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Newsletter

No. 144  
SEPTEMBER, 1970

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All Newsletter correspondence, etc., should be addressed to:

Bernard L. Shapiro  
Department of Chemistry  
Texas A&M University  
College Station, Texas 77843

# UNIVERSITY of PENNSYLVANIA

PHILADELPHIA 19104

## *The School of Medicine*

JOHNSON RESEARCH FOUNDATION  
DEPARTMENT OF  
BIOPHYSICS AND PHYSICAL BIOCHEMISTRY

August 10, 1970

Professor B.L. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station, Texas 77843

Dear Professor Shapiro,

Thank you for your blue note. It is not always easy to find something relevant to the wide audience of TAMUNMRN, but I hope the following will do.

The use of Mn(II) as a paramagnetic probe for studying the binding of divalent ions to biological macromolecules is well documented. The main feature is that as a result of the binding the longitudinal proton relaxation rate (PRR) of water is usually enhanced and measurements at very low concentrations of the substance of interest are feasible. For trivalent ions, however, a suitable probe has yet to be found since the binding of Fe(III) leads to a decrease of the PRR rather than to an enhancement.

We have studied the effects of binding on the PRR due to gadolinium (III). For this preliminary investigation bovine serum albumin (BSA, M.W. 69000) was chosen since it is known to bind metal ions. It was found that bound Gd(III) is about 4.5 times more effective in relaxing the water protons than free Gd(III). It was also found (from a protein titration) that BSA has 4 binding sites for Gd(III) with a dissociation constant of approximately  $5 \times 10^{-5}$  M. It appears that Gd(III) may be a very useful probe for studying the binding of trivalent ions to biological macromolecules.

Sincerely yours,



Jacques Reuben

Title: Gadolinium(III) as a probe for studying binding of trivalent ions to biological macromolecules by NMR

**Glaxo**

Glaxo Research Ltd, Greenford, Middlesex

Telephone: 01-422 3434  
Telegrams: Glaxotha, London  
Telex: 22134  
Code: New Standard, Bentleys

Please address reply to:

4th August, 1970.

Prof. B.L. Shapiro,  
Texas A & M University,  
Texas.

Dear Professor Shapiro,

Sensitivity Test for N.M.R. Spectrometers

We are interested in comparing the sensitivities of n.m.r. spectrometers produced by different manufacturers and have observed that although makers normally specify the minimum signal-to-noise ratio for a 1% (v/v) solution of ethylbenzene in carbon tetrachloride, the conditions specified for conducting the test vary considerably. Often, no limit is set on the depth or volume of solution in the spinning-tube and, hence, on the quantity of ethylbenzene. The minimum, routinely practical, depth of solution needed for a standard 5 mm spinning-tube varies from one instrument to another. It is clear that the facility to use a smaller depth of solution represents an effective increase in sensitivity.

We suggest that a standard weight rather than a concentration should be used in the manufacturer's specification and, further, we believe that a crystalline solid, which can be easily weighed, provides a better standard than a liquid, such as ethylbenzene.

We have conducted tests on several Varian, Perkin-Elmer and Jeolco 60 MHz spectrometers, in which we measured in the conventional way the signal-to-noise ratio for the largest peak of the AB quartet formed by the 21-methylene group of a 7 mg sample of Reichstein's D diacetate (3 $\beta$ ,21-diacetoxy-17 $\alpha$ -hydroxy-5 $\alpha$ -pregnane-17,20-dione) dissolved in the minimum volume of CDCl<sub>3</sub> needed to give a satisfactory signal with the particular instrument. Standard 5 mm spinning-tubes were used and the spectra were recorded at a standard sweep rate of 1 Hz/sec. or at the nearest equivalent rate; the R.F. power and filtering of each instrument were adjusted for optimum performance. Reichstein's D diacetate was selected, since it is readily soluble in CDCl<sub>3</sub>, gives a suitable AB quartet and, above all, happened to be in the laboratory cupboard at the right moment.

Contd./

- 2 -

We now realise that Reichstein's D diacetate is not generally available in n.m.r. laboratories and suggest that the highest peak of the methylene quartet shown by phenacetin in  $\text{CDCl}_3$  solution (cf. Varian Spectrum Catalogue, 1962, No. 267) be used instead. Phenacetin is available commercially in an analytical grade; 5 mg appears to be a suitable standard quantity.

We would like to know whether other readers have had thoughts on standard sensitivity tests for commercial n.m.r. spectrometers and would like to hear of their experiences.

Yours sincerely,

J.E. Page

R.A. Fletton

G.F.H. Green

James E Page

R.A. Fletton

G.F.H. Green

**digital**

August 10, 1970

Dr. Bernard Shapiro  
Department of Chemistry  
Texas A & M University  
College Station, Texas

"Simplified Signal Averaging with the PDP-8/I"

Dear Barry:

With this letter I wish to start my subscription to TAMUNMR. Bob Kurland described his instrumental set-up for  $^{13}\text{C}$  signal averaging in the June issue, and I would like to describe the computer program we developed for use with this instrumentation.

The basic method is that of computer control of the experiment rather than of occasional interaction of two instruments. The PDP-8/I computer has available with it an interface called the AX-08 which contains analog to digital and digital to analog converters (ADC's and DAC's). By loading the DAC with a number between 1 and 512 a voltage is produced which can be used to drive, for instance, a scope display. We used this voltage to drive the Fluke digital frequency synthesizer, model 633A in external sweep mode. This meant that for each input voltage there was a discrete, reproducible frequency which could be applied to the spectrometer.

Each time the voltage was advanced in this manner, the computer program examined the ADC input from the spectrometer, and added it into memory. We found that we were able to increase our S/N markedly by sampling each of these 512 points several times (up to 20) during each sweep. This method, known as boxcar averaging, really amounts to 20 sub-sweeps during a single sweep. It has been deprecated by others but we found it to be of great value in our proton and  $^{13}\text{C}$  experiments.

The upshot of this instrumental method is that there is really no difference between the spectrometer and computer sweeps as a function of time, since they are one in the same. Thus, using the DAC's to drive a plotter, while still connected to the

cont.

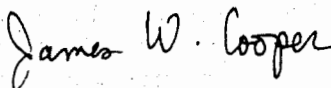
Dr. Bernard Shapiro  
August 10, 1970

synthesizer, we were able to plot out the accumulated spectrum, adjust the plotter pen to hover over any line of interest, and read the line's exact frequency from a counter.

This method seems to be far superior to the more common approach where the computer and spectrometer sweep concurrently, but asynchronously, after the triggering by a sync pulse. In this latter method, the nonlinearity of the two sawtooth sweeps may lead to disastrous errors in spectrum calibration.

As you can see from the letterhead, I have finished my post-doc with Dr. Kurland and am now with Digital Equipment Corporation, where I am still involved in this same area of work. I'll be glad to send further details on this method of signal acquisition to any interested parties.

Sincerely,



James W. Cooper  
Applications Programmer  
NMR and Related Chemical Applications

JC/jb

The University of Oklahoma

620 Parrington Oval, Room 221

Norman, Oklahoma 73069

August 10, 1970

Professor Bernard L. Shapiro  
 Department of Chemistry  
 Texas A & M University  
 College Station, Texas 77843

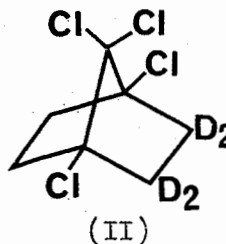
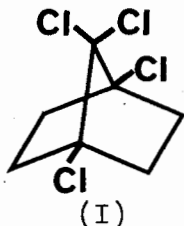
Dear Dr. Shapiro,

LINEAR FREE ENERGY CORRELATIONS OF NMR CHEMICAL SHIFT PHENOMENA

Your pink notice catches me on the short side of completion of a study involving ~~nmr~~ investigation of solute-solvent interactions. In the hope of maintaining our TAMUNMR Newsletter subscription, I offer the following progress report as a pacifier:

Some time ago (IITNMR Newsletter, 108, 2, September, 1967) we reported our observations regarding solvent effects on the ~~nmr~~ spectrum of 1,4,7,7-tetrachloronorbornane, (I). In particular, we observed that the appearance of the  $A_4B_4$  spectrum of I in benzene was quite different from that in pyridine; the spectra have since been published, (J. Org. Chem., 34, 1109 (1969)). Compound I possesses a pronounced molecular dipole moment ( 3.30 D ). This led us to wonder if in fact the difference between the appearance of the spectra of I obtained in these two solvents might be due to a specific solvation interaction. One might envision this as an interaction between the lone pair on the pyridine nitrogen and the positive end of the molecular dipole associated with I. This effect, of course, would be absent were pyridine replaced by benzene as solvent.

