Texas

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No. 188

May, 1974

Α.	Pines Shtuyot Bemitz Agvaniyot	,
R.	Mondelli  1H and 13C Spectra of Thietane-1-Oxide	3
J.	I. Musher Position Available	5
J.	Ray and B. Santini A Simple Instrumentation Amplifier Between an NMR and Signal Averaging Computer	6
R.	L. Lichter Postdoctoral Position Available	9
Р.	D. Ellis Hydrocarbon <sup>13</sup> C Chemical Shifts and Postdoctoral Position Available	10
G.	J. Martin and M. L. Martin Dependence of the Coalescence Temperature in Two-Site Chemical Exchange Studies	12
J.	F. Garst Postdoctoral Position Available	15
J.	T. Clerc, H. P. Meier and E. Pretsch T <sub>1</sub> Error Analysis for the Inversion Recovery Technique	16
D.	J. Sardella Allylic Couplings and Bond Fixation in α-Pyrones	18
F.	W. Wehrli Solvent Suppression in Proton FT NMR on the XL-100: Some Practical Hints	20
н.	C. Clark Position Available	25
М.	C. Thorpe 13C NMR of Isomeric Dithianes	26
C.	K. Tseng and A. Mihailovski Trifluoroacetic Acid as a Solvent	28
Α.	Briquet, J. C. Duplan and J. Delmau T <sub>1</sub> Measurements Showing Convection Streams	31

Newsletter

(Cont'd on p. (i))

A monthly collection of informal private letters from Laboratories of NMR. Information contained herein is solely for the use of the reader. Quotation is <u>not</u> permitted, except by direct arrangement with the author of the letter, and the material quoted <u>must</u> be referred to as a "Private Communication". Reference to the TAMU NMR Newsletter by name in the open literature is strictly forbidden

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G.	Gurato and G. Zenoni  19F NMR Analysis of Fluoride of Perfluorocyclohexanesulphonic Acid and of 3-Trifluoromethyl-perfluorocyclopentane-1-Sulphonic Acid	34
R.	R. Fraser The Conformation of N-nitroso- $\underline{r}$ -2-cis-6-dimethyl-4-phenylpiperdine, $\underline{I}$	39
L.	D. Colebrook, F. H. Hund and S. Icli Spectra of Internal Rotational Isomers in a Chiral Solvent	42
Н.	J. Jakobsen, R. S. Hansen, S. A. Linde and S. Sørensen Selective Population Transfer (SPT) in FT NMR; A Useful Technique Equivalent to TSI in CW NMR	44

Deadline Dates: No. 189: 3 June 1974

No. 190: 1 July 1974

All Newsletter Correspondence, Etc. Should Be Addressed To:

Dr. Bernard L. Shapiro Department of Chemistry Texas A&M University College Station, TX. 77843 U.S.A.

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DEPARTMENT OF CHEMISTRY

BERKELEY, CALIFORNIA 94720

March 28, 1974

Professor B. L. Shapiro
Department of Chemistry
Texas A & M University
College Station, Texas 778

77843

Dear Barry:

We have constructed a device which is quite useful for nmr in all types of systems. This has several novel features, e.g.:

- 1. Produces a 11166 pulse 387T sequence with independently adjustable widths and delays. This eliminates all shifts and couplings in any spectrum, including chemical shifts, Kondo shifts, scalar coupling, dipolar coupling, quadrupolar coupling, Knight Shifts, Day Shifts, and Suhl-Nakamura coupling, leaving a simple, clear, easily interpretable spectrum consisting of one sharp line. Analysis of this spectrum is performed automatically.
- 2. Eliminates automatically large extraneous peaks due to solvents. The solvent to be eliminated is selected by pushbutton out of a collection of 1704 entries. Solvent peaks may also be added.
- 3. Sensitivity is enhanced by transferring polarization from a large sample maintained in Saclay at 2°K. Polarization is running out so competition is keen and proposals must be submitted for permission to transfer.
- 4. Operates with 130 frequency synthesizers and a magnetic field calibrated according to the theorem  $\omega_o = \gamma H_o$ . The lock is external and operates on  $^5\text{Li} ^{11}\text{Li}$ ; it is powerful and can be put on another campus.
- 5. The machine smiles during a spin tickling experiment.
- 6. Equipped with a 45 KW power transmitter. This is supplied with an answering service to reply to complaints from radio amateurs in Siberia.
- 7. Optical detection is optional. Sensitivity is so enormous that half a photon has been detected. The other half is being kept spin-locked under close guard of a hyperfine component.
- 8. The device operates in any frame including the rotating, pulsating, and randomly fluctuating frames.
- 9. Can do DEFT, TDFT, PRFT, WEFT, LEFT, GLBM, etc., by selecting a combination of four letters. If no experiment exist for a selected combination, the device will invent one and publish a preliminary communication.

This work was hardly supported.

Best regards,



188-

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Milano, March 15th, 1974

Prof. Bernard L. Shapiro
Dept. of Chemistry
College of Science
Texas A & M University
College Station, Texas 77843
U. S. A.

Dear Professor Shapiro,

I have received today your pink letter, so I must apologize again for my delay and I send you immediately our contribution, whose title is:

"  $\frac{1}{1}$ H and  $\frac{13}{1}$ C spectra of thietane-1-oxide "

We have continued our n.m.r. investigations on four-membered rings, in particular thietanes. In thietane derivatives only a few complete analysies have been published, whereas many stereochemical assign/ments are based on approximate n.m.r. results. Moreover the absence of complete experimental data of unsubstituted molecules and the lack of knowledge on their conformation did not allow an adequate test of the theoretical studies. We have analyzed the H and 13C spectra of thietane-1-oxide in order to determine the ring conformation and evaluate whether a correlation is possible between structure and n.m.r. parameters, particularly J's, normally used for stereochemical assignments.

The analysis was carried out first on the spectrum at 300MHz which, although strongly coupled, gave the four chemical shifts and the approximate values of the three-bond couplings and  $^2J_{CD}$ . The refinement of these parameters and the determination of the remaining ones were obtained by iterative procedure, performed on the spectrum at 100 MHz (fig. 1). From the n.m. r. data in the range of temperature + 190° and - 140°C, evidence was obtained that a strongly puckered structure with the sulphinyl oxygen equatorial is rather more populated than the other one, although the ring inversion rate is fast enough to average the coupling constant values even at low temperature. Increasing the temperature, a slight increase in the population of the less stable conformer (sulphinyl oxygen axial) has been deduced from variations of  $^3J_{AC}$   $^3J_{BD}$   $^4J_{BB}$ , and  $^4J_{AC}$ .

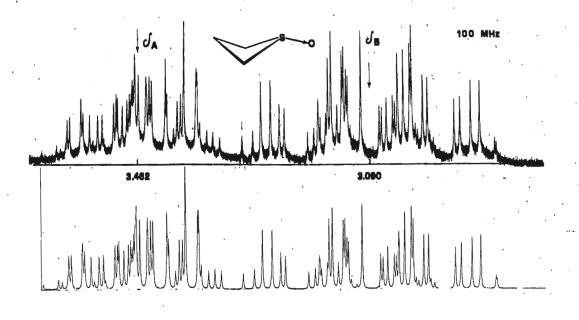
AA' A complete discussion on the coupling constant values in thietane-1-dioxide and related four-membered heterocycles will appear shortly in a full

The <sup>13</sup>C spectrum without <sup>1</sup>H decoupling is reported in Figure 2 together with all the coupling constant values, that could be obtained complete of their signs from the <sup>2nd</sup> order analysis.

Sincerely yours

(R. Mondelli)

Joseph Mandle .



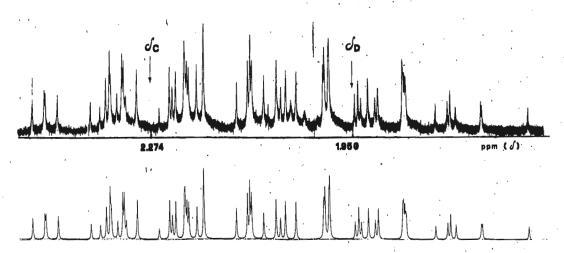
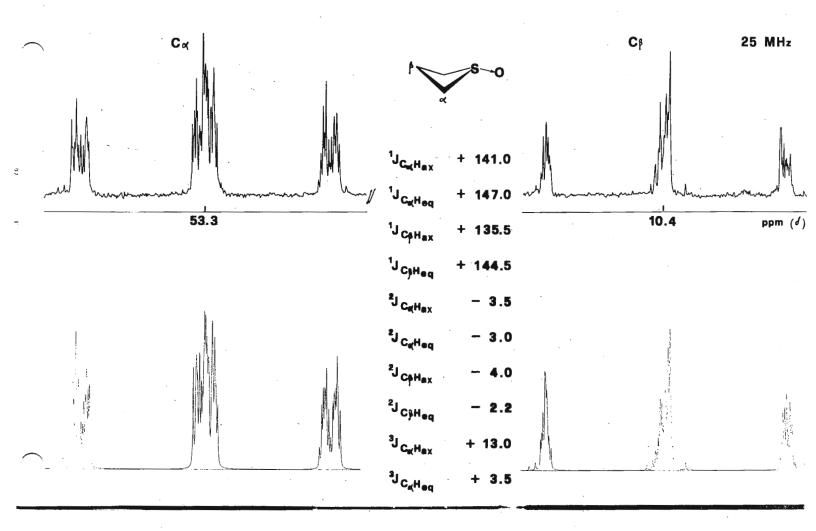


Figure 1

2 <sub>J</sub> AB	- 10.29 ±-0.005 Hz	acetone-d <sub>6</sub> 30°C
<sup>2</sup> J <sub>CD</sub>	- 12.69 <b>±_</b> 0.006	RMS 0.031
3 <sub>JBD</sub>	+ 12.53 ± 0.005	
J <sub>AC</sub>	+ 1.83 ± 0.005	*The probable error for
3 <sub>JBC</sub>	+ 10.63 ± 0.006	each parameter should
$^3$ J <sub>AD</sub>	+ 7.49 ± 0.005	increased at least by
<sup>4</sup> J <sub>AA</sub> ,	+ 6.10 ± 0.008	factor of two.
4J <sub>BB</sub> ,	- 0.57 ± 0.007	
4JAB'	$-$ 0.95 $\pm$ 0.006	





# BELFER GRADUATE SCHOOL OF SCIENCE

2495 Amsterdam Avenue / New York, N.Y. 10033 / (212) 568-8400

29 March 1974

Dr. B.L. Shapiro Department of Chemistry Texas A&M University College Station, Texas 77043

Dear Barry,

I am also interested in an organic chemist interested in working on novel compounds of Se (non-toxic), Te and I. Salary is \$8000-\$8500. The position begins after Sept. 1.

