McFarlane, Birchall& Pereira, etc.). A new piece of insight is that relativistic effects seem to play a rôle in making 1K negative , already for elements as light as Se or Te. The relativistic increase of $^1K(XH)$ in case (b) is again beautifully confirmed.

We still keep finding that relativistic effects increase the anisotropy $\Delta K = K_{\parallel} - K_{\perp}$, as predicted in /2/, and have tracked down this to an s(A)-p_{1/2}(B) contribution, for which $K_{\parallel} = -K_{\perp}$.

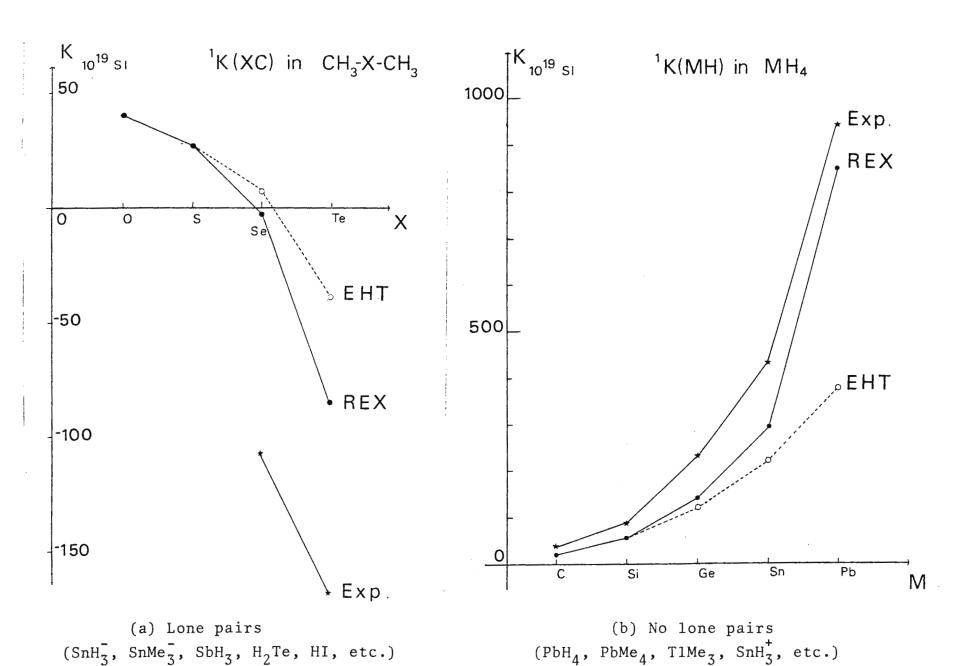
Sincerely yours,

Pekka PYYKKÖ

Laurent WIESENFELD

/1/ L.L. Lohr, Jr. and P. Pyykkö, Chem. Phys. Letters 62 (1979) 333; L.L. Lohr, Jr., M. Hotokka and P. Pyykkö, QCPE 12 (1980) 387.

/2/ P. Pyykkö, Chem. Phys. 22 (1977) 289.



263-26



HALL-ATWATER LABORATORIES MIDDLETOWN, CONNECTICUT 06457

TEL.: (203) 347-9411

DEPARTMENT OF CHEMISTRY

July 18, 1980

Professor B. L. Shapiro Department of Chemistry Texas A & M University College Station, Texas 77843

Dear Dr. Shapiro,

One of the many goals of the spectroscopist is to obtain good spectra from lousy samples. For those of us in NMR the main sources of poor spectra (besides malfunctioning spectrometers and operators) are either field and/or sample inhomogeneity. Recently people have started putting a whole new class of inhomogeneous samples into NMR tubes: intact cells and tissues. As a distinguished colleague has said: "These samples are inherently not isotropic and it will be years until those genetic engineers get any isotropic organisms for us to study." In the meantime it is of interest to find ways in getting the information which is obscured by the poor quality of living samples. One approach is to use DANTE2 to selectively excite a magnetically homogeneous portion of the sample while proton decoupling is applied. If the FID is then acquired without proton decoupling the proton coupled spectrum from a magnetically homogeneous sample can be obtained. The ability to resolve proton couplings might offer information that could aid in assigning the 31P peaks as well as obtaining some conformational information about cellular phosphates in vivo. An example is shown in the figure. The bottom spectrum is of 2' CMP in an inhomogeneous field. If a DANTE pulse train is applied to excite the center of the ³ P peak and then a non-selective pulse applied to the 31P nuclei the middle spectrum results. This illustrates the width of the selective excitation relative to the inhomogeneous linewidth. If the sample is excited only by the selective pulse train in the presence of proton decoupling and then the FID acquired without proton decoupling then the top spectrum is obtained. The proton-phosphorus coupling is about 7 Hz. The method has fairly good signal to noise and the spectra shown were all obtained by accumulating 4 transients. The method has not been tested on any real live samples yet.