Our recent work leads us to believe that the type of interaction described above is probably not responsible for the effects which we observe. In particular, we have calculated the chemical shift and coupling constant parameters ( via LAOCOON III ) for the AA'BB' spectrum of deuterium-decoupled II, below, in several monosubstituted benzene solvents, (PhX, where X = halogen, CN, NMe<sub>2</sub>, NO<sub>2</sub>, CF<sub>3</sub>, OMe, CH<sub>3</sub>, CHO, -C≡CH, CH<sub>2</sub>CN).



Department of  
Chemistry



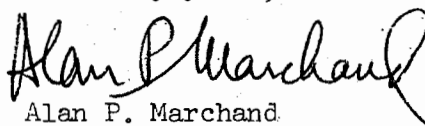
Letter to Professor B. L. Shapiro  
August 10, 1970

We then attempted to correlate the difference in chemical shifts,  $\Delta\delta$ , between the upfield and downfield halves of the centrosymmetric AA'BB' pattern with Hammett  $\sigma$  values. Acceptable correlations have been obtained. Interestingly, electron donating substituents, X, in PhX describe one line, and electron withdrawing substituents fall on another correlation line, suggesting that fundamentally different interactions are involved in the collision complexes formed between II and these two different types of monosubstituted benzenes.

Another pertinent observation has emerged: the spectrum of II in pyridine is essentially identical to that of II in nitrobenzene! This suggests that the effect of the nitrogen atom on the spectrum of II in pyridine is due mainly to the enhanced electronegativity of nitrogen relative to carbon. (In nitrobenzene, a "ring nitrogen atom" is approximated in electronegativity by the ring carbon atom to which the highly electronegative NO<sub>2</sub> substituent is attached).

We therefore look to electronic effects (and not to lone pair- dipole association effects) to explain our original observations. Work currently in progress is aimed toward delineating the relative contribution of inductive and resonance effects of the substituent X in the solvent PhX to the overall solvent effect upon  $\Delta\delta$  in the nmr spectrum of II. This work is being carried out in collaboration with Dr. Anna Segre De Angelis (Mellon Institute, Carnegie-Mellon University, Pittsburgh, Pennsylvania, on leave from the Istituto di Chimica delle Macromolecole del C. N. R., Milan, Italy) and Dr. Arthur M. Ihrig (Department of Chemistry, Texas Christian University, Fort Worth, Texas).

Sincerely yours,



Alan P. Marchand  
Associate Professor



## ARGONNE NATIONAL LABORATORY

July 31, 1970

Professor B. L. Shapiro  
Department of Chemistry  
Texas A and M University  
College Station, Texas 77843

Automatic Liquid Nitrogen Delivery Line for HR-220

Dear Barry:

The Varian HR-220 NMR Spectrometer System uses a cryostat which contains liquid helium (LHe) and liquid nitrogen (LN). The super-conducting solenoid is immersed in LHe and the jacket surrounding the LHe is kept filled with LN. It is desirable to maintain a high, constant level of LN in the outer dewar to keep the LHe loss to a minimum. An automatic LN filling system has been designed to deliver a predetermined quantity of LN. The system has the following operating parameters: First, an adequate supply (160 l), available under approximately 12 lbs. pressure. Second, adequate pressure relief valves, properly installed, to relieve pressure built up in the transfer lines when the solenoid valves are closed. Third, well-insulated transfer lines to conserve LN.

The LN supply is stored at a distance of 12 feet from the cryostat. The total length of the transfer line is approximately 22 feet. The transfer line is constructed of 3/8" stainless steel pipe and insulated with 5/8" O.D. rubber tubing together with 5/8" x 1 5/8" polystyrene type insulation. Connections from the transfer line to the dewar are made with 1/2" x 5/8" pure gum tubing and secured with clamps. The connection to the 160 l LN supply is made with 3/8" stainless steel flexible tubing and secured with flare fittings to insure a good mechanical connection. The problem of pressure in the transfer line after filling was solved by using conventional pressure relief valves; one is set at 18 lbs., and another set at 20 lbs. The two valves are mounted on a 10 inch heat riser located near the LN supply. The other closed loop of the transfer line is protected by the two relief valves furnished with the instrument which are set to release at approximately 1 lb. The thermistor probe is located as close as possible to the dewar so the maximum length of transfer line can be pre-cooled.

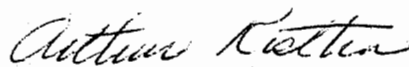
-2-

The levels of LN inside the dewar determine the points at which the system turns on and off. This is controlled by the LN sensor unit. The turn-on/off points are adjustable. When the turn-on level is reached, relay  $K_1$ , located in the HR-220 console, is energized. This causes relay  $K_2$  to energize. The contacts of relay  $K_2$  permit 110V AC to energize solenoid valves (1) and (2). These normally closed valves open and allow the LN to pre-cool the transfer line. Relay  $K_3$  is energized because of the low resistance of the thermistor probe, which is located in the transfer line near solenoid valve (3). When the transfer line is cooled, and LN begins to flow, the thermistor probe causes relay  $K_3$  to de-energize, which in turn causes solenoid valve (2) to close and solenoid valves (3) and (4) to open. The dewar then begins to fill with LN and this continues until the sensor unit causes relay  $K_1$  to de-energize. In turn,  $K_2$  de-energizes and solenoid valves (1), (3) and (4) close. Relay  $K_3$  will again energize as soon as the resistance of the thermistor allows  $Q_1$  to conduct sufficient current.

Timer C, energized when the system turns on, provides a timed safety factor should a malfunction such as a broken transfer line, leaky connections or lack of LN occur. If the timer times out before completion of the filling operation, its relay contacts open, causing relay  $K_4$  to come on and solenoid valves (1), (2), (3), and (4) to turn off. At the same time, relay  $K_4$  causes an alarm to sound. The alarm can be turned off by  $S_1$  or  $S_3$ .

This system has been used for more than a year and has been found to be highly reliable.