Yours sincerely,

Lew

Jeremy I. Musher

Standard Oil Company (Indiana)

Standard Oil Research Center Post Office Box 400 Naperville, Illinois 60540 312-420-5111

April 1, 1974

Dr. B. L. Shapiro
Department of Chemistry
Texas A & M University
College Station, TX. 77843

TITLE: A simple instrumentation amplifier between an NMR and signal averaging computer.

Dear Barry:

Please credit this contribution to the Standard Oil (Indiana) subscription.

We would like to describe a simple and effective amplifier to provide signal conditioning between an existing NMR spectrometer and the Analog to Digital converter (ADC) of a signal averaging system. The circuit is suitable for both CW and FFT operation. It provides a high degree of common mode rejection in order to avoid ground loops and noise pick-up. It will also generate a ±10V offset voltage to properly DC null the NMR signal before it reaches the ADC. As a result the experimenter may obtain optimal digital resolution and long signal averaging times without overranging the storage word in the computer system.

Referring to the attached schematic,  $A_1$  and  $A_2$  are respectively, a variable voltage reference and a summing amplifier.  $A_3$ ,  $A_4$  and  $A_5$  form a classical differential instrumentation amplifier. Each input may be floated relative to ground. In normal operation, inputs 2 and 3 are connected to the analog output of the NMR receiver. A shield may be connected to the common of the circuit. It is essential that this shield is <u>not</u> connected to the spectrometer or the noise rejection properties of the amplifier will be lost. The resistors around  $A_5$  must be closely matched to achieve the highest possible common mode rejection ratio. The signal from the instrumentation amplifier is summed with the variable reference voltage in  $A_2$ . The meter circuit on the output of  $A_2$  provides a convienent means to null any DC component from the NMR signal before the signal is passed on to the computer for averaging.

Additional details of the circuit include two zener diodes across the input to protect against pulse break-through damage. A spring loaded switch increases the sensitivity of the meter circuit momentarily for precise balance near DC null. The decoupling scheme is shown. This scheme

B. L. Shapiro April 1, 1974

and proper layout practices must be employed to avoid oscillation. The basic bandwidth of the circuit is about 100 KHz with a gain of unity. The bandwidth may be restricted by the addition of  $\mathbf{C}_{\mathbf{X}}$  to the feedback path of A2. A more effective approach is to use a tunable active filter on the output circuit, before the ADC. The operational amplifier types illustrated were carefully chosen for their almost ideal phase shift vs frequency characteristics and should not be substituted.

This circuit has been used successfully between a Bruker HFX-90 and a Nicolet 1070, an XL-100 and Nicolet 1080 and in installations involving IBM computer systems.

We hope that this circuit will prove helpful to anyone who is attempting to optimize the transfer of analog data between an NMR and signal averaging computer, regardless of the types involved.

Sincerely yours,

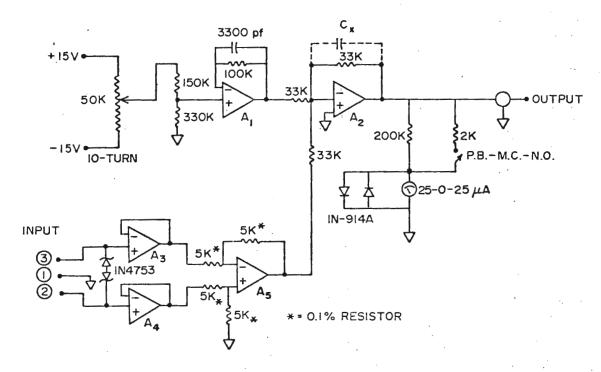
Joe Ray

Standard Oil, Inc. (Indiana)

Bet Senden

Bob Santini
Department of Chemistry
Purdue University
W. Lafayette, IN. 47907

GJR:slk

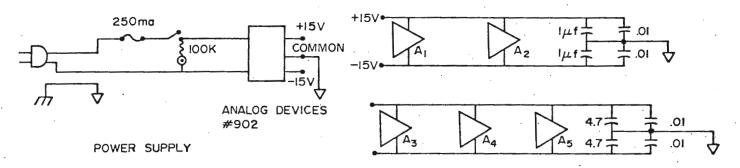


A<sub>1</sub> to A<sub>5</sub> = MOTOROLA MC-1456CG or SIGNETICS 5556C

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# Hunter College

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Telephone

April 11, 1974

Dr. Bernard L. Shapiro Department of Chemistry Texas A & M University College Station, Texas 77843

Dear Barry:

Title: Postdoctoral Position Available

I am seeking a postdoctoral research associate for a one- or two-year period beginning 1 September 1974 to collaborate in doing nitrogen-15 nmr. The research areas include structure-chemical shift correlations, medium effects on chemical shifts, and applications to biochemically and pharmacologically important compounds. We also are determining coupling constants between 15M and other nuclei. All the chemical shift work is carried out at the natural-abundance level of 15M on a JEOL PS/PFT-100 spectrometer with deuterium lock. In addition, we have become involved in some aspects of instrument modification.

Interested individuals, who should have some nmr experience and preferably some acquaintance with FT nmr, should contact me at the above address.

Hunter College is located in the upper east side of Manhattan and is easily accessible to all parts of New York City. Needless to say, we are an equal opportunity employer.

See you in Raleigh.

Sincerely,

Robert L. Lichter Assistant Professor

RLL:bd



# UNIVERSITY OF SOUTH CAROLINA COLUMBIA, S. C. 29208

Department of Chemistry (803) 777-5263

April 1, 1974

Professor B. L. Shapiro Department of Chemistry Texas A & M. University College Station, Texas 77843

TITLE: Hydrocarbon <sup>13</sup>C Chemical Shifts and Postdoctoral Position Available

Dear Barry:

We have been involved recently with the development of a semi-empirical theory of nmr chemical shifts, which could be applied to any atom in the first long row of the periodic table. To properly check the calculations for hydrocarbons, we obtained the  $^{13}\text{C}$  chemical shifts of seven hydrocarbons, i.e., CH<sub>4</sub>, C<sub>2</sub>H<sub>6</sub>, C<sub>3</sub>H<sub>8</sub>, C<sub>4</sub>H<sub>2</sub>, C<sub>2</sub>H<sub>2</sub>, C<sub>3</sub>H<sub>4</sub>, and C<sub>2</sub>H<sub>4</sub>. These shifts were determined on a Varian CFT-20 in CS<sub>2</sub> solvent with 10% acetone-d<sub>6</sub> added for a lock material. The temperature was -80°C. The data are summarized in the accompanying table. It is important to note that the value of the chemical shift between CH<sub>4</sub> and C<sub>2</sub>H<sub>6</sub> is -11.23 ppm. Our value is excellent agreement with the one reported by Strong, et al., J. Magn. Resonance, 9, 145 (1973).

Also I would like to let your readers know of the availability of a nonteaching postdoctoral position in my laboratories. The research involves the study of the mmr parameters (shifts and  $T_1$ 's) for the pyridine dinucleotides interacting with several enzymatic systems. The nuclei of interest are 1H,  $^2\mathrm{H}$ ,  $^{13}\mathrm{C}$ ,  $^{15}\mathrm{N}$  and  $^{31}\mathrm{P}$ . Furthermore, the postdoctoral (hopefully) will be involved in some fundamental modification to the XL-100, e.g. quandrature phase detection, etc. The salary for this position is \$8500. The University of South Carolina is an equal opportunity/ affirmative action employer.

I would ask any interested individuals to write or phone me (803-777-6039) for further information.

Sincerely,

Paul D. Ellis

Assistant Professor

Carbon-13 Chemical Shifts of Some Representative Hydrocarbons

	Chemical Shif	t	•
Compound	δ (CS <sub>.2</sub> ) <sup>a</sup>	δ (TMS) <sup>b</sup>	δ (CH <sub>4</sub> ).
CH <sub>4</sub>	195.61	2.8	0.0
<sup>C</sup> 2 <sup>H</sup> 6	184.38	-8.4	-11.23
CH3CH2CH3	175.67	-17.1	-19.94
CH <sub>3</sub> CH <sub>2</sub> CH <sub>3</sub>	175.50	-17.3	-20.11
H <u>C</u> ≅C−C≡ <u>C</u> H	126.34	-66.5	-69.27
HC <u>=C</u> −C <u>=</u> CH	124.63	-68.2	-70.98
$^{\mathrm{C}}_{2}^{\mathrm{H}}_{2}$	119.67	-73.1	-75.94
H <sub>2</sub> C=C=CH <sub>2</sub>	118.18	-74.6	-77.43
<sup>C</sup> 2 <sup>H</sup> 4	<b>69.</b> 34	-123.5	-126.27
H <sub>2</sub> C=C=CH <sub>2</sub>	-19.78	-212.6	-215.39

a) Internal CS $_2$  as determined on a Varian CFT-20 operating at  $\sim 80\,^{\circ}$ C locked to a 10% acetone-d $_6$ . Positive chemical shifts denote shifts to higher shielding.

b) External TMS using the relationship  $\delta \, (\text{TMS}) \, = \, \delta \, (\text{CS}_2) \,$  -192.8

UNIVERSITÉ DE NANTES U E R DE CHIMIE

38, Bd Michelet - 44 - NANTES B. P. 1044 Tél. (40) 74-50-70

> Professor Bernard L. SHAPIRO Department of Chemistry TEXAS A-M University College Station

> > TEXAS 77843

Dear BARRY,

Dependence of the coalescence temperature in two-site chemical exchange studies. A complement to Professor GUTOWSKY'S letter.