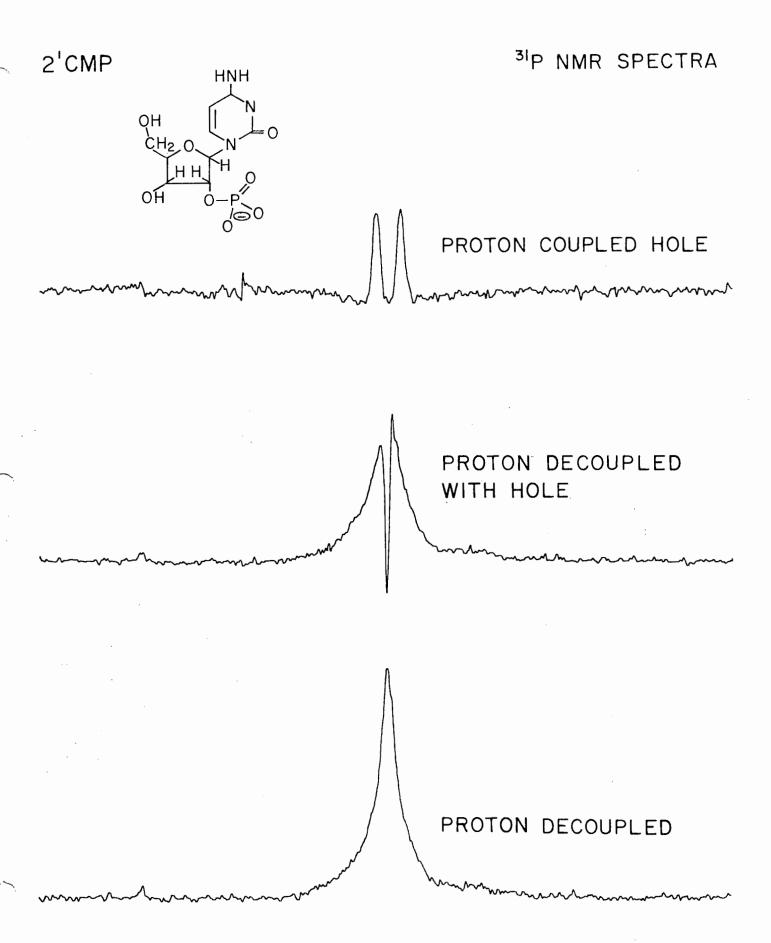
1. B.A.Still, quoted in "The Middletown News," July 14, 1980, p.31.

2. G.Morris and R.Freeman, J.Magn.Res., 29, 433-462 (1978).

Sincerely,

Philip H. Bolton Assistant Professor

PHB:1b





Oklahoma State University

Department of Chemistry / (405) 624-5920 / Stillwater, Oklahoma 74074

July 21, 1980

Professor B. L. Shapiro Department of Chemistry Texas A&M University College Station, Texas 77843

Dear Professor Shapiro:

13C Relaxation in Crosslinked Polystyrene Gels

Interest in determining the role of crosslinked polystyrene structure in polystyrene-supported catalysts led us to investigate chain mobility in these crosslinked, solvent-swollen gels. The polymers contain varied amounts of divinylbenzene as crosslinker and 25 wt % chloromethylstyrenes to be used for binding catalysts. Spectra were obtained at 25.2 MHz on our Varian XL-100(15)-Nicolet TT-100 system at 30°C with the polymers swollen or dissolved in CDCl $_3$.