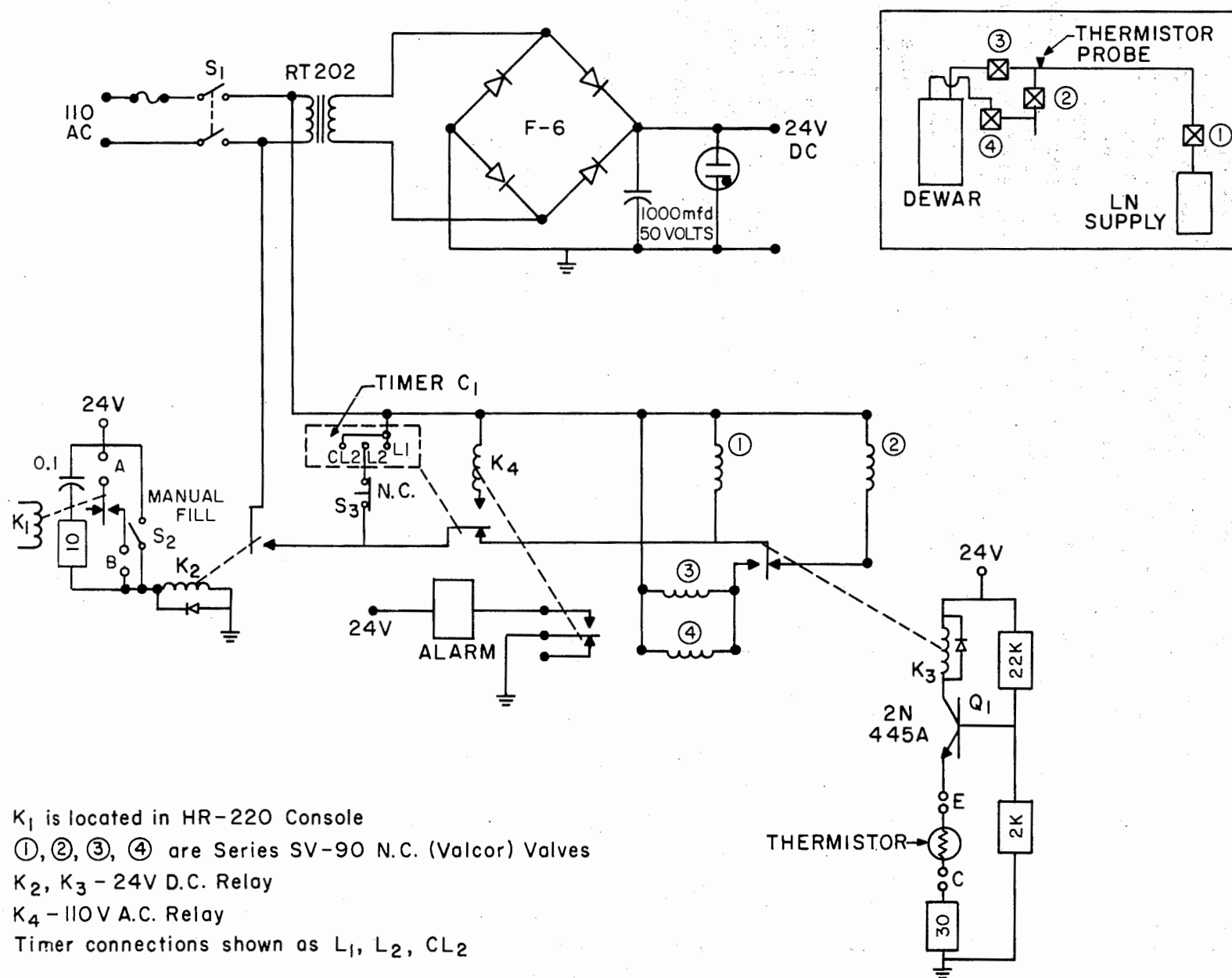
Sincerely, yours,



Arthur Kostka  
Chemistry Division

AK/skj

Note: Credit to Joseph J. Katz





LABORATORIUM  
VOOR  
ORGANISCHE CHEMIE

B-9000 GENT, August 10th 1970.

KRIJGSLAAN 105  
(België - Europa)

Tel. (09) 22.60.95

Professor Bernard L. SHAPIRO,  
Department of Chemistry,  
Texas A & M University,  
College Station,

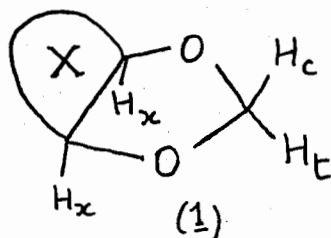
TEXAS 77843. U.S.A.

PMR features of some 1,3-dioxolanes.

Dear Barry,

We have lately been interested in some PMR features in 1,3-dioxolanes, i.e.  $^2J(O_2CH_2)$ . This parameter is found to depend [1,2] on the magnitude of the torsion angles between the  $\sigma(C-H)$  bonds involved and the adjacent lone electron pairs, and our aim is to trace this behaviour in pentacyclic compounds in order to obtain conformational information. Our dioxolane series is not yet worked through, but some readers might be interested in some striking features already obtained. The table gathers some parameters, and the following is noteworthy:

- 1) There is a small fluctuation in  $^2J(O_2CH_2)$  in 1,3-dioxolanes  
(1)  $0 < ^2J(O_2CH_2) < |1.2| \text{ cps.}$
- 2) The sign is positive, as follows from the decrease in absolute value of  $^2J(O_2CH_2)$  with increasing polarity of the medium, causing a negative contribution [3]. There seems to exist some confusion in the literature [2,4] with respect to the sign of  $^2J$  in dioxolanes, although the present result confirms the findings of Smith and Cox [3].
- 3)  $^2J(^{13}C-H_2)$  is slightly bigger for the lowfield dioxy methylene hydrogen atom than it is for the highfield counterpart. A  $\beta$ -cis substituent causes a deshielding, thus  $^2J(^{13}C-H_2)$  is larger for  $H_c$  than for  $H_t$  (1). It is clear that for a symmetric pseudorotation of the dioxolane framework the amount of eclipsing of  $H_c$  with the adjacent p lobes is equal to that of  $H_t$ . Therefore the phenomenon can't be explained by a preferential neighborhood [5] of the electron cloud to one of the H-atoms.







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(België - Europa)

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4) Only the lowfield hydrogen atom  $H_C$  is involved in long range coupling with  $H_X$  (nmr). This is a further example of a non-classical long-range phenomenon [6]

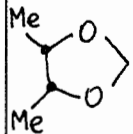
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Anteunis M. and Swaelens G.; Org. Magn. Res.: in press.  
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- [2] Cahill, R., Cookson R.C. and Crabb T.A.; Tetrah. 25 (1969), 4681.
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- [4] Rommelaere Y. and Anteunis M.; Bull. Soc. Chim. Belges, in press.  
See also Baggett N., Duxbury J.M., Foster A. and Webber J.:  
J. Chem. Soc. (c) 1966, 208  
Inch T. and Williams N.J.; J. Chem. Soc. (c) 1970, 263.
- [5] Yonezawa T. and Morishima I.; J. Mol. Spectr. 27 (1968), 210.
- [6] cf. for a discussion on the controversy on the possibility for non-classical long range Anteunis M. and Schamp N.; Bull. Soc. Chim. Belges 79 (1970) 427.
- [7] Rattet L., Williamson A. and Goldstein J.; J. Mol. Spectr. 26 (1968) 281.  
Quoted 1.0 cps by Buniacombe J.B., Foster A., Jones B. and Willard J.; Chem. Comm. (1965), 174.
- [8] From bandwidth eventually before and after decoupling experiments or from comparison with internal reference (i.e.  $CH_2Cl_2$ ).

Laboratory for NMR Spectroscopy.

M.J.O. ANTEUNIS

TABLE.