An alternative to Professor's GUTOWSKY (1) way of obtaining enthalpies and entropies of activation from the rate constant at the coalescence point, is conveniently offered by the simultaneous use of H and 13C resonance.

Some results concerning the hindered rotation around C-N bonds in amidines are represented in the table. The presence of many diastere-otopic nuclei improves the quality of the method as shown for compound 2. Consideration of different sites is certainly a source of discrepancies although the intrinsic variation of the chemical shift with the temperature must not be excluded in the one-site multi-frequencies experiments.

The method making use of both  $^{1}\text{H}$  and  $^{13}\text{C}$  resonance is somewhat cheaper than the elegant procedure of Professor GUTOWSKY since:  $\Sigma$  US \$ (A 60 + HA 100 + HR 220 + further developments of superconducting solenoids)  $\gg$ 

∑US \$ (CFT 20 + XL 100)

It may also be noted that a twenty fold range in frequency and rates may easily be covered by this technique.

	<sup>1</sup> H (a)				<sup>13</sup> с (ъ)			
	CFT 20*		CFT 20* XL 100		CFT 20*		XL 100	
	۵νω	Тс	Δλω	Tc	VĵΦ	Tc	ω(۵	Тс
C <sub>6</sub> H <sub>5</sub> -N=CH-N(CH <sub>3</sub> ) <sub>2</sub> 1 (a)	6.2	-	7.8	278	116	-	145	318
C <sub>6</sub> H <sub>5</sub> -N=CH-N(CH <sub>2</sub> CH <sub>3</sub> ) <sub>2</sub> 2 ba b	(13.4	-	16 <b>.</b> 8	275 -	128 56	_	160 70	304 294

\* At the present time, the values relative to the CFT 20 spectrometer are virtual since we do not dispose of such apparatus, but this negligeable fact does not detract from the validity of the method. Other data will be abailable in ref. (2).

The accuracy of the results is good for the  $\Delta G^*$ , acceptable for the  $\Delta H^*$  and poor for the  $\Delta S^*$  values; it is limited, as Professor GUTOWSKY points out, by the accuracy of the temperature measurements. With the vast shortage of funds which characterizes our day and age, greater accuracy perhaps should be looked for in a drastic improvement of the "magic box", rather improperly called "temperature controller V6040" (4), or in self-made devices, than in superconducting solenoids.

### Sincerely yours.

		G.J. MARTIN	M.L. MARTIN
	$\Delta$ G*/T kcal.m <sup>-1</sup>	ΔH* kcal.m <sup>-1</sup>	ΔS* cal.Kelv <sup>-1</sup>
1	14.9/318 + 0.1	11.9 + 2	- 8 <del>+</del> 4
<u>2</u>	14.3/304 <sup>±</sup> 0.1	12.2 + 1.8	- 7 <sup>±</sup> 4

- (1) H.S. GUTOWSKY TAMU Newsletter 186, 25, (1974) -
- (2) M.L. FILLEUX, N. NAULET, J.P. DORIE, G.J. MARTIN Tetrahedron Letters 1974 in press -
- (3) G.J. MARTIN, D. LE BOTLAN, M.L. FILLEUX TAMU Newsletter, 182, 30,(1973) -
- (4) I suppose that the problem of controlling the temperature in NMR inserts is not limited to this apparatus but we have not direct experience of other commercial systems.

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- Failsafe

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### DEPARTMENT OF CHEMISTRY

# THE UNIVERSITY OF GEORGIA ATHENS, GEORGIA 30602

April 4, 1974

Prof. B. L. Shapiro Department of Chemistry Texas A&M University College Station, Texas 77843

# POSTDOCTORAL POSITION

Dear Prof. Shapiro:

I solicit candidates for a postdoctoral position in my laboratory to begin July 1, 1974, or as soon thereafter as possible.

The research will involve CIDNP from reactions of stable free radicals (radical anions) in low magnetic fields. CIDNP will be detected after these very fast reactions are complete using Fourier nmr spectroscopy. The radical pair theory and its parameterization will be tested through efforts to simulate the observed CIDNP spectra through calculations based on those models.

A candidate must be well versed in magnetic resonance theory as well as practice. In particular, he should be familiar with treatments of time-dependent phenomena involving fairly complex spin systems and with the associated computer programming. It would seem that the best candidates would probably be physical chemists with experience in DNP, other double resonance areas, or similar fields. Prior experience in CIDNP is not required.

A candidate must be able to work independently. His expertise will not be coincident with mine (physical organic chemistry, kinetics and mechanisms), but complementary.

The salary will be \$8500/yr.

Candidates should contact me directly.

Sincerely

John F. Garst

Associate Professor



# Eidgenössische Technische Hochschule Zürich. Laboratorium für Organische Chemie

CH-8006 Zürich, Universitätstrasse 16 April, 4 1974. Tel. (01) 32 62 11

Prof. B. L. Shapiro

Department of Chemistry

Texas A & M University

College Station, Texas 77843

U. S. A.

# T, Error Analysis for the Inversion Recovery Technique

Dear Prof. Shapiro,

We have recently studied some parameters affecting the accuracy and precision of  $T_1$  values measured by the inversion recovery technique. The most important factors are errors in signal intensity measurements, pulse inhomogeneity, and incorrect pulse width.

Errors in signal intensity measurements do not affect the accuracy of the  $T_1$  values. Pulse inhomogeneity and incorrect  $180^{\circ}$  pulse setting, however, give raise to a systematic error. If a linear regression of  $\ln(A_0 - A) / 2 A_0$  on  $\tau$  (or similar) is used for the evaluation, this bias is removed, as it affects only the intercept.

The precision of  $T_1$  values is most seriously affected by the variance of the signal intensity and to a lesser degree by the pulse inhomogeneity. The accuracy of the  $90^{\circ}$  pulse does not influence accuracy and precision of the result. In practice, the contribution of the variance of  $\tau$  as well as of inproper  $180^{\circ}$  pulse setting can be neglected. Pulse inhomogeneity is difficult to measure. However, as errors in pulse width are generally small, a measure for pulse inhomogeneity is

given by  $I = -A_{(180^{\circ} - 0 - 90^{\circ})} / A_{(90^{\circ})}$ . If one further assumes that A and  $A_{o}$  are measured with equal precision, the variance of a  $T_{1}$  value calculated from a single point with given A/A<sub>o</sub> can be estimated. In Fig. 1 the ratio between the relative standard deviation of  $T_{1}$  and the relative standard deviation of  $A_{o}$  is plotted against A/A<sub>o</sub> for two different values of the pulse inhomogeneity measure I.  $SD(T_{1})/SD(A_{o})$ 

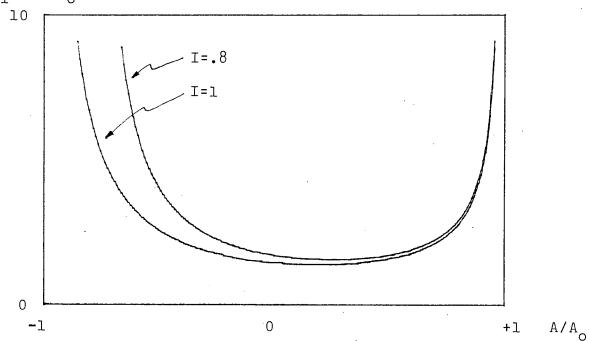


Fig. 1. Plot of the estimated ratio between the relative standard deviation of  $T_1$  and the relative standard deviation of  $A_0$  against  $A/A_0$ .

As the precision in  $T_1$  detoriorates rapidly for extreme values of  $A/A_0$ , it is suggested to exclude data points with  $|A/A_0|$  near 1.

A more detailed discussion will be submitted for publication in Org.Magn.Res.

Yours sincerely

J.T.Clerc

H.P.Meier

E.Pretsch

i.T. Cerc

Ar. Mein

E. Crehola

# HARVARD MEDICAL SCHOOL

DEPARTMENT OF BIOLOGICAL CHEMISTRY

HARVARD MEDICAL SCHOOL

# PETER BENT BRIGHAM HOSPITAL

Mailing Address:
BIOPHYSICS RESEARCH LABORATORY
Peter Bent Brigham Hospital
Boston, Massachusetts 02115

April 5, 1974

Professor Bernard L. Shapiro Department of Chemistry Texas A & M University College Station, Texas 77843

Dear Barry:

# Allylic Couplings and Bond Fixation in $\alpha$ -Pyrones

Since the relationship between  $\pi$ -bond order and the magnitude of allylic couplings involving methyl groups is well-established, it was not surprising that the nonequivalent methyls of 4-methylaminopent-3-en-2-one had widely differing allylic couplings:

One might thus anticipate that for a system in mobile equilibrium, such as acetylacetone

the allylic coupling should be approximately the average of those in the above model. In fact,  $^4\mathrm{J}$  <0.2 Hz, suggesting either 1) the model is a poor one, 2) there is no enol-enol equilibrium or 3) the allylic couplings in the model have opposite signs.