Representative data are in the table. The T₁s decrease with crosslinking up to 4% divinylbenzene and may increase slightly with 6% divinylbenzene, suggesting that the T₁ minimum occurs at ~4% crosslinking. The linewidths increase with crosslinking from values nearly the same as those of soluble polymers at $\leq 1\%$ crosslinking to broad bands at 10% crosslinking. The linewidths depend on crosslinking, not on polymer concentration, because the uncrosslinked polymer has identical linewidths in 10, 19, and 25% solutions in spite of a tremendous increase in viscosity. NOE ratios, determined by gated decoupling and fully decoupling successive spectra, decrease as crosslinking increases. The most interesting data arose from the discovery in NOE measurements that the aliphatic carbon signal areas in polystyrenes were always smaller than theoretical values when compared with the aromatic signal areas. On a per carbon nucleus basis the aliphatic signal decreases from ~87% of its theoretical area in uncrosslinked samples to ~20% in the 10% crosslinked sample when compared with a poly(ethylene glycol) internal standard. The relative signal areas were obtained from both gated decoupled and fully coupled spectra, and by both electronic and cut-and-weigh integrations. Results using all of these methods agree to within 10% in all but one sample. We attribute the signal area decrease to increasing numbers of dipolar coupled carbon nuclei in the polymer backbone as the crosslinking increases. Our results are the first systematic study of crosslinking level on 13C relaxation in polymer gels of which we are aware.

Please credit this contribution to the Oklahoma State University account.

Sincerely, Warren T. Ford

Warren T. Ford

Associate Professor

								Signal Area Ratios from Gated Decoupled Spectra				
Sample ^a	Wt % Polymer in CHCl ₃	<u>o,m</u>	Methine	- <u>C</u> H ₂ C1	—Lin <u>o,m</u>	ewid <u>P</u>	lth, Hz— Methine	Aromatic PEG	Aliphatic PEG	Aliphatic Aromatic		
Polystyrene	10	124	95		20	14	15	0.98	0.82	0.89		
Uncrosslinked copolymer	10				20.5	14	15			0.87		
	19				21.5	15	15	₩ ₩	→	0.84		
	25	123	97	141	21	15	15.5			0.87		
0.5% DVB	11				25.5	20	14	0.96	0.84	0.88		
1.0% DVB	10	112	93	155	40		16	1.12	0.86	0.77		
2.0% DVB	17	100	72	132	95		22.5	1.06	0.76	0.72		
4.0% DVB	26	93	73	104	180		36	0.93	0.50	0.54		
6.0% DVB	30	113	78	110	240		~65	1.04	0.45	0.43		
10.0% DVB	33			~ =	~470			0.48	0.20	0.42		

1

GBF Mascheroder Weg 1 D-3300 Braunschweig-Stöckheim



Gesellschaft für Biotechnologische Forschung mbH

Abteilung

Physikalische Meßtechnik

Ihre Nachricht vom

Texas 77843,

USA

College Station,

Telefon 05 31/70 08-1

Telefondurchwahl 0531/7008 362

Datum 21.7.80

Dear Professor Shapiro,

Professor B.L. Shapiro,

Department of Chemistry,

Texas A & M University,

One bond J(CC)

The wealth of data that now exists in the literature for J(CC) has been tabulated in two recent reviews 1,2, where all the data up to the beginning of 1980 have been collated. By far the most numerous are those for coupling over one bond. These have been summarised in the enclosed scheme which we hope will be of practical use for those chemists and biochemists working in this area of NMR.

The scheme shows the range and distribution of \$^1J(CC)\$ values, abscissa, according to the formal hybridisation of the coupled carbons which is indicated along the left-hand ordinate as follows:

A-both coupled carbons formally with sp\$^3 hybridisation, B-one coupled carbon formally with sp\$^3 hybridisation and the other with sp\$^2 hybridisation, C-one coupled carbon formally with sp\$^3 hybridisation and the other is a carbon of a carbonyl function, D-one coupled carbon formally with sp\$^3 hybridisation and the other with sp hybridisation, E-both coupled carbons formally with sp\$^2 hybridisation (other than aromatic and carbonyl carbons),

F-both coupled carbons formally with sp\$^2 hybridisation (both aromatic or aromatic and olefinic carbons), G-both coupled carbons formally with sp\$^2 hybridisation, one of which is a carbonyl function, H-one coupled carbon formally with sp\$^2 hybridisation, and I-both coupled carbons formally with sp hybridisation.