PMR parameters for bicyclic formal acetals, in  $\text{CCl}_4$  (unless otherwise stated).  $^4J(\text{H}_t, \text{H}_x) = 0$  in all cases. Shifts in ppm from TMS (internal).

x(1)	$^2J$	$^4J(\text{H}_t, \text{H}_x)$ $\Delta H_c$	$^2J((\text{O})^{13}\text{C}_2-\text{H})$ (c) $\text{H}_c$ (t) $\text{H}_t$	$\Delta H_t$	$\Delta H_x$
5	0	0.5(tr) 4.90	-	4.53	-
6	1.15[77] 1.05( $\text{CCl}_3$ ) 0.95(DMSO)	0 5.05	c 166.0 t 165.0	4.80	3.9
7	0	0.4[81] 5.10( $\text{CHCl}_3$ )	c 163.8 t 161.0	4.73 ( $\text{CHCl}_3$ )	4.05 ( $\text{CHCl}_3$ )
8	0.45 0.2[81](DMSO)	0.2[81] 4.83	c 163.0 t 161.0	4.50	3.85
9	1.07 0.98( $\text{CCl}_3$ ) 0.95(acetone- $d_6$ ) 0.80(DMSO)	0.2[81] 4.80	c 166.0 t 165.0	4.52	3.80
10	0.85 0.70( $\text{CCl}_3$ ) 0.70(acetone- $d_6$ )	0.15[81] 5.0( $\text{CCl}_3$ )	c 164.5 t 163.0	4.74 ( $\text{CCl}_3$ )	3.90 ( $\text{CCl}_3$ )
12	1.10 1.00( $\text{CCl}_3$ ) 0.95(DMF)	0.1 4.92	c 163.5 t 162.0	4.68	3.78
	0.63	<0.5 4.97	-	4.87	-

LABORATORIUM  
VOORT  
ORGANISCHE CHEMIE



GENT, August 10th 1970.  
KRIJGSLAAN 105  
(Belgie - Europa)  
Tel. (09) 22.60.95

## NORTHWESTERN UNIVERSITY

EVANSTON, ILLINOIS 60201

DEPARTMENT OF CHEMISTRY

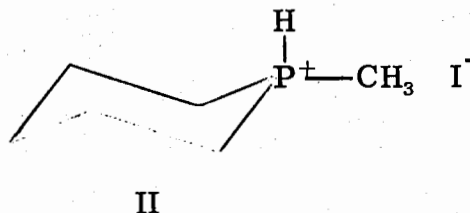
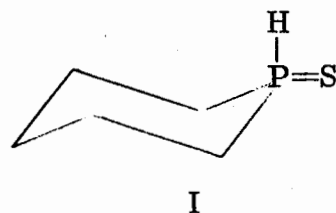
August 12, 1970

Professor Bernard L. Shapiro  
 Department of Chemistry  
 Texas A and M University  
 College Station, Texas 77843

Dear Barry:

Conformational Studies of Phosphacyclohexanes

Continuing our studies of phosphacyclohexanes, we have prepared compounds I and II.



The splitting pattern of the proton on phosphorus was examined to determine whether the proton is axial or equatorial. In the parent compound, the corresponding proton is almost entirely axial.<sup>1</sup> For the sulfide I, the spectrum of the P-H proton at 90 MHz is a broad triplet independent of temperature to  $-70^\circ$ . We estimated the coupling constants to be  $J_{aa} = 7.5$  Hz and  $J_{ae} = 2.5$  Hz. These values favor the conformation for I drawn above, with the proton axial.

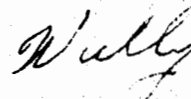
For the methiodide II, the spectrum of the P-H proton at 90 MHz with irradiation of the methyl resonances is a broad singlet. This observation is not evidence for an equatorial orientation, but arises because the axial and equatorial  $\alpha$  protons have a small chemical-shift difference. For  $J_{aa} = 8.0$  Hz,  $J_{ae} = 2.0$  Hz, and  $\Delta\nu_{ae} = 7$  Hz, the calculated spectrum is a deceptively simple singlet rather than a triplet of triplets. In the other cases,  $\Delta\nu_{ae}$  is about 45 Hz, so this problem does not arise. Thus, we cannot determine the location of the P-H proton in II because of second-order effects.

Double resonance experiments on II showed that  $^1J_{P-H}$  and  $^2J_{P-C-H}$  have opposite signs. Since  $^1J_{P-H}$  is probably positive,  $^2J_{P-C-H}$  must be negative.

Sincerely yours,



Joseph B. Lambert



Wallace L. Oliver, Jr.

- (1) J. B. Lambert, W. L. Oliver, Jr., and G. F. Jackson, III, Tetrahedron Lett., 2027 (1969).

DIVISION OF PHYSICAL CHEMISTRY

4-16 The Royal Institute of Technology

S-100 44 STOCKHOLM 70

Sweden

Cable address: Technology

Stockholm August 17, 1970

NUCLEAR MAGNETIC RESONANCE GROUP  
THE ROYAL INSTITUTE OF TECHNOLOGY  
STOCKHOLM 70 SWEDEN

Professor Bernard L. Shapiro  
Texas A & M University  
College of Science  
College Station  
Texas 77843 USA

Dear Barry:

Thank you for the reminder. I hope the equilibrium will be restored by the following report of work carried out together with Dr. Pinhas Lindner of this group. The short title of our contribution is:

Water in Tendon Collagen under Tension

Tendons from male rats, about 100 days old, were assembled in a loading device to be inserted into the spectrometer probe under controlled tension and humidity conditions. Prior to the measurements some of the samples were stored for five months at 75 % relative humidity, and for 3 months at 90 % R.H. under a constant load of 550 gr.

The tendon tension and the PMR spectra were observed at various equilibrium water contents. The results are exemplified in Figures 1 and 2 showing, respectively, the variation of the tendon tension at constant length and the corresponding width of the shifted band as functions of the regain.

The measurements establish the existence of a structural transition zone at about 20 % regain and a relation between the load at constant length and the water content of tendons expressed by the equation



$$\frac{d\sigma}{dw} = -a\sigma$$

with the solution

$$\sigma = \sigma_0 \exp [-a(w-w_0)]$$

where  $\sigma_0$  and  $w_0$  represent respectively the initial mean stress and water content of the regain interval considered.

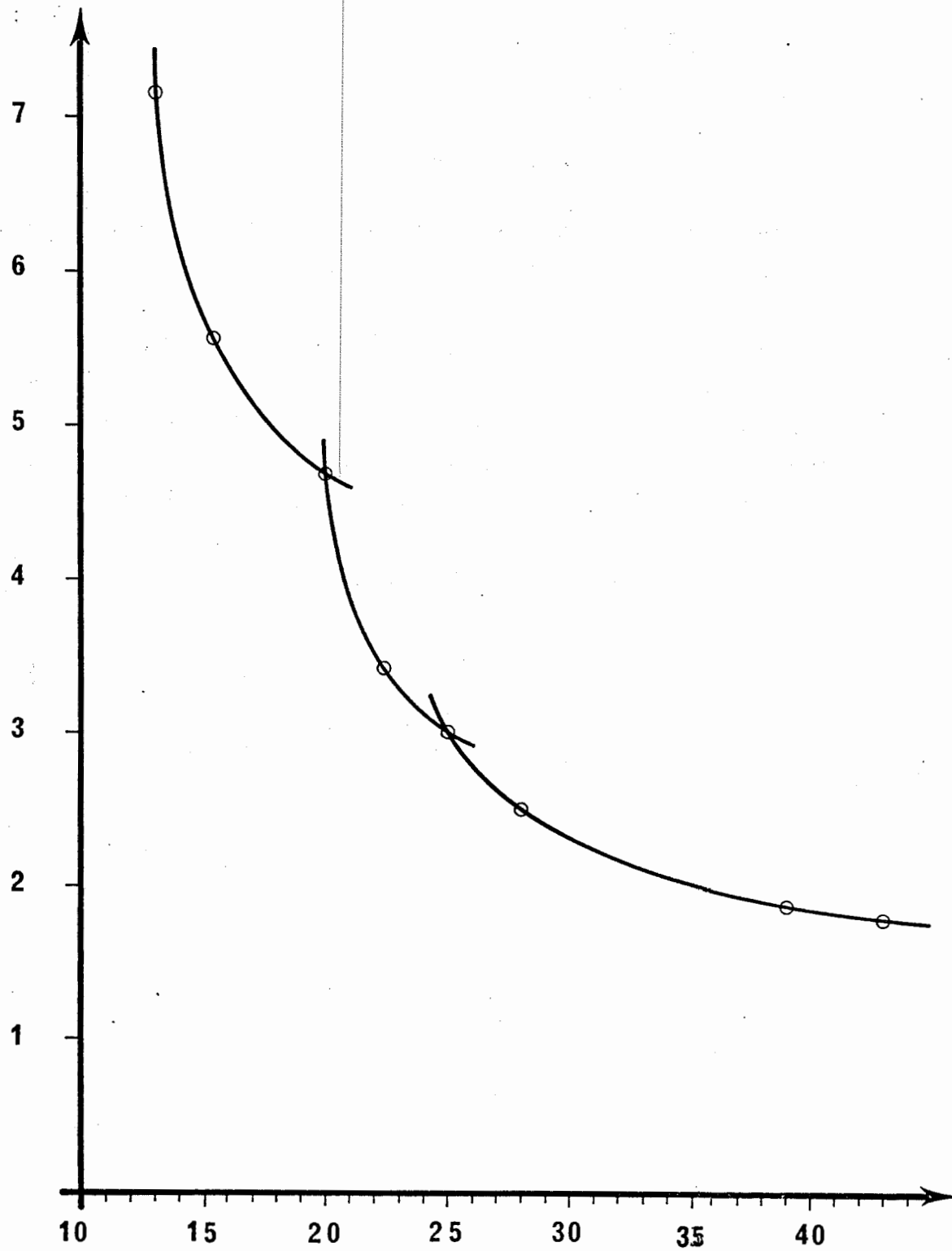
These and other observations indicate that the internal water balance of the tendons plays an important role in relaxation and load redistribution processes. We would appreciate to hear from any reader interested in the function of the mucopolysaccharides in tendons.