In an attempt to explore the possibility that systems involving considerable bond fixation may exhibit positive allylic couplings, I looked at the spectra of some methyl-substituted  $\alpha$ -pyrones (supplied by Dr. George Vogel of Boston College).

In CDCL solutions J exhibits methyl absorption at  $\delta$  2.26 (singlet) and  $\delta$  2.21 (doublet; J = 1.0 Hz). Coupling between the methyls is  $\delta$  0.2 Hz. No sign information was obtained.

Compound II gives a spectrum ideally suited for sign determinations. The methyl absorption ( $\delta$ 2.16; doublet of doublets, J = 1.2 and 0.25 Hz) clearly shows coupling to both ring protons (meta coupling = 1.4 Hz). Double resonance experiments indicate

which, of course, is not supportive of the original hypothesis. The question, thus, remains open, but the indication of considerable bond fixation in  $\alpha$ -pyrones is interesting and entirely consistent with their chemistry.

Best wishes to all.

Sincerely,

Dennis J. Sardella Visiting Lecturer

DS:bn



Prof. B.L. Shapiro Department of Chemistry Texas A & N University College Station

Texas 77843 U.S.A.

April, 11, 1974

Dear Barry,

Suggested Title:

Solvent suppression in proton FT NMR on the XL-100: Some practical hints

With the rapid growth of biochemical applications of proton FT NMR, instrument users are confronted with the problem of detecting weak solute resonances ( $10^{-3}$ - $10^{-5}$  molar) in the presence of the large residual HDO line. The problem has in some instances erroneously been interpreted as one of dynamic range due to the limited computer word length. This can easily be overcome by block averaging and double precision storage techniques. The limitation, however, rather lies in the ADC's dynamic range. Though the XL-100's ADC has a word length of 13 bit (12 bit plus sign), the strong solvent response forces one to reduce the spectrum amplitude in order to avoid overloading of the ADC. This on the other hand means that weak signals may be below the ADC's detection threshold. Apart from this, a solvent line, which is two orders of magnitude more intense than the resonances of interest, render the observation of signals near the solvent line impossible (spinning sidebands etc.). It is for these two reasons often mandatory to suppress the perturbing solvent line.

It should be pointed out that none of the subsequently discussed methods is essentially new. The purpose of their presentation is rather to compare the presently available methods with respect to their efficiency and to encourage instrument users to apply them.

The techniques to be discussed are:

- (i) Inversion-recovery method as described by Patt and Sykes<sup>1</sup> and Benz, Feeney and Roberts<sup>2</sup>
- (ii) Irradiation of the HDO peak
- (iii) Partial saturation using a (90°-t) sequence in the steady state regime.
- ad (i): All this method requires is (a) a standard XL-100 program, (b) a minor hardware modification allowing a short field gradient pulse to be applied to the Y shim coils, (c) a precise determination of T<sub>1</sub> (HDO) and (d) that T<sub>1</sub> (solute) (HDO).

The idea behind the method is to apply the 90° observing pulse at the moment the HDO magnetization passes through zero. A standard (180°-t-90°-T) sequence would require  $T \ge 5 \cdot T_1$  (HDO) thus excessively lengthening the experiment. Instead Benz et al. 2 have determined the steady state conditions allowing shorter values of T to be taken. After  $T_1$  (HDO) has been determined (which is a matter of seconds), a suitable pair of t, T values can be taken from the curve on page 117 in Ref. 2. In order to allow for the steady state to build up, it is important, however, that SS = 1 is used. This means that the first four pulses will be neglected. Fig1a) shows a proton FT spectrum obtained after accumulation of 500 4-sec transients under standard conditions from a 5 x  $10^{-3}$  molar solution of tryptophan in 99,75 % D<sub>2</sub>O in a 5 mm-tube. The spectrum in Fig1b) was obtained by using a (1800-HSP-t-900-T) sequence under otherwise unaltered conditions. Apart from the 160-fold reduction of the HDO peak's intensity, the spectrum shows a considerably improved signal-to-noise, because of a better ADC resolution for these signals.

ad (ii): Irradiation of the HDO peak at its exact frequency is an alternative way of suppressing the solvent resonance. This requires the FT homodecoupling accessory, in principle a simple T cable which permits the hetero decoupler output to be fed onto the observe transmitter coils. Important in this experiment is a precise setting of the decoupler frequency. This is best achieved by varying the decoupler frequency until a minimum response is observed on the oscilloscope. Fig 1 c) shows a spectrum obtained under the same conditions as the one in Fig 1 a) except that the HDO line was irradiated. The solvent suppression factor is about 30 and the signal-to-noise of the solute line comperable to that observed in Fig (b).

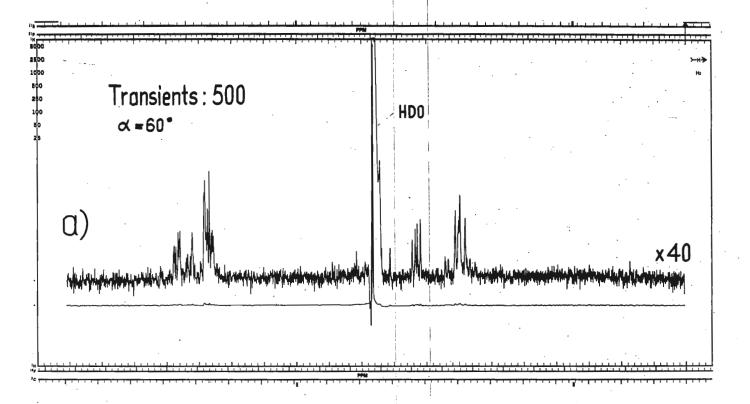
ad (iii): The simplest but also the least efficient among the methods discussed, consists of applying  $90^{\circ}$  pulses. If steady state conditions are selected and  $T_1$  (solute) $< T_1$  (HDO) this should result in a solvent line intensity reduction. The spectrum shown in Fig 1 (c) has been obtained under the same conditions as the one in Fig 1 (a) except that the first four pulses were neglected and that  $90^{\circ}$  pulses were applied.

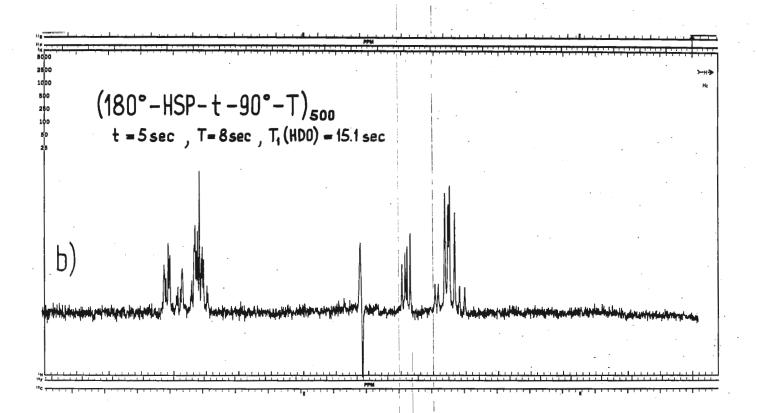
VARIAN AG Research Laboratory

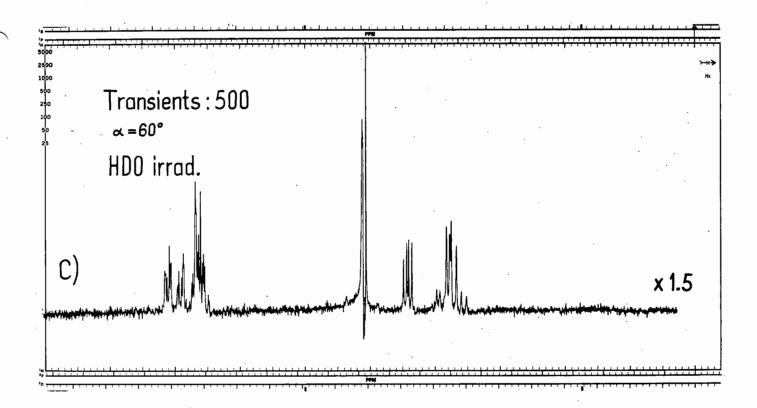
F.W. Wehrli

<sup>&</sup>lt;sup>1</sup> S.L. Patt and B.D. Sykes, J. Chem. Phys. <u>56</u>, 3182 (1972)

<sup>&</sup>lt;sup>2</sup> F.W. Benz, J. Feeney and G.C.K. Roberts, J. Mag. Res. 8, 114 (1979)







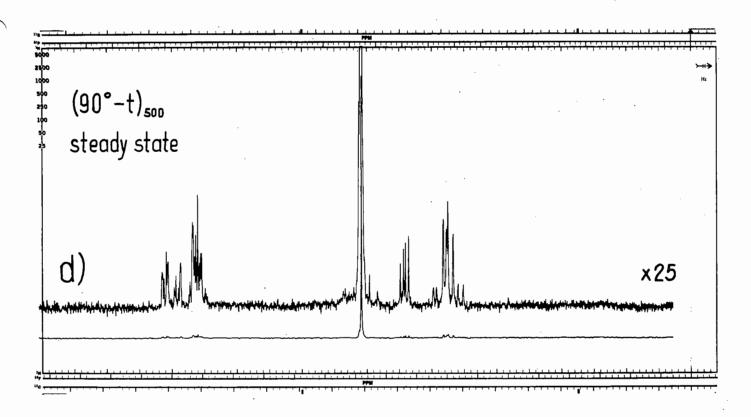


Fig. 1 a)-d): 100 MHz FT spectra of 5 x 10<sup>-3</sup> molar tryptophan in 99,75% D<sub>2</sub>O (5 mm tube) under experimental conditions as described in the text.