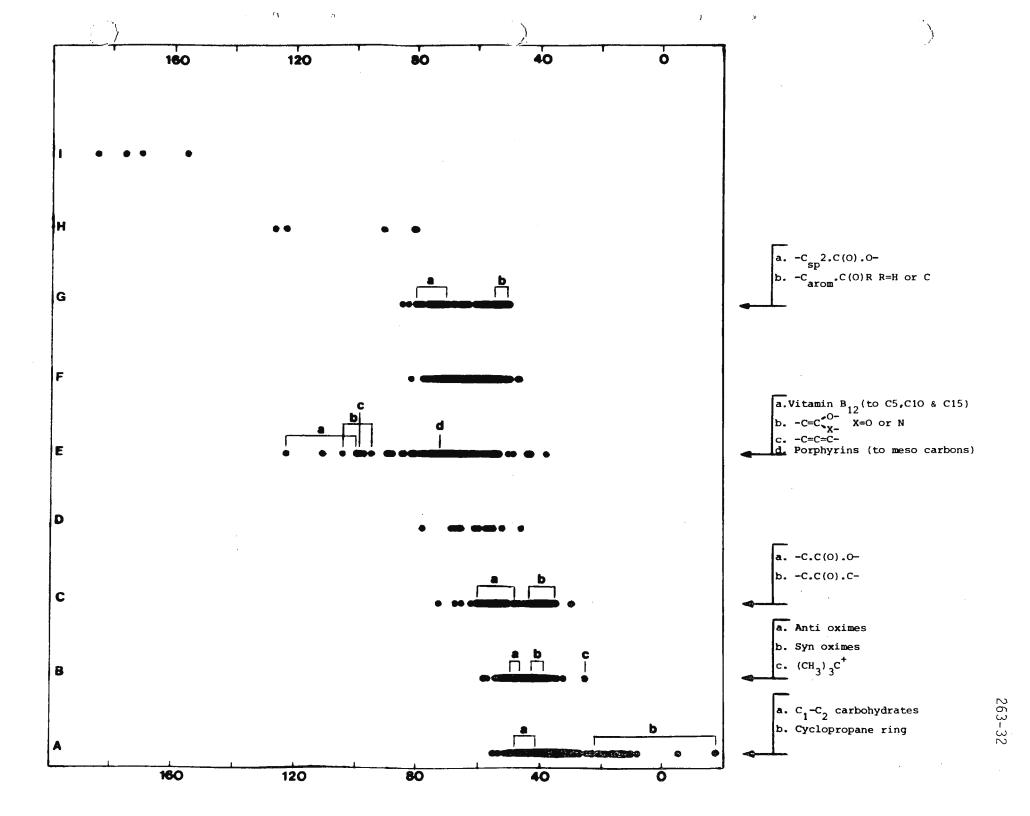
This work was done in collaboration with Dr. P.E. Hansen of Roskilde University, Denmark.

Yours sincerely,

Victor Wray

1.V.Wray, Prog.Nucl.Magn.Res.Spectrosc., 13, 177 (1979). 2.V.Wray and

P.E. Hansen appendix to P.E. Hansen, Annual Reports NMR Spectrosc., in press.



UNIVERSITÄT TÜBINGEN PHYSIKALISCHES INSTITUT Prof. Dr. O. Lutz D-7400 TÜBINGEN 1, den 22.07.1980 Morgenstelle Telefon (0 70 71) 29 67 14

Professor B.B. Shapiro
Department of Chemistry Texas A & M University
College Station
Texas 77843 U.S.A.

Hetero-Nuclei in Solid State

Dear Professor Shapiro,

Extending our NMR studies of heteronuclei in liquids to the solid state we succeeded in the following topics:

- 1) For the spin 1/2-nuclei ⁷⁷Se and ¹²⁵Te large chemical shifts and partly small line widths have been observed in IIb- and Pb-selenides and tellurides (1).
- 2) The anisotropy of the shielding of ²⁰⁷Pb found in lead nitrate powder (A.Nolle, Z.Naturforsch. 32a, 964(1977)) has been investigated now in a lead nitrate single crystal (a gift by Prof. Haussühl, Köln). The shielding tensors and the characteristic vectors could be given (2,3)
- 3) For ¹²⁵Te in CdTe, HgTe and PbTe powders a line splitting was observed which has been analyzed in terms of direct dipoledipole-coupling and isotropic and anisotropic indirect spin-spin-coupling (4). The procedure could be confirmed by a comparison with a single crystal measurement in CdTe (A.Nolle, Z.Physik B34, 175(1979)).
- 4) For the quadrupolar nuclei ³³S and ⁶⁷Zn, which have rather low receptivities, narrow signals have been detected in powdered chalcogenides. Absolute shielding constants could be given for ⁶⁷Zn (5).

A typical signal of 67 Zn in zinc selenide powder is given in Fig. 1.