Yours Sincerely,

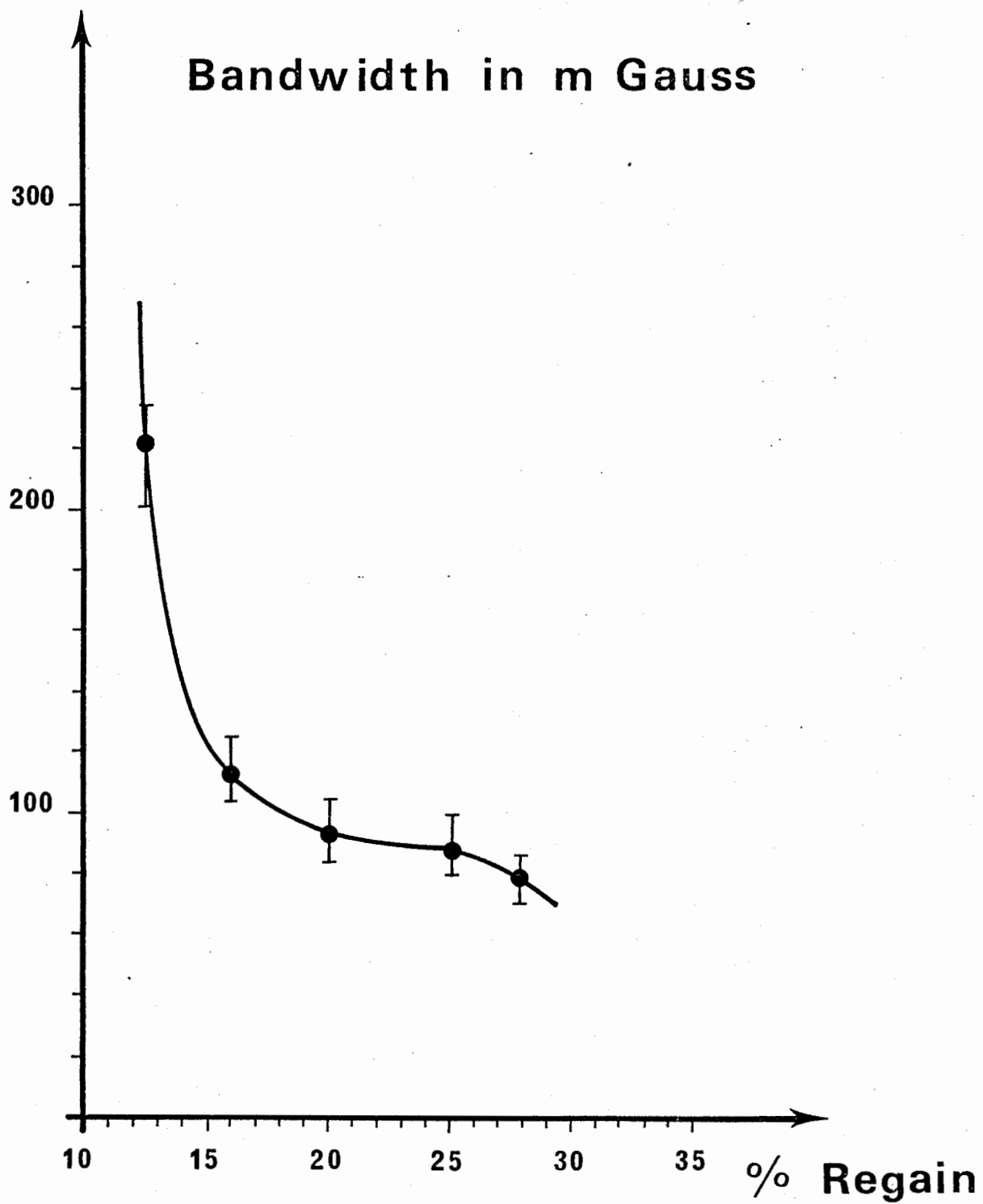


Erik Forslind  
Professor of Chem. Phys.

Load kg



% Regain



ADVANCED TECHNOLOGY  
DEPARTMENT 400  
ABBOTT LABORATORIES  
NORTH CHICAGO, ILL.

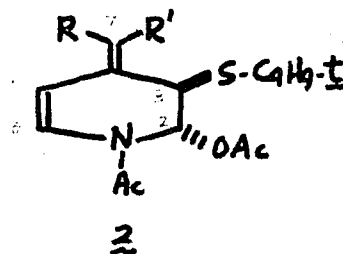
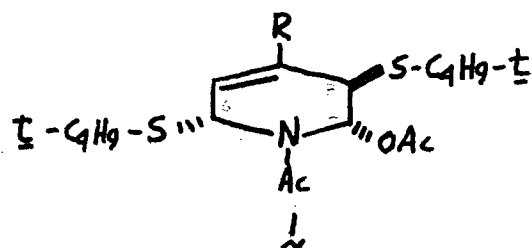
August 19, 1970

Dr. Bernard L. Shapiro  
Department of Chemistry  
Texas A&M University  
College Station, Texas 77843

Dear Professor Shapiro:

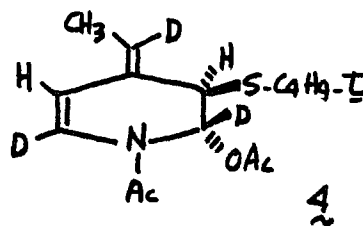
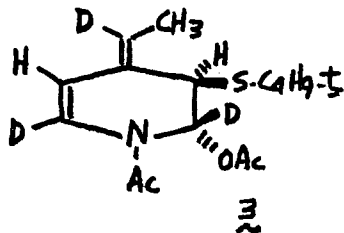
CONFIGURATION OF 4-ALKYLIDENE-1,2,3,4-TETRAHYDROPYRIDINES

The reaction of 4-alkylpyridine N-oxides with mercaptans in acetic anhydride yields mixtures of 2- and 3-alkylmercapto-4-alkylpyridines and a number of solid by-products which have been shown to be 4-alkyl-1,2,3,6-tetrahydropyridines 1 (1-3). A similar reaction in the presence of triethylamine affords different solid by-products which have recently been shown to be 4-alkylidene-1,2,3,4-tetrahydropyridines 2 (4,5).



When 4-picoline or 4-isopropylpyridine N-oxide were reacted, only a single by-product (2 R=R'=H or CH<sub>3</sub>) was isolated. However, when 4-ethylpyridine N-oxide was the starting material, a mixture (60:40) of two compounds resulted (2 R or R' = H or CH<sub>3</sub>). Various attempts to separate this mixture have failed although a sample enriched in one isomer (~90%) was obtained by fractional crystallization. The nmr spectra were consistent with the anticipated mixture of geometric isomers. Suitable but untested rationale could be developed to assign configurations to the isomers based on the chemical shifts of the ring protons (c.f. later discussion); however, a more definitive proof was sought.