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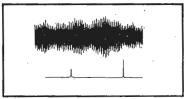
STOCHASTIC RESONANCE — This technique involves random noise modulation of the rf excitation frequency, leading to a broadband excitation whose response is measured and Fourier transformed much as in conventional pulsed-FT nmr. Its principal advantage lies in the fact that much less rf power is needed to excite the chemical shift range of interest and that it has the same Fellgett's advantage of FT nmr.

RAPID SCAN NMR — In this technique, a spectral region is swept at a rate much greater than allowed for by slow passage conditions and the resulting ringing spectrum correlated with a spectrum of a single ringing line or of a theoretical line. Its principal advantage lies in the ability to acquire data rapidly even in the presence of strong solvent lines, without requiring that these lines be part of the signal averaged data.

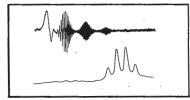
HOMOSPOIL T<sub>1</sub> SEQUENCE — This sequence allows the rapid measurement of long T<sub>1</sub>'s without the long 5xT<sub>1</sub> waiting period needed by the inversion-recovery sequence. It is accomplished by a -(90-spoil-tau-90-sample-spoil)- sequence.

QUADRATURE DETECTION NMR — This technique allows  $\sqrt{2}$  enhancement over conventional pulsed nmr since the rf carrier can be placed in the center of the spectral region and dual phase detectors used to receive the response. The two resulting free induction decays are Fourier transfermed using a complex transform which results in the frequency domain spectrum.

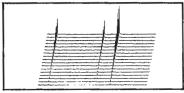
Please phone for details on these applications or to discuss how the vers. ...3-80 data system may be used for your experiments. Complete systems start at \$20,000. If you are already a Nicolet data system user, you may order free software for the above techniques from our Nicolet Users Society (NUS) library.



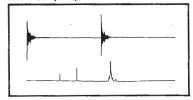
R. R. Ernst, J. Mag. Res. 3 10-27 (1970); R. Kaiser, J. Mag. Res. 3 28-40, (1970); J. Cooper and R. Addleman, 13th Experimental Nmr Conference (1972); E. Lippmaa, private communication.



J. Dadok and R. F. Sprecher, 13th and 14th Experimental Nmr Conferences (1972, 1973); E. Becker, paper in press.



J. L. Markley, W. J. Horsley and M. P. Klein J. Phys. Chem. 55 3604 (1971); R. Freeman and H. D. W. Hill, Ibid. 54 April (1971); G. G. McDonald and J. S. Leigh, Jr., J. Mag. Res. 9 358 (1973).



J. Schaeffer and E. O. Stejskal, 15th Experimental Nmr Conference (1974), and J. Mag. Res. March (1974) (in press). J. D. Ellett, etal. Adv. Mag. Res. 5 117 (1971)

### NICOLET INSTRUMENT CORPORATION









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# The University of Western Ontario, London, Canada

Faculty of Science Department of Chemistry April 24, 1974

Dr. B.L. Shapiro, Department of Chemistry, Texas A. & M. University, College Station, Texas 77843 U.S.A.

Dear Dr. Shapiro:

# POSITION AVAILABLE

I would like to draw to the attention of the readers of TAMU newsletter that the Chemistry Department, University of Western Ontario is beginning a search for a Senior Scientific Officer. I would be pleased to receive inquiries from anyone interested. A brief description follows.

Applications are invited for the post of Senior Scientific officer from persons familiar with the operation and scientific capabilities of spectroscopic instrumentation. The position involves responsibility for the maintenance and operation of major instruments in the Department with emphasis on n.m.r. and mass-spectroscopic facilities, together with an involvement in the professional or research activities of the Photochemistry Unit within the Department. The appointee will be a senior member of the support staff of the Department and should hold a Ph.D. or M.Sc. degree in Chemistry. The appointment is for 2 years in the first instance. Salary is negotiable, but not less than \$12,000. Applications should be sent to Dr. H.C. Clark, Head, Department of Chemistry, University of Western Ontario, London, Ontario N6A 3K7, as soon as possible.

Yours sincerely,

H.C. Clark, Head.

# Southern Research Institute

KETTERING-MEYER LABORATORY



2000 NINTH AVENUE SOUTH BIRMINGHAM, ALABAMA 35205 TELEPHONE 205-323-6592

April 15, 1974

Prof. B. L. Shapiro Department of Chemistry Texas A & M University College Station, Texas 77843

Dear Barry:

<sup>13</sup>C NMR of Isomeric Dithianes.

Most of our recent <sup>13</sup>C work has been on substituted purines, but this is scheduled to appear in the <u>Journal of Magnetic Resonance</u> very soon (July), and, in any case, is too lengthy for the newsletter. Meanwhile, here are some <sup>13</sup>C data on a pair of <u>cis-</u>, <u>trans-</u> isomers of a 2,5-disubstituted-1,3-dithiane to keep the subscription going.

Originally, we identified these isomers by means of their  ${}^1\mathrm{H}$  nmr spectra, two helpful references  ${}^1,{}^2$ , and  $\mathrm{Eu}(\mathrm{fod})_3$  addition. The  ${}^1\mathrm{H}$  chemical shifts of the methyl groups for both isomers are in agreement with that expected for an equatorial methyl group at  $\mathrm{C}_2$ , so that the predominant conformations should be as drawn.

Both completely decoupled and proton-coupled <sup>13</sup>C spectra were obtained for each isomer. The proton-coupled spectra made it possible to assign all peaks in the spectra of both isomers, and to obtain some of the coupling constants.

# Southern Research Institute

Prof. B. L. Shapiro April 15, 1974

# Chemical Shifts<sup>3</sup>

Isomer	$C_2$	$C_4(C_6)$	$\mathrm{C_5}$	$CH_3$	
cis-	41.82	38.62	38.72	${21.06}$	
<del>trans</del> -	41.00	37.89	47.69	20.00	

# Coupling Constants (Hz)

Isomer	$\frac{^{1}\mathrm{J}_{\mathrm{C_{2}H_{2}}}}{}$	$\frac{^{1}\mathrm{J}_{\mathrm{C_4}\mathrm{H_4}}}{-}$	$\frac{^3J_{\mathrm{C_4H_2}}}{-}$	$\frac{^{1}\mathrm{J}_{\mathrm{C_{5}H_{5}}}}{$	$\frac{^{1}\mathrm{J_{C_{Me}H_{Me}}}}{}$	$\frac{^{2}\mathrm{J}_{\mathrm{CMeH_{2}}}}{}$
cis- trans-	~ 153 ~ 150	~ 139 ~ 140	~7		$129.4 \pm 0.3$ $129.4 \pm 0.3$	$4.3\pm0.3$ $4.3\pm0.3$

As you will note, the only carbon with a significantly different chemical shift in the two isomers is  $C_5$ . In addition, in the proton-coupled spectra  $C_5$  showed different splitting patterns in the different isomers. In the cis- isomer,  $C_5$  appeared as doublet of triplets (being split by only one of the protons at  $C_4(C_6)$ ,  $^2J_{\begin{subarray}{c}C_5H_4\\\end{subarray}}=4.7\pm0.3).$  In the trans- isomer,  $C_5$  was split nearly equally by both protons at  $C_4(C_6)$ , and appeared as doublet of "quintets" with  $^2J_{\begin{subarray}{c}C_5H_4\\\end{subarray}}\approx4.5\mbox{Hz}.$ 

Sincerely,

Martha C. Thorpe Senior Chemist

Molecular Spectroscopy

### MCT:mw

- E. L. Eliel and R. O. Hutchins, J. Amer. Chem. Soc., 91, 2703-15 (1969).
- 2. R. J. Abraham and W. A. Thomas, <u>J. Chem. Soc.</u>, 335-41 (1965).
- Chemical shifts are in ppm downfield from internal TMS, and should be accurate within ± 0.02 ppm. Solvent: CDCl<sub>3</sub>.

WESTERN RESEARCH CENTER



# Stauffer Chemical Company

1200 South 47th Street / Richmond, California 94804 / Tel. (415) 233-9361

April 23, 1974

Professor B. L. Shapiro Department of Chemistry Texas A&M University College Station, Texas 77843

# Trifluoroacetic Acid as a Solvent

We wish to caution NMR researchers on the use of trifluoroacetic acid as a solvent.

In our attempt to obtain a carbon-13 NMR spectrum of 2-hydroxy-4-(ethylamino)-6-(isopropylamino)-triazine (I) we had to use trifluoroacetic acid as a solvent. For spectral comparison, we also used the same solvent for 2-chloro-4-(ethylamino)-6-(isopropylamino)-triazine (II). To our surprise, the two carbon-13 NMR spectra were identical.

I:X = OHII:X = C1

The chemical shifts with respect to TMS are listed as follows:

C(2), C(4), C(6): 145.2, 149.3, 150.2

C(7) : 33.0

C(8) : 7.7

C(9) : 41.6

C(10) : 15.7

Upon evaporating the solvent, both compounds gave identical IR and proton NMR spectra. Apparently trifluoroacetic acid contained water which reacted with II in the presence of acid to form I.

Sincerely yours,

C. K. Tseng

A. Mihailovski

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Dear Professor Shapiro,

# $T_1$ measurements showing convection streams.

In order to measure spin-lattice relaxation times of protons at low resolution we apply the usual fast adiabatic passage technique /1//2/, on a Varian HA-100 spectrometer operating in the H.R. mode. For example, phenyl selenol relaxation times bave been studied in the  $30^{\circ}\text{C}$  -  $150^{\circ}\text{C}$  temperature range. When oxygen is removed /2//4/ the  $T_1$  values of selenic proton /3/ are about 20 seconds so we can use a saw thooth generator with a period of 1.6 s to obtain the field sweep through the resonance.

Similar observations have been obtained with dimethyl sulfoxide and substances dissolved in this liquid.