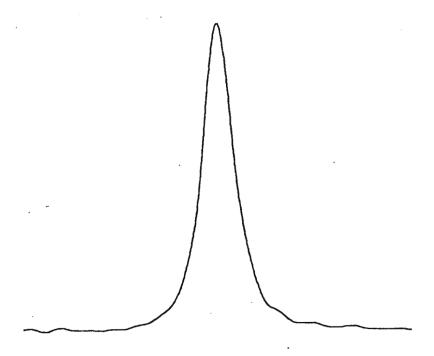


Fig. 1: ⁶⁷Zn NMR signal in ZnSe powder, Larmor frequency at 2.11 T: 5.633 MHz, line width: 49 Hz, number of FID: 10800, measuring time: 15 h.

Sincerely yours

(O. Lutz)

- (1) 77 Se and 125 Te Nuclear Magnetic Resonance Investigations in II - VI and IV - VI Compounds
- W. Koch, O. Lutz, and A. Nolle, Z. Physik A289, 17 (1978) (2) Angular Dependence of the 207 Pb NMR linewidth in Pb(NO $_3$) $_2$ due to ¹⁴N-²⁰⁷Pb Direct Dipole-Dipole-Interaction
 - O. Lutz, A. Nolle, Z. Physik B35, 245 (1979)
- (3) Nuclear Magnetic Shielding Tensors of 207Pb2+ in Pb(NO3)2 O. Lutz, A. Nolle, Z. Physik B36, 323 (1980)
- (4) 125 Te NMR Studies of Indirect and Direct Dipole-Dipole-Coupling in Polycristalline CdTe, HgTe, and PbTe R. Balz, M. Haller, W.E. Hertler, O. Lutz, A. Nolle, and R. Schafitel, J. Mag. Res., in press
- (5) ⁶⁷Zn and ³³S Nuclear Magnetic Shielding in Zinc Chalcogenides M. Haller, W.E. Hertler, O. Lutz, and A. Nolle Solid State Comm. 33, 1057 (1980)

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Pr. B.L. SHAPIRO

Texas A.M. University

College of Science COLLEGE STATION

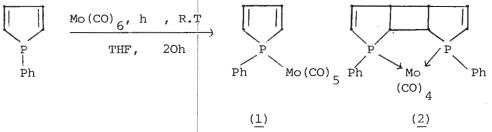
Texas 77 843 U.S.A.

Paris, le July 25, 1980

ITLE: Further Phosphole Structures by ¹H, ¹³C, ³¹P NMR

Dear Barry,

In a preceding letter we have described the "4 + 2" dimerization of 1-phenyl-3,4-dimethylphosphole by reaction with Mo(CO) under U.V. However, when reacting 1-phenylphosphole with molybdenum hexa-carbonyl under U.V.; a "2 + 2" dimer is obtained instead of the expected "4 + 2" dimer:



The original "head to head" structure of complex (2) was established by NMR and mass spectroscopy: H NMR (CDCl , internal TMSi, 100 MHz, P decoupling): 6 = 3.12 (d x m, J(H-P) 17.7 Hz, 2H, saturated cyclic Ch); 6.15 (ABX system, 2J(H-P) 25 Hz, J(H-H) 7.64 Hz, 2H, olefinic cyclic C α H); 6.37 (ABX system, J(H-P) 34 Hz, 2H, olefinic cyclic C α H); 7.36 (m, 6H, Ph meta-para); 7.61 (m, 4H, Ph ortho) ppm. P NMR (CDCl): α P = 89 ppm. C NMR (CDCl , internal Me α Si, H decoupling): α = 46.25 (d, J(C-P) 19 Hz, saturated cyclic C α); 128.2 - 132.2 (m, Ph and olefinic cyclic C α); 140.2 (s, olefinic cyclic C α) ppm.

Since only minor changes in the 13 C chemical shifts are observed when complexing a chelating diphosphine with a Mo(CO) $_4$ moiety (1), the attributions have been made for (2) by comparison with the 13 C spectrum of 1-phenylphosphol-2-ene 1. The perfectly symetrical structure of complex (2) is well in evidence on the 13 H and 31 P NMR spectra.

We have no explanation for this with result at the present time.

A regards (COV C)

G. MAVEL, C. SANTINI, F. MATHEY, R. MANKOWSKI-FAVELIER

Siège Social, Direction Générale, 18 bis, bd de la Bastille, 75012 Paris. Tél. (1) 340.38.98



DEPARTMENT OF HEALTH, EDUCATION, AND WELFARE PUBLIC HEALTH SERVICE NATIONAL INSTITUTES OF HEALTH

Professor Barry Shapiro Texas A & M University College Station, Texas 77843 NATIONAL INSTITUTE ON AGING GERONTOLOGY RESEARCH CENTER BALTIMORE CITY HOSPITALS BALTIMORE, MARYLAND 21224

July 9, 1980

Dear Professor Shapiro:

I am looking for a recent Ph.D. who is highly skilled in NMR techniques to work with me on a number of projects that involve mechanisms of biological processes. Applicants who are not citizens of the U.S. will be considered under the Fogarty fellowship program. If you know of a qualified individual, please let me know, or have the person contact me directly.