The deoxidative substitution was repeated using 4-ethylpyridine N-oxide- $\alpha,\alpha,2,6\text{-d}_4$  as the starting material and a mixture of tetradeuterio isomers (3 and 4) isolated.



The results of a series of nuclear Overhauser effect experiments on the mixture are summarized in the table. Simultaneous irradiation of the vinyl methyls, necessitated by their similar chemical shifts, resulted in a significant increase in the integrated intensity of the resonances of H-5<sub>major</sub> and H-3<sub>minor</sub> and had no effect on the remaining resonances. Large intensity changes of the resonances of H-3 of both isomers resulted from irradiation of the t-C<sub>4</sub>H<sub>9</sub> groups. These results require that the major isomer is 4 and the minor isomer is 3.

These configurational assignments enable the chemical shift data, collected in the Table, to be rationalized. It appears that ring protons are markedly deshielded when the methyl group is cis. Recently, similar correlations have been made in conjugated alkadienes (6).

- (1) F.M. Hershenson and L. Bauer, J. Org. Chem., 34, 665 (1969).
- (2) F.M. Hershenson and L. Bauer, ibid., 34, 660 (1969).
- (3) R.S. Egan, F.M. Hershenson and L. Bauer, ibid., 34, 665 (1969).
- (4) B.A. Mikrut, F.M. Hershenson, K.F. King, L. Bauer, and R.S. Egan, Abstracts, 158th National Meeting, American Chemical Society, New York, Sept., 1969, No. 49.
- (5) B.A. Mikrut, Thesis, University of Illinois at the Medical Center, Sept., 1970.
- (6) C.G. Cardenas, Tetrahedron Letters, 4013 (1969).

	Chemical Shift CDCl <sub>3</sub>	n.O.e. % enhancement		
		{CH <sub>3</sub> }	{ <u>t</u> -C <sub>4</sub> H <sub>9</sub> major}	{ <u>t</u> -C <sub>4</sub> H <sub>9</sub> minor}
H-3 <sub>major</sub>	5.30	0	22.6	11.3
H-5 <sub>major</sub>	5.78	32.4	0	0
H-3 <sub>minor</sub>	5.54	20.0	13.6	14.8
H-5 <sub>minor</sub>	5.82	0	0	0

	Chemical Shifts, C <sub>5</sub> D <sub>5</sub> N solution		
	Major	Minor	$\Delta = \delta_{\text{major}} - \delta_{\text{minor}}$
H-2	6.25	6.37	-0.12
H-3	5.48	6.08	-0.60
H-5	5.85	...	...
H-6	6.76	6.62	+0.14
H-7	5.68	...	...
CH <sub>3</sub> -7	1.72	1.84	---

Sincerely,



Richard S. Egan  
nmr lab, D-408

vc



## CALIFORNIA INSTITUTE OF TECHNOLOGY

PASADENA, CALIFORNIA 91109

DIVISION OF CHEMISTRY AND CHEMICAL ENGINEERING  
GATES AND CRELLIN LABORATORIES OF CHEMISTRY

August 18, 1970

Professor Bernard L. Shapiro  
 Department of Chemistry  
 Texas A and M University  
 College Station, Texas 77843

A General Iterative Method of Calculating Exchange-Broadened NMR Spectra

Dear Barry,

In total lineshape analyses of exchange-broadened nmr spectra, usually the line-shapes are calculated from one or more unknown parameters, and the resulting calculated spectra are compared visually to the experimental curves. The unknown parameters are adjusted and the process is repeated so as to obtain a satisfactory fit. This visual fitting can be time consuming, especially if a large number of unknown parameters are involved. We have recently been obtaining computer simulations of experimental spectra utilizing a general method which allows simultaneous optimization of several unknown parameters.

Experimental spectra are digitized by storing single or multiple scans on a CAT which is interfaced to a key punch. The calculated spectrum is adjusted to fit the experimental spectrum by a general iterative least-squares method (1) that has found wide use for normal spectral analysis. (2) In the present case, however, the amplitude of each spectral point is fitted as opposed to transition frequencies. A least-squares solution is achieved when  $\phi$  defined by eq. 1 is minimized

$$\phi = \sum_{i=1}^N (A_{\underline{i}}^{\text{exp}} - A_{\underline{i}}^{\text{calc}})^2 \quad \text{I}$$

where  $A_{\underline{i}}^{\text{exp}}$  and  $A_{\underline{i}}^{\text{calc}}$  are the respective experimental and calculated amplitudes of the  $\underline{i}$ th point of the spectrum and  $N$  is the number of digitized spectral points. In general, for a given number of nuclear spins and chemical exchange sites,  $A_{\underline{i}}^{\text{calc}}$  is a function of several unknown parameters, i. e.,

$$A_{\underline{i}}^{\text{calc}} = f_{\underline{i}}(X_1, X_2, \dots, X_{\underline{j}}, \dots)$$

where the  $X$ 's refer to chemical shifts, coupling constants, rate constants, and/or relaxation times.

In each iteration, the procedure (1) involves solving eq. 2 for  $\vec{X}$ , the vector of corrections to the unknown parameters,  $X_{\underline{j}}$ .

$$D^T \vec{A} = D^T D \vec{X} \quad \text{II}$$

where

$$D_{ij} = \frac{\partial A_i^{\text{calc}}}{\partial X_j} \quad \text{III}$$

and

$$\vec{A}_i = A_i^{\text{exp}} - A_i^{\text{calc}}$$

The partial derivatives in eq. 3 are estimated numerically by

$$\frac{\partial A_i^{\text{calc}}}{\partial X_j} \approx \frac{f_i(X_1, X_2, \dots, X_j + h, X_{j+1}, \dots) - f_i(X_1, X_2, \dots, X_j, \dots)}{h}$$

where the function  $f_i$  is evaluated using the DNMR program (3), and  $h$  is a small increment in the  $j$ th unknown.

Figure 1 shows experimental and calculated proton spectra of the *t*-butyl resonance in 3-chloro-2,2,3,4,4-pentamethylpentane. In the one nucleus, two-site calculation (two of the three sites are accidentally degenerate), the unknown parameters consist of one rate constant and two chemical shifts. Above the coalescence temperature, the two chemical shifts were allowed to vary in the minimization sequence, but with a constant difference. Without this restraint, convergence was not achieved.

In Figure 2, the experimental and computer-simulated proton-decoupled natural-abundance  $^{13}\text{C}$  spectra of 1,1,3,3-tetramethylcyclohexane are shown. The invariant peak is due to the quaternary carbon, the presence of which allows convergence while varying  $T_2^*$ . The exchange-broadened peaks are due to the methyl carbons and the other resonances are not shown. The other unknown parameters in the one nucleus, three-site calculations were the rate constant for conformational inversion, the three chemical shifts, and the population of the quaternary site. The latter parameter permits taking into account differences in Overhauser enhancement of methyl vs. quaternary carbons. Again, the chemical-shift differences between the two exchanging sites was held constant for spectra above the coalescence temperature.

Initial values of the unknown parameters were estimated from the experimental spectra and convergence was rapid. Five or less iterations were necessary for each spectrum in Figures 1 and 2.

The principal advantage of this general method is the small time expenditure required for accurate, total lineshape analyses.

Yours sincerely,

*Bruce L. Hawkins*

*Jack*

Bruce L. Hawkins

John D. Roberts

- (1) W. C. Hamilton, "Statistics in Physical Science", The Ronald Press Co., New York, 1964, p. 150.
- (2) S. M. Castellano and A. A. Bothner-By, "Computer Programs for Chemistry", D. T. Detar, Ed., Vol. I, W. A. Benjamin, Inc., 1968, pp. 10-39.
- (3) G. Binsch, J. Amer. Chem. Soc., 91, 1304 (1969).