Quantitative analysis. Let V be the total volume of the sample and v' be the fraction of this volume in which nuclei are adiabatically reversed. Furthermore we suppose that only a fraction of v' induce the bulk of the signal into the receiver coil. Let v. be this volume which is centered at the receiver coil level and corresponds to 1 cm of the sample vertical size. The signal in the receiver arises from two different species of nuclei : A part of it arises from the reversed spins which were initially in the volume v', let be  $p_1(t)$  the corresponding fraction ; an other part results from the non reversed spins which were initially outside the volume v' but came into the receiver volume v because they have pulled in by convection streams, let be  $1 - p_1(t)$  the corresponding fraction. Then,

<sup>/1/</sup> L.E. DRAIN, Proc. Phys. Soc. (London) 62 A, 301, (1949)

<sup>/2/</sup> J.C. DUPLAN, A. BRIGUET and J. DELMAU, J. Chem. Phys. 54, 3648, (1971)

<sup>/3/</sup> L. LALEZARI and N. SHARGHI, Spectroch. Acta <u>23A</u>, 1948, (1967)

<sup>/4/</sup> R.E. RONDEAU, J. Chem. Phys. Educ. 46, 504, (1969).

<sup>/5/</sup> J.W. EMSLEY, F. FEENEY, L.H. SUTCLIFFE, High Resolution N.M.R. Spectroscopy, First Edition, chapter 7, page 271, Pergamon Press (1965)

the total magnetization which influences the receiver coil may be written:

$$M_z(t) = p_1(t) M_z(t) + (1 - p_1(t)) M_o$$
 (1)

The evolution of  $M_{\chi}(t)$  is driven by the Bloch decay:

$$\frac{d}{dt} M_z(t) = \frac{M_o - M_z(t)}{T_1}$$
 (2)

Eq (1) and Eq (2) give:

$$M_z(t) - M_o = (M_z(t) - M_o)p_1(t)$$
 (3)

with the initial conditions :

$$M_{z}(0) = -M_{0}$$
 $p_{1}(0) = 1$ 
(5)

The behaviour of  $p_1(t)$  is reflected by the product  $(M - M_z(t))e^{t/T}1$  since  $T_1$  and  $M_0$  are known and the variations of  $M_z(t)^0$  are recorded.

Motion of "labeled" nuclei : The experimental curves shown in Fig.2 have been obtained at 130°C with phenyl protons signals of C6H5SeH. The full lines represents the decay of (M - M(t))/M and they correspond to the evolution of the magnetization of the volume v after the spins have been reversed in the volume  $v^*$ . From the preceding curves we can the decay of  $p_1(t)$  by multiplying by  $e^{t/T_1}$  \* - dashed lines. We have obtained theses results using decreasing liquid heights in 5 mm o-d N.M.R. tubes successively 45,25 and 15 mm. The tubes are identically fixed in the spinner so the surface of the liquid has not the same position and v' is not constant.

Of course the quantity (M  $_{\rm c}$  - M  $_{\rm c}$ (t))/M tends toward zero, but this limiting value is reached only after several oscillations. One can reasonably assume that the spins "labeled" in the volume v' at time zero will spread over the whole sample volume V after a long enough time. Then the value of  $p_1(t)$  tends to the ratio  $v^*/V$  which is a non zero value. This prediction is well tested by the experiment.

A close relationship exists between the period of the observed oscillations and the convection motions. These streams appear well regulated. But in course of time the streams tubes get out of shape because the mixing of the liquid and the amplitude of the oscillations of p, (t) decrases.

Cautions in high temperature N.M.R. studies. Since this parasite effect must be carefully avoided during relaxation times measurements we though it would be useful to summarize some remarks about how to avoid it. Care must be taken even for spinning samples because the inertia introduced by rotations prevents the convection streams, it also hinders the homogeneization of the temperatures amid the samples.

We also looked at the effects of the convection currents on

<sup>\*</sup> The T,'s value is independently measured by the adiabatic method after the gradient has disappeared.

saturation recovery experiments /6/. The recovery of the apparent magnetization is in that case nearly exponential but with a time constant shorther than the usual  $T_1$ 's value. Indeed, the convection motions alter the initial conditions of the different spins; those which are not in the volume  $v^*$  at zero time, have soon recovered some magnetization.

Finally we would like to point out that our observations provide further support to the idea that magnetic labeling of the nuclei is a powerful method to reveal internal motions in small liquid samples /7/.

Sincerely yours.

Menhor

A. BRIGUET

J.C. DUPLAN

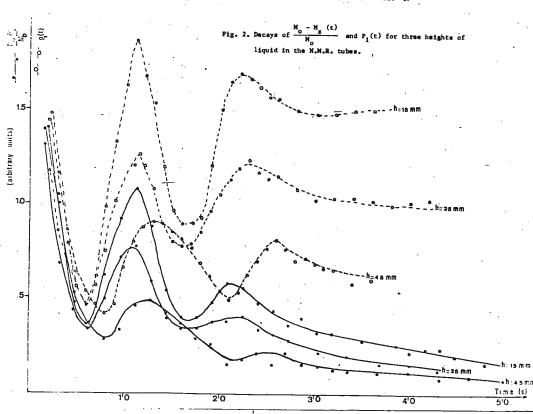
J. DELMAU

Fig. 1. Magnetization recovery of the phenyl protons in purum C.H.SeH





non spinning sample (h = 60 mm, Température ~ 130 °C)



/6/ A.L. VAN GEET and D.N. HUME, Anal. Chem. 37, 979, (1965)

ibid <u>37</u>, 983, (1965)

/7/ J. PACKER, Mol. Phys. 17, 355, (1969)

# MONTEDISON

Prof. B.L. Shapiro,
Texas A & M University,
College Station, Texas 77843
Dpt. of Chemistry U S A

27 march 1974

BLS/lmk

DIPE/CER

Dear Prof. Shapiro:

just now i have received your letter of the 25<sup>th</sup> february and I have learnt that I will go on receiving TAMU NMR Newsletter. I have already received the february issue and I hope you will be able to send me the back numbers from october 73'. In the mean time I send you the present contribution.

Subject: <sup>19</sup>F NMR analysis of fluoride of perfluorocyclo-hexanesulphonic acid and of 3-trifluoromethyl-perfluorocyclopentane-l-sulphonic acid.

From the electrochemical fluorination of the chloride and fluoride of the benzensulphonic acid, beside the main product perfluorocyclohexane sulphonic acid  $C_6F_{11}SO_2F$  (I), also the side products 3-trifluoromethylperfluorocyclopentame-l-sulphonic acid  $CF_3C_5F_8SO_2F$  (II) and  $C_6F_{10}C1SO_2F$  were obtained.

We have examined with accuracy the values of the different chemical shifts and coupling constants of I and II.

(II) is constituted of two geometric isomers cis (IIa) and trans (IIb). Near the cis-trans isomerisation phenomenum, for each of the two geometric isomers, two conformers in mutual equilibrium are found: (1),(2) (see table 1). For the cis isomer the two conformers in equilibrium are: (1) the pseudo-axial-axial conformer and (2) the pseudo equatorial-equatorial conformer (1) and form (1) is the overwhelming  $(n_1 \gg n_2)$ . For the trans isomer the two conformers are: (1) the pseudo-axial-equatorial and (2) the pseudo-equatorial-axial conformer and in this case the distribution of the populations is  $n_1 \cong n_2$ .

The attributions of <sup>19</sup>F MMR spectra of (1) were conducted in strict analogy with the Sutcliffe analysis of perfluoromethylcyclohexane  $(CF_3C_6F_{11})^{(2)}$ .

In table 1 the structure formulae of (I), (IIa) and (IIb) with the labelling of the fluorine atoms are reported.

The chemical shifts were measured relatively to CFCl, and expressed in ppm according to the  $\delta$  scale. The coupling constant are given in Hz but the sign was not determined.

In the next tables the small letters e, a are used for the equatorial and respectively axial fluorine atoms.

In table 2 the chemical shifts (  $50\% \text{ v/v CFCl}_3$  solutions) and some coupling constants of (I) are reported.

In table 3 the chemical shifts and the geminal coupling constants of (IIa) and (IIb) are reported (same concentration).

We hope to be able to send within few months the present work to Organic Magnetic Resonance Journal with further explanations and details.

We are indebted to Prof. L. Cavalli of "Società Italiana Resine, Laboratori Analisi", for the helpful discussions, we had during his former activity at the "Centro Ricerche of Bollate" (Montedison SpA), and to Prof. G. Troilo of the "Istituto di Chimica Industriale-Facoltà di Ingegneria-Università di Padova" for the preparation and purification of the products .-

Yours sincerely

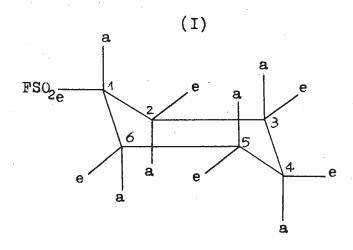
Giorgio Gurato (Giorgio Gurato)

(Giorgio Zenoni)

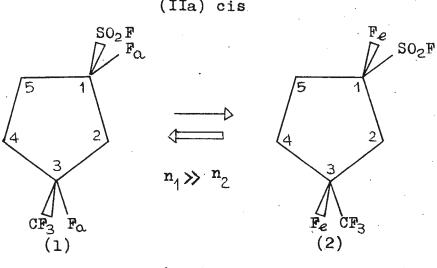
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Table 1