Thank you very much for your attention.

Sincerely,

Gunther L. Eichhorn

Chief, Laboratory of Cellular &

Molecular Biology

THE UNIVERSITY OF BRITISH COLUMBIA

2075 WESBROOK MALL VANCOUVER, B.C., CANADA V6T 1W5

DEPARTMENT OF CHEMISTRY

8 July, 1980

Professor Bernard L. Shapiro TAMU NMR Newsletter Department of Chemistry Texas A & M University College Station, TX 77843 U.S.A.

New Campus Instrument Center at Ohio State University

Dear Barry,

I have the pleasure to announce that I will be joining the faculty of Ohio State University in September, 1980, as professor of chemistry and biochemistry, and director of the campus instrument center.

The center equipment will initially include: Bruker WH-300 FT-NMR spectrometer, Kratos MS-25 GC-mass spectrometer, and computerized Kratos MS-9 mass spectrometer. Further expansion in NMR and mass spectroscopy is contemplated over the next few years. Users will initially include members of the chemistry, biochemistry, and pharmacy departments.

Our own research on NMR (1 H, 19 F, 31 P) of 5S RNA, and dispersion-absorption plots for analyzing NMR line shapes will continue at 0.S.U., and I will be sending a contribution shortly to TAMU in these areas.

Sincerely,

Alan G. Marshall Associate Professor

INSTITUT FÜR PHYSIK DER UNIVERSITÄT BASEL EXPERIMENTELLE KERNPHYSIK

Klingelbergstrasse 82, Telefon 061 - 44 22 80

Prof. Dr. P. Diehl

Prof. B.L. Shapiro
Department of Chemistry
Texas A & M University

College Station, Texas 77843
USA

Direct couplings providing poor structural information

Dear Barry,

In the field of NMR of oriented molecules we quite often come across cases of direct couplings which turn out to carry very poor structural information, i.e. the coupling can be interpreted by a large range of internuclear distances with almost negligible change in bond angle. As the analysis is always performed by computers nobody has tried to explain this peculiarty and to give a general condition for its occurence. In the present newsletter I would like to make up for this neglect.

The relation for a direct coupling: $D = (-k/r^3)[(Sx-Sy)\cos^2\alpha + Sy]$ may be interpreted as a graph in two dimensions: $(1/r^3)[(Sx-Sy)\cos^2\alpha + Sy] + D/k = 0$ respectively $x [a \cdot y + b] + D/k = 0$ axy + bx + D/k = 0

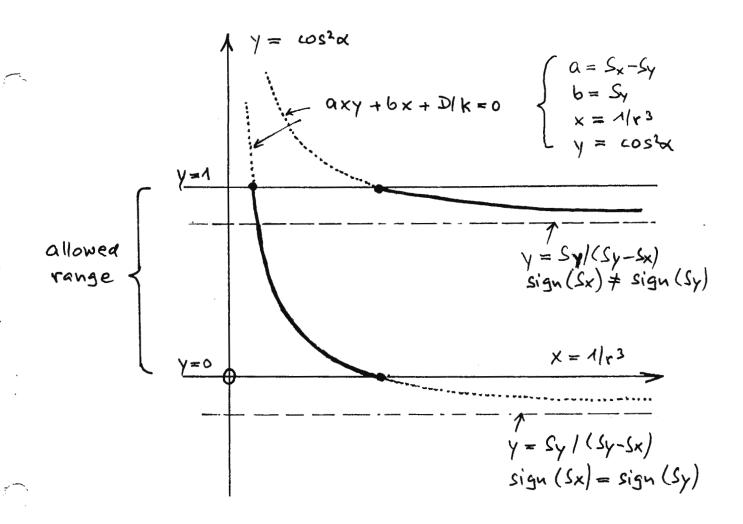
This relation corresponds to a hyperbola with the following two asymptotes:

 $x = 1/r^3 = 0$ and y = -b/a = Sy/(Sy-Sx)Since $y = cos^2\alpha$, only the part of the hyperbola which lies between y = 0 and y = 1 is a solution of the problem. It now becomes immediately obvious that if the asymptote y = Sy/(Sy-Sx) lies in the allowed range between y=0 and y = 1, the corresponding direct coupling defines the internuclear distance very poorly (see the figure!).