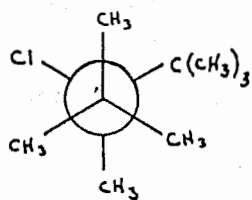
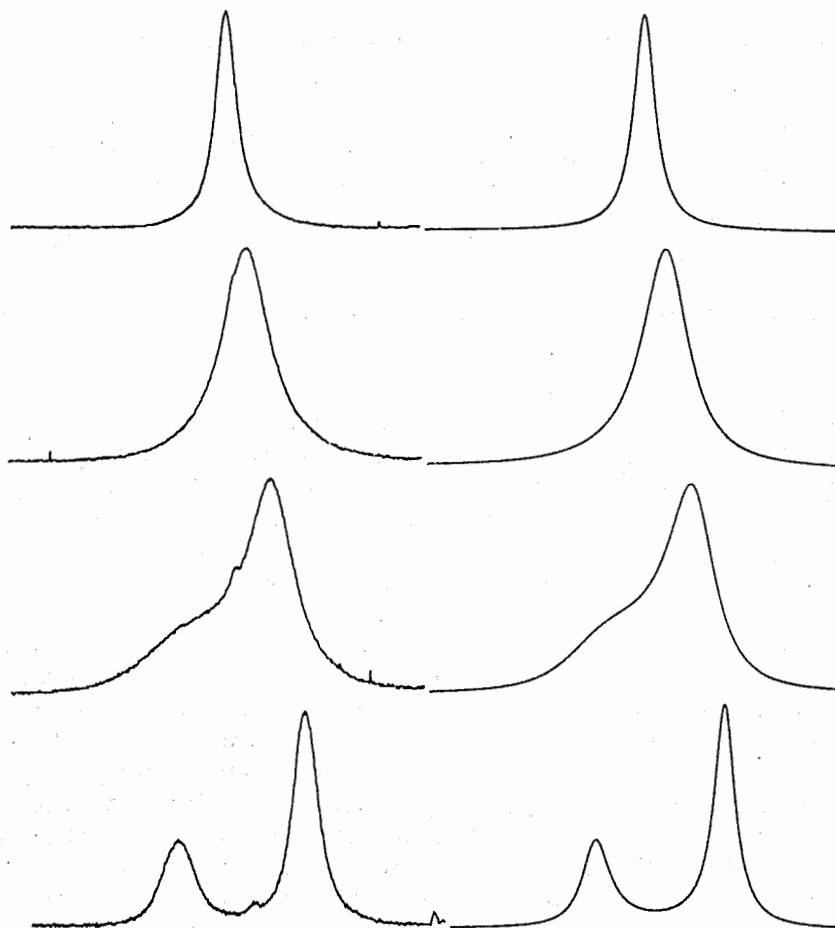


Figure 1.

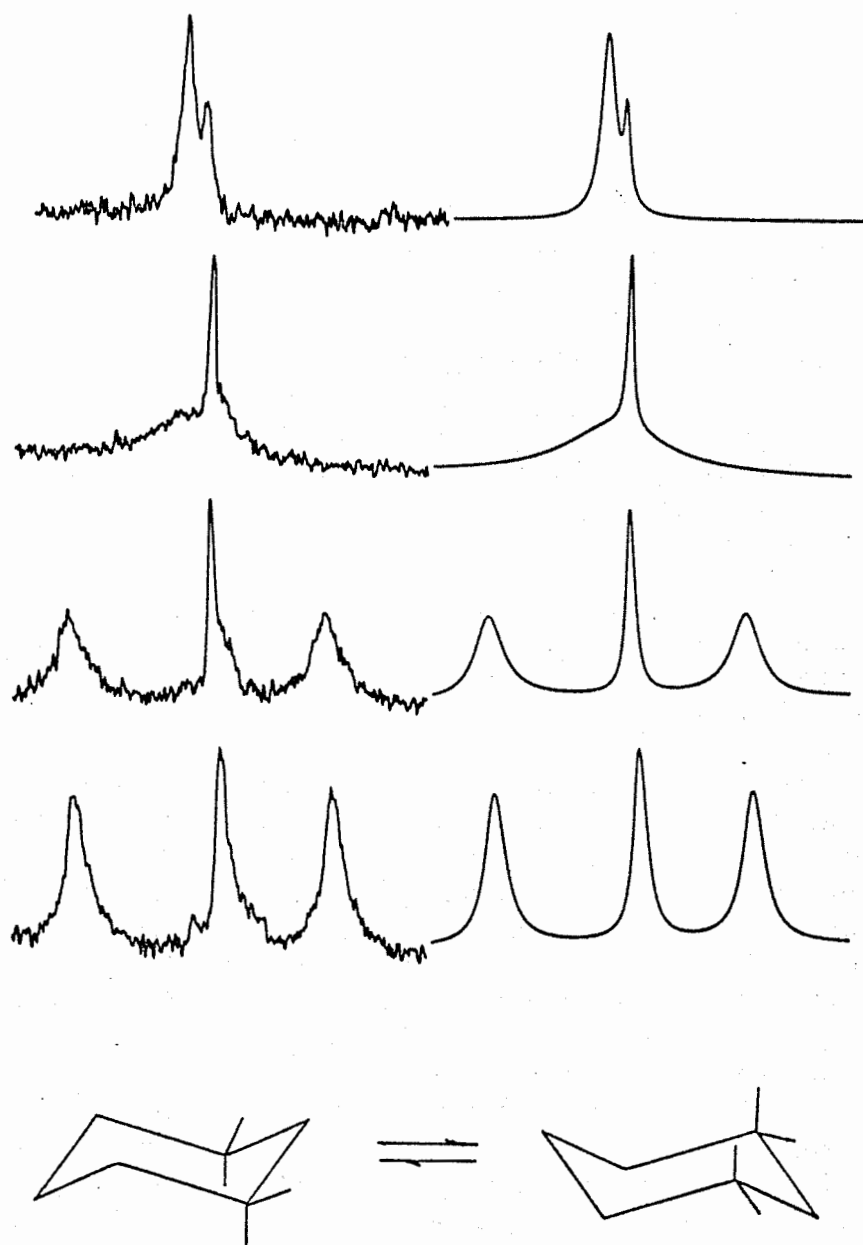


Figure 2.

STANFORD UNIVERSITY  
STANFORD, CALIFORNIA 94305

DEPARTMENT OF CHEMISTRY

20 August 1970

Professor Bernard L. Shapiro  
Department of Chemistry  
Texas A & M University  
College Station, Texas 77843

On Use of HA-100 Console with HR-60 System  
Modified Pen Holder for HA-100, T-60, some A-60's

Dear Barry:

While most of our colleagues are concerned with the conversion of their nmr systems to higher fields and frequencies, we have recently had occasion to reverse this process.

After obtaining some relaxation measurements at 100 MHz, one of our students wanted to repeat some of the measurements at 60 MHz, only to find that our HR-60, which has no field-frequency lock, was not stable enough. Since both instruments are located in the same room, we decided to try using the HA-100 console to control the 60 MHz RF unit, probe and field.

We were pleased to find that the extension cables (approx. 20 ft) required to make the interconnection did not diminish the signals below a useful level. Thus it was not necessary to move any heavy equipment. The conversion now requires less than five minutes and involves the switching of a few connections from standard arrangement to an interconnecting cable which we have tied together in a permanent harness.

For this operation, the regular cables for the homogeneity control and slow sweep units of the HR-60



Professor Bernard L. Shapiro  
20 August 1970

-2-

have been relocated so that the controls may be set near the HA-100 console. We did not attempt to extend the homogeneity cables. Attempts to extend the slow sweep cables led to a poor flux stabilizer lock. Possibly this could have been compensated for by readjusting the amplifier gain and magnet regulation controls on the V-3506. Parallel connection of the two scopes (optional) permits monitoring the signal while adjusting RF unit or probe at the HR-60 console. All other adjustments are made from the HA-100 console area.