(IIa) cis



(IIb) trans

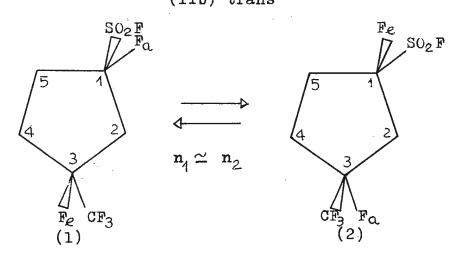


Table 2

#### NMR data of (I)

```
Chemical shifts (ppm)
                              Coupling constants (Hz)
ഗ്(le)
           = +63.5
                           J(2a-2e)=J(6a-6e) = 295 \pm 0.5
                           J(3a-3e)=J(5a-5e) = 287 \pm 3
\delta(2a,6a) = -117,1
                                               = 288 \pm 3
\delta (3a,5a) = - 122.4
                           J(4a-4e)
\delta(4a)
         = -123.7
\delta(2e,6e) = -129.9
\delta (3e,5e) = -139.0
\delta (4e) = -141.7
\delta(la) = - 165.8
```

#### Coupling constants related to 1a band (-CF)

$$J(1a-3a,5a) = 23,5$$
  
 $J(1a-2e,6e) = 15.9$   
 $J(1a-2a,6a) = 4.2$   
 $J(1a-3e,5e) = 2.1$   
 $J(1a-SO_2F) = 2.1$   
 $J(1a-4a,4e) = 0.0$ 

Coupling constants related to  $-SO_2F$  band  $J(SO_2F-2a,6a) = 16.7$   $J(SO_2F-2e,6e) = 7.8$   $J(SO_2F-1a) = 2.1$   $J(SO_2F-3e,5e) = 2.2$   $J(SO_2F-3a,5a;4a,4e) = 0.0$ 

Coupling constants related to 4e band 
$$J(4e-3e)=J(4e-5e)=14.1$$
 or 12.7  $J(4e-3a)=J(4e-5a)=12.7$  or 14.1  $J(4e-2e)=J(4e-6e)=6.4$   $J(4e-2a)=J(4e-6a)=1.6$ 

Table 3

#### NMR data of (IIa) and (IIb)

#### Chemical shifts (ppm) (IIa) cis (IIb) trans $\delta(SO_2F) = + 59.18$ $\delta(SO_2F) = + 59.50$ $\delta_{AV}(CF_3) = -72.66$ (cis+trans) $\delta(2a)$ $\delta(2a)$ = -116.52= - 113.99 $\delta(4a)$ = - 119.14 $\delta(2e)$ = -116.83= - 123.90 $\delta(5a)$ o (5a) = - 120.35δ(2e) = -123.230 (4a) = - 123.21δ(4e) = - 131.76 $\delta$ (5e) = -123.41∂(5e) = -132.32o(4e) = -125.61δ(la) = -163.41 $\delta(1a,e) = -163.99$ $\delta(3a)$ = - 183.87 $\delta(3e,a) = -185.50$

#### Geminal coupling constants J(e,a) (Hz)

J cis	J trans						
$J(2c) = 283.80 \pm 0.1$	$J(2t) = 276.50 \pm 0.3$						
$J(4c) = 268.95 \pm 0.25$	$J(4t) = 268.48 \pm 0.55$						
$J(5c) = 274.35 \pm 0.15$	$J(5t) = 270.60 \pm 0.10$						

#### UNIVERSITÉ D'OTTAWA



#### UNIVERSITY OF OTTAWA

DÉPARTEMENT DE CHIMIE

DEPARTMENT OF CHEMISTRY

April 18, 1974.

Title: The Conformation of N-nitroso-r-2-cis-6-dimethyl-4-phenylpiperidine,  $\underline{I}$ ,

Professor B.L. Shapiro, Dept. of Chemistry, Texas A and M University, College Station, Texas 77843.

Dear Barry:

We wish to describe the results of a computer analysis of the 13-spin system giving rise to the 220 MHz spectrum of the N-nitrosopiperidine derivative, I. Comparison of the vicinal coupling constants with similar parameters in the literature provides evidence that the compound possesses a distorted chair conformation in which the two methyl groups are axial, e.g.  $H_{\sim}^{\sim}$ 

PHE

Compound I was prepared by two successive methylations of N-nitroso-4-phenylpiperidine. Since the 100 MHz spectrum of I was exceedingly complex the 220 MHz spectrum was run for us by Dr. A. Gray, Ontario Research Foundation, Toronto. A portion of the spectrum showing all but the aromatic and methine proton absorptions appears in Fig. 1A. In order to obtain the spectral parameters for the seven protons attached to the piperidine ring the LAOCN3 program was used. Iterative fit by computer was not used as the matching of assignments was complicated by broadening due to unresolved long-range couplings. Each calculated spectrum was compared to the experimental and another calculation made with adjusted parameters until acceptable agreement was obtained. The calculated spectra included a first order perturbation by the two methyl groups on both the vicinal and the next nearest protons. Agreement was deemed acceptable for correspondence in line positions of <0.1 Hz. The parameters which give the calculated spectrum shown in Fig. 1B appear in the Table.

It can be seen that coupling between protons at carbons 3,4 and 5 are typical for protons on a cyclohexane ring. However couplings between protons on carbons 2 and 3 (and 5 and 6) are not normal for a piperidine ring i.e.  $J_{ae}$  should be 3-3.5 Hz and one can estimate, by considering effects of electronegativity and their orientation dependence, a value for  $J_{ee}$  of 2.2 Hz. The observed  $J_{ae}$ 's (6.3, 6.5) and  $J_{ee}$ 's (1.5) indicate the presence of a distortion due to the severe repulsive 1,3-diaxial methylmethyl interaction. Should this distortion produce a  $10^{\rm O}$  increase in the dihedral angle between the two equatorial protons  $(\phi_{ee})$  and a concomitant increase in  $\phi_{ea}$ , the predicted

values for  $J_{ea}$  and  $J_{ee}$  would change to 5.2 and 1.0 Hz. Thus a distortion of this type can account, at least qualitatively, for the observed vicinal coupling constants.

Proton	Shift			Coupling Constants to Other Protons <sup>a</sup>						
	(Hz from TMS at 220 MHz)		•				•			
****		2	3e	3a	4	5e	5 <b>a</b> -	6	СН3	
2	1143.5		1.5	6.5				1.0	7.0	
3e	402.5			-13.7	3.2	2.3			0.2	
3a	390.9				13.0				0.3	
4	733.0					3.2	12.6			
5e	442.2						-13.3	1.5	0.2	
5 <b>a</b>	467.4	,						6.3	0.5	
6	1150.5								7.0	

a) effects of unresolved long range couplings were simulated by using a linewidth of 1 Hz for the print out in Fig. 1B.

The conclusion that the title nitrosamine exists in a distorted chair conformation having two methyl groups axial substantiates the earlier proposals of Harris and Spragg<sup>3</sup> and Chow and Colon<sup>4</sup> that the nitrosamine of  $\underline{cis}$  2,6-dimethylpiperidine exists "predominantly" in a chair conformation having both methyl groups axial. Furthermore the parameters obtained for  $\underline{l}$  rule out the presence of boat conformations to greater than a few percent.

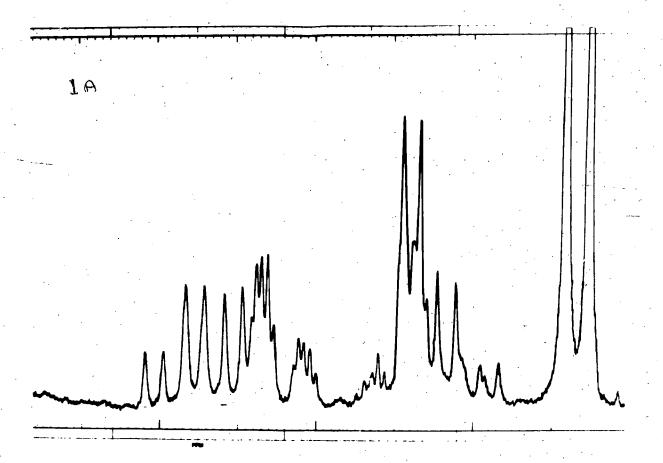
Best regards,

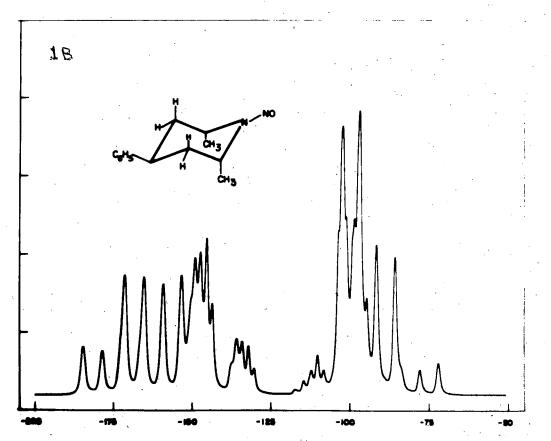
R.R. Fraser,

Prof. of Chemistry.

#### RRF/ma

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#### **DEPARTMENT OF CHEMISTRY**

April 26, 1974

Dr. Bernard L. Shapiro
Department of Chemistry
Texas A and M University
College Station, Texas 77843
U.S.A.

Dear Barry:

#### Spectra of Internal Rotational Isomers in a

#### Chiral Solvent

We have determined the proton spectra of some aryl substituted hydantoins, which are known to have slow rates of internal rotation at normal probe temperatures, in the chiral solvent (+)-phenyltri-fluoromethylcarbinol. Under achiral conditions, the rotational isomers of these compounds are enantiomeric.

Since, in the chiral solvent, the solvated rotamers should bear a diastereomeric relationship to each other, it is expected that they should show distinguishable nmr spectra. We were interested in the extent to which the signals from corresponding nuclei in the two rotamers would be split.