The condition O<Sy/(Sy-Sx)<l is fulfilled if Sx and Sy have opposite signs. Here we have the simple reason for the well known fact that if the molecular plane contains directions of S=0, the structure of the molecule is usually poorly defined.

Yours sincerely

Peter Diehl



University of Illinois at Urbana-Champaign

School of Chemical Sciences Urbana, Illinois 61801

July 22, 1980

Professor B. L. Shapiro Department of Chemistry Texas A & M University College Station, Texas 77843

Dear Barry:

500, 360 MHz Postdoctoral Positions

I shall have several postdoctoral positions opening up in my group for the coming year, for work on the NMR of systems of chemical and biological interest using our new 500 and 360 MHz widebore spectrometers.

Areas of primary interest are as follows:

- 1) ¹³C and ²H NMR of solids at 11.7Tesla (500 MHz ¹H frequency), with special reference to studies of membrane and protein-crystal dynamics.
- 2) 13C 2D-FT at 360 MHz (Oxford WB-360 and Nicolet 1180 data system), studying in particular magnetically ordered protein crystals and oriented membranes.
- 3) Metal-ion NMR (Fe, Mo, Zn) in systems of biological interest.
- 4) High-field magic-angle spinning

Candidates should submit a letter of application, a curriculum vitae with a list of publications, and arrange to have three letters of recommendation sent to me at the following address:

Professor Eric Oldfield School of Chemical Sciences University of Illinois at Urbana-Champaign Urbana, Illinois 61801

The starting salaries are negotiable and the appointments are renewable for up to three years. I will be happy to answer any preliminary inquiries by phone, at (217) 333-3374 or (217) 333-8328.

Best regards,

Yours sincerely,

Eric Oldfield



(303)491-6480

Department of Chemistry

Colorado State University
Fort Collins, Colorado
80523

July 24, 1980

Title: Permanent NMR Staff Position

Professor Bernard L. Shapiro TAMU NMR Newsletter Department of Chemistry Texas A&M University College Station, TX 77843

Dear Barry:

I hope that some of your readers will be interested in the following announcement.

Colorado State University seeks an NMR Spectroscopist for a permanent staff position in the Department of Chemistry. The starting annual salary will be in the range \$16,000 to \$20,000, depending on background and experience. The candidate must have an advanced degree in chemistry and a strong background in nmr. The responsibilities will be weighted heavily toward providing nmr service on the Department's spectrometers, and will also involve supervision, spectrometer instruction, maintenance, and repair. Collaboration with faculty on NMR problems is encouraged. Applications should include curriculum vitae, bibliography and three letters of recommendation, and should be sent to D.F.S. Natusch, Chairman, Department of Chemistry, Colorado State University, Fort Collins, Colorado 80523. Colorado State University is an EEO/Title IX Employer. Equal Opportunity Office: 314 Student Services Building.

Sincerely,

Gary . Maciel

Professor of Chemistry

NORTHWESTERN UNIVERSITY

EVANSTON, ILLINOIS 60201

DEPARTMENT OF CHEMISTRY

July 28, 1980

Professor Bernard L. Shapiro Department of Chemistry TAMU NMR Newsletter Texas A&M University College Station, Texas 77843

Dear Barry:

We are looking for a manager of our NMR equipment. The following ad describes the position.

NMR Specialist

Challenging position in the Chemistry Department Analytical Laboratory for a professional with experience in the operation and maintenance of NMR spectrometers and/or expertise in high-frequency analog and digital electronics. Knowledge of organic chemistry desirable. Current NMR instrumentation includes: JEOL FX-90Q, Varian CFT-20, T-60, and EM-360, and Perkin-Elmer R-20B. In addition, it is expected that a 250 MHz superconducting FT-NMR system will be acquired. Opportunity to work with other instruments, such as the Nicolet FT-IR spectrometer. Must have an advanced degree or comparable experience.

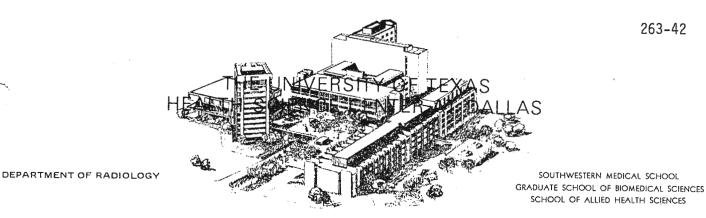
Send resume to: Manager of Employment, Northwestern University, 720 University Place, Evanston, IL 60201

An equal opportunity affirmative action employer m-f.