Ι

IIa Ar = 2-methy1
IIb Ar = 2,3-dimethy1

IIc Ar = 2, 4-dimethy1

IId Ar = 2,5-dimethy1

Dr. Bernard L. Shapiro

We find that the extent of splitting of signals is strongly dependent on the location of the nuclei in these molecules. Thus, in the 3-aryl hydantoin, I, only the ortho methyl signals are resolvable (0.05 ppm), the 5,5-dimethyl signals being unaffected. Other 3-aryl hydantoins which were examined also failed to show additional splitting in the chiral solvent.

In the 1-ary1 series (II) the 5,5-dimethy1 proton signals are slightly broadened in the chiral solvent, while the ortho ary1 methy1 signal is split (0.12-0.13 ppm) in all cases examined. In the 2,3-dimethy1pheny1 compound (IIb) the meta methy1 signal is split by 0.12 ppm, whereas in the 2,5-dimethy1pheny1 compound (IId) the meta methy1 signal is split by 0.06 ppm. In contrast, the para methy1 signal of the 2,4-dimethy1 compound (IIc) is unsplit.

The known solvating behaviour  $^2$  of the solvent suggests that solvation should occur by hydrogen bonding to the carbonyl groups of the hetero ring. A feasible explanation for the observed chemical shift differences between the diastereomeric solvates may be based on a model involving a secondary interaction between the  $\pi$  electron systems of the solute and solvent molecules such that the maximum anisotropy of the solvent phenyl group is experienced by the nuclei in the region of the solute aryl group. Thus the 5,5-dimethyl groups are affected in the 1-aryl but not the 3-aryl series. The lack of splitting of a para methyl group (IIc) may result from a similar relationship with the solvated solvent phenyl groups in the two rotamers.

Best regards,

Yours sincerely,

auric.

L.D. Colebrook F.H. Hund S. Icli

W.H. Pirkle, J. Amer. Chem. Soc., <u>88</u>, 1837 (1966).

<sup>&</sup>lt;sup>2</sup> W.H. Pirkle and S.D. Beare, ibid., <u>91</u>, 5150 (1969).

#### KEMISK INSTITUT

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LABORATORIET FOR ORGANISK KEMI HANS JØRGEN JAKOBSEN 8000 Århus C, den April 30, 1974 Telefon (06) 124633 HJJ/ATL

Professor Bernard L. Shapiro Department of Chemistry Texas A & M University COLLEGE STATION, Texas 77843 USA

Dear Professor Shapiro:

## Selective Population Transfer (SPT) in FT NMR. A Useful Technique Equivalent to TSI in CW NMR

In connection with our interest in analysis of undecoupled  $^{13}$ C FT NMR spectra, we would like to inform your readers of a technique that we have applied in numerous cases for assignment and relative sign determination of  $^{13}$ C-X coupling constants in  $^{13}$ C spectra [1]. Furthermore, in molecules with degenerate proton transitions (e.g.  $AX_n$  spin systems;  $A = ^{13}$ C,  $X = ^{1}$ H), the method may be especially useful for sensitivity enhancement [2].

The SPT technique is equivalent to the transitory selective irradiation (TSI) experiment in CW NMR [3], i.e. a specific transition in the 1H satellite spectrum is perturbed in a manner so as to approach inversion of the corresponding energy level populations. Thus in a 13C FT experiment a weak irradiating field  $\underline{H}_2$  field  $(\gamma \underline{H}_2/2\pi = 0.1-0.5 \text{ Hz})$  is applied to one of the <sup>1</sup>H transitions during a time  $\tau$ , given by  $YH_2\tau = \pi$  (i.e.  $\tau = 1-5$  sec), immediately before probing the <sup>13</sup>C intensity change by excitation with a strong  $\underline{H}_1$  rf pulse. Almost complete population inversion may be obtained for molecules where  $T_1 >\!\!\!> \tau$  ( $T_1$ is the shortest spin-lattice relaxation time for all connected transitions). In a repetitive  $\underline{H}_1$  pulse experiment the  $\underline{H}_2$  field is simply gated as described by Freeman and Hill [4]. This pulse sequence has also recently been applied by Pachler and Wessels to the  $^{1:3}$ C spectrum of chloroform [5]. The SPT  $^{1:3}$ C spectra in figures 1 and 2 serve to illustrate the application of the technique for assignment and relative sign determination of 13C-X couplings, and for sensitivity enhancement in 13C FT NMR, respectively.

For comparison with first order spectral intensity distributions arising from ordinary equilibrium populations (i.e. the binomial coefficients), we have calculated (fig. 3) the relative intensities for the A transitions in  $AX_n$  (A =  $^{13}$ C, X =  $^{1}$ H) spin systems immediately after the SPT technique has been applied to one of the lines (degenerate transitions) in the X doublet. This has been performed assuming complete population in-

version between the levels for the degenerate transitions and using  $\gamma_H/\gamma_C = 4$ . As seen in figure 2 the relative intensities in the ënhanced spectra are in good agreement with those predicted (fig. 3).

A few preprints (refs. [1] and [2]) are available.

Sincerely yours,

Hans J. Jakobsen J. J. Horsen

H. J. Jakobsen R. S. Hansen

S. Aa. Linde S. Sørensen

- [1] S.Sørensen, R.S.Hansen and H.J.Jakobsen, J.Magn.Resonance, 1974.
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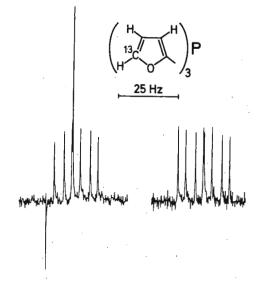


Figure 1. 25.16 MHz <sup>13</sup>C SPT NMR spectrum (Varian XL-100-15 spectrometer, S124-XL Fourier transform accessory, Varian 620L 16K computer) of the C5 carbon in tris-2-furylphosphine; high frequency (low field) part is at the left and low frequency (high field) part at the right. The spectrum was obtained by application of the SPT technique to the highest frequency line in the H4 satellite spectrum,  $\gamma \underline{H}_2/2\pi = 0.25 \text{ Hz}$  and  $\tau = 2.0$  sec. The experiment shows that

$$^{2}J_{C5-H4}$$
 >  $^{3}J_{C5-H3}$ ,
 $^{1}J_{C5-H5}$  x  $^{3}J_{H4-H5}$  > 0,
 $^{3}J_{C5-H3}$  x  $^{3}J_{H3-H4}$  > 0
(i.e.  $^{3}J_{C5-H3}$  > 0), and
 $^{3}J_{C5-P}$  x  $^{4}J_{H4-P}$  > 0

(i.e.  $J_{C5-P} > 0$ ).

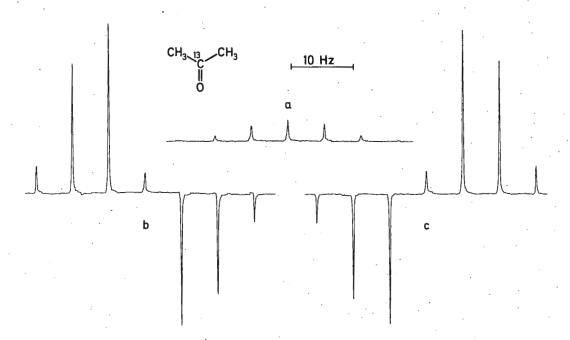


Figure 2. <sup>13</sup>C NMR spectra of the carbonyl carbon atom in acetone  $(^2J_{CCH} = 5.92 \text{ Hz})$ . (a) Normal undecoupled spectrum. (b) and (c) SPT spectra obtained by gated irradiation of the low (b) and high (c) frequency line in the <sup>1</sup>H inner satellite doublet spectrum,  $(^{2}H_{2}/^{2}\pi = 0.45 \text{ Hz}$  and  $(^{2}H_{2}/^{2}\pi = 0.45 \text{ Hz})$  and  $(^{2}H_{2}/^{2}\pi = 0.45 \text$ 

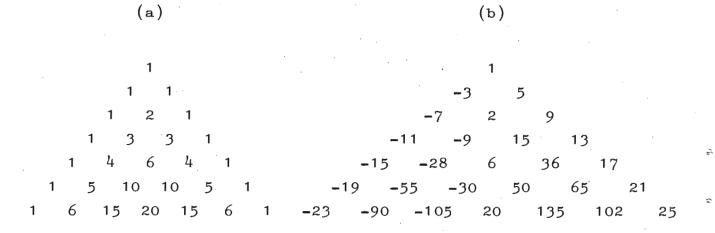


Figure 3. Relative intensities for the A spectrum of  $AX_n$  (A = <sup>13</sup>C and X = <sup>1</sup>H) spin systems. (a) Relative intensities corresponding to the populations at thermal equilibrium (Pascal triangle). (b) Relative intensities corresponding to complete inversion of the equilibrium populations for the degenerate transitions due to one of the lines in the X doublet  $(\gamma_H/\gamma_C = 4)$ .



### AUTHOR INDEX

Briguet, A	•							31
Clark, H. C.		•			•			25
Clerc, J. T.				•				16
Colebrook, L. D			•			•	•	42
Delmau, J								31
Duplan, J. C.								31
Ellis, P. D.		•						10
Fraser, R. R.								39
Garst, J. F.							•	15
Gurato, G								34
Hansen, R. S.								44
Hund, F. H			•					42
Icli, S								42
Jakobsen, H. J.								44
ichter, R. L.					• .			.9
inde, S. A.								44
Martin, G. J.			.•				•	12
Martin, M. L.			•		•			12
Meier, H. P.								16
Mihailovski, A.								28
Mondelli, R.								3
Musher, J. I.								5
Pines, A								1
Pretsch, E								16
Ray, J								6
Santini, B								6
Sardella, D. J.								18
Sørensen, S.								44
Thorpe, M. C.								26
Tseng, C. K.								28
Wehrli, F. W.								20
Zenoni G	-	•	•	•	•	•	-	34



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