Sincerely,

Joseph B. Lambert

Title: Position Available for NMR Manager



July 28, 1980

Professor B.L. Shapiro Department of Chemistry Texas A&M University College Station, Texas 77843

Title: "Who shot J.R., and does anyone have a used pulse spectrometer for sale?"

Dear Dr. Shapiro:

In initiating my subscription to the Newsletter, I should like to let your readers know that after a very fruitful and instructive tenure with Dr. Donald Hollis, I am starting my own NMR laboratory at the University of Texas Health Science Center at Dallas. is my intention to pursue NMR studies of cellular metabolism and also to investigate the potential of high field imaging based on metabolic states. It is envisioned that this second area will be directed toward basic research in physiologic (and pathophysiologic) studies of perfused organs and in vivo in small animals. It is likely that I will be able to support a postdoctoral fellow to work in these areas by late spring or early summer of 1981. Well qualified senior graduate students planning on completing their degrees within this time frame are encouraged to contact me.

Dr. Donald Twieg (also at this institution) is interested in obtaining a used pulse spectrometer (without computer) in good condition. Please contact him at 214/688-2259 or write to him at this address:

> Department of Radiology University of Texas Health Science Center 5323 Harry Hines Blvd. Dallas, Texas 75235

if you have a used spectrometer for sale.

Sincerely,

Ray L. Nunnally, Ph.D. Assistant Professor

dh

UNIVERSITY OF GRONINGEN 263-43



DEPARTMENT OF ORGANIC CHEMISTRY
ZERNIKELAAN, GRONINGEN
THE NETHERLANDS
TEL: 050 - 11
JBFNE/tld

Dear Janos,

Professor J.H. Fendler
Department of Chemistry
Texas A & M University
College Station
Texas 77843
USA

Groningen, June 20, 1980

We would like to bring to your attention our interest in offering a highly qualified Ph.D. a one, two or three year appointment (tenure not absolutely excluded) in our Division starting in September 1980 or January 1981. The candidate must be well-versed and experienced in modern NMR techniques and willing to devote upto 50% of his time to the service aspects of our XL-100 (with \$^{13}{\rm C}\$ probe) and the 200 MHz Nicolet apparatus, which will arrive in December of this year. We would strongly encourage research although our first need is high quality service. The technical assistance will consist of (at least) one part or full time technician and at least one or possibly two, (part-time) graduate students as assistants. Our Organic Division consists of six full Professors of which five have an active research program. The Division has at present sixty graduate students and ten technicians. The salary ranges from US \$ 20-25,000,- depending on age and experience and has all the advantages of a civil-service appointment.

I enclose: a) Recent biographical data on members of the Division.

b) Summary of the structure of the Chemistry Department.

5

c) Additional data on Groningen and the University.

If you can recommend someone strongly for this position please let him write to the Chairman of the Division (Prof. Dr. H. Wynberg). The candidate should enclose biographical data and names of two persons familiar with his work, who can provide letters of reference.

We would expect the candidate to be between 24 and 35 years of age, to have completed the requirements of the Ph.D. degree or its equivalent and be motivated to enter academic life by way of NMR research, theory, perhaps a teaching task and service duties. Willingness to devote time and energy to work with graduate students and staff members on NMR problems connected with their research is essential.

J.B.F.N. Engberts
Professor of General Chemistry

NT-Series Fourier Transform Superconductive Magnet NMR Spectrometers

The NT-Series has been conceived and designed to provide optimum performance while being fully adaptable to new techniques with minimal cost and difficulty. More than just a collection of instruments, the NT-Series represents a completely modular approach to FT-NMR instrumentation that allows the user to expand his system as his research needs grow and to easily accommodate new experimental techniques as they develop.

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- Precise digital plotting with full annotation of spectral parameters and flexibility of hardcopy format.

The multiple-technique NT-Series spectrometers provide the user with the ability to easily adapt to the newest techniques and experimental configurations.

Some of these are:

- High-resolution studies of solids with Waugh-Pines crosspolarization and magic-angle spinning
- High-sensitivity wide-bore ¹³C studies of high molecular weight polymers.
- Automated T₁ and T₂ measurements.
- Chemical dynamics studies.
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- ³¹P experiments on living organs.



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