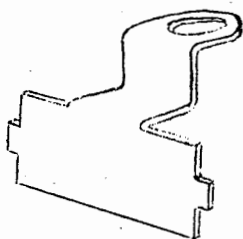
We have had a modified pen holder made up for use with the LeRoy lettering pens used on the HA-100, T-60 and some A-60's which does not require removal of the pen from the holder to cap the pen. We believe it is more conducive to cap the pen if no adjustment of holder is required. The holder is of 1/16 inch sheet metal (see diagram).

Sincerely yours,

*M. Bramwell*  
M. Bramwell

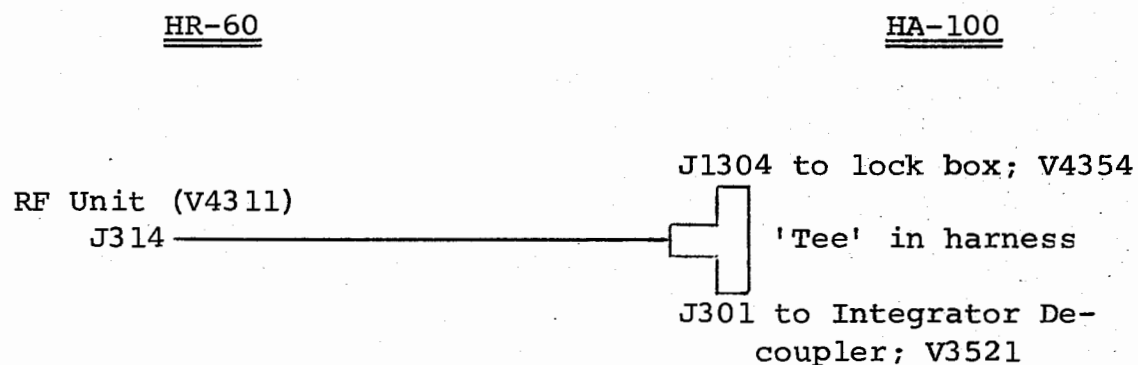
*Lois J. Durham*  
L. Durham

*S. Seaver*  
S. Seaver

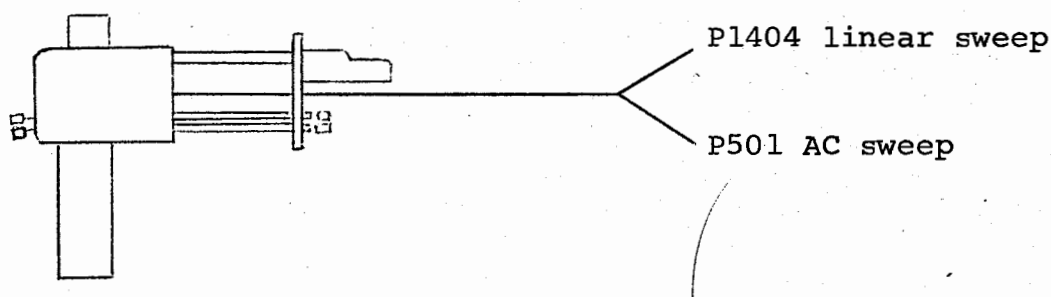


Please credit subscription  
to L.J.D.

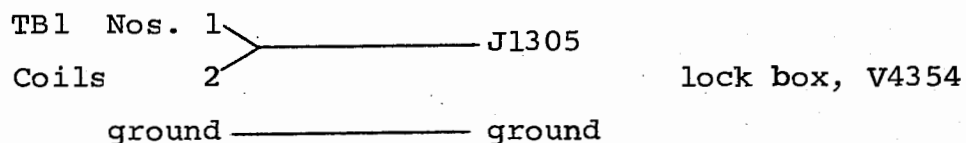
Schematic for production of an HA-60 hybrid by  
crossing an HR-60 with an HA-100



Probe



Flux Stabilizer (V3506)



Scope

vert —————

horiz. —————

Scope

vert

horiz.

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**BROCK**  
**University**

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St. Catharines, Ontario

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144-29  
Glenridge Campus  
684-7201 Ext. 317

August 20, 1970

Dr. B. L. Shapiro,  
Department of Chemistry,  
Texas A & M University,  
College Station,  
Texas 77843  
U.S.A.

Dear Dr. Shapiro:

Effects of hydrogen fluoride contamination  
on nmr spectra of  $\text{BF}_3$  adducts

A number of  $^{19}\text{F}$  studies of  $\text{BF}_3$  adducts have shown the presence of small amounts of a second fluorine-containing species in addition to the simple donor-acceptor adduct. If the Lewis base involved is sufficiently weak, the second  $^{19}\text{F}$  peak can arise from the  $\text{BF}_3$  adduct of water present in incompletely dried materials (1); if the adduct is unstable, various decomposition products may appear. However, even in stable systems in which the Lewis base is too strong to be displaced from its adduct by water, a second peak, usually small, may appear. Our recent work suggests that in at least some such cases the extra peak arises from contamination of the system by HF, rather than from adducts of other than 1:1 stoichiometry or from an associated form of the 1:1 adduct, as have on occasion been postulated. Some results from the  $\text{BF}_3$ -tetramethylurea (TMU) system illustrate this. We have observed similar effects in trialkylamine and N,N-dialkylamide systems.

$\text{BF}_3$  is known to form a 1:1 adduct with TMU; thus, when TMU and  $\text{BF}_3$  in a 2:1 molar ratio are introduced into methylene chloride as solvent, all of the  $\text{BF}_3$  should be present as the 1:1 adduct. Such a solution, when prepared directly from commercial  $\text{BF}_3$ , gave the two-peak room temperature  $^{19}\text{F}$  spectrum shown in Fig. 1a. It was possible to resolve the smaller high-field peak into a 1:1:1:1 quartet with  $J_{\text{BF}_3-\text{TMU}} = 1.1$  Hz, reminiscent of  $\text{BF}_4^-$  (2). The presence of  $\text{BF}_4^-$  was confirmed by a pronounced increase in the size of this peak, with no change in the fine structure, when  $n\text{-Bu}_4\text{N}^+\text{BF}_4^-$  was added to the solution (Fig. 1b). Careful purification of  $\text{BF}_3$ , in particular a trap-to-trap distillation from a  $-112^\circ$  trap, gave a sample

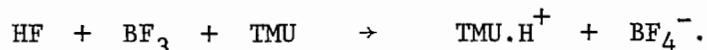
(continued.../2)

Dr. B. L. Shapiro

- 2 -

August 20, 1970.

having a  $^{19}\text{F}$  spectrum having only one major peak; we could not, however, completely eliminate the second peak (Fig. 1c). Additions of anhydrous HF to such a solution caused the  $\text{BF}_4^-$  peak to grow at the expense of the  $\text{TMU.BF}_3$  peak. HF,  $\text{BF}_3$ , and TMU apparently react as follows, until the HF is used up:



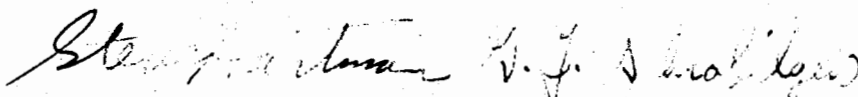
The remaining  $\text{BF}_3$  forms  $\text{TMU.BF}_3$ . The relative  $^{19}\text{F}$  peak areas would thus indicate the proportion of HF in  $\text{BF}_3$ .

It should be noted that some samples of commercial  $\text{BF}_3$ , in particular those taken from nearly-empty cylinders in which the less volatile HF had become concentrated, contain up to 20% HF; normal purification procedures may not be designed to handle such large amounts of impurities. Thus it seems likely that some of the studies reported in the literature have been carried out using impure material.

All samples in which  $[\text{TMU}] > [\text{BF}_3]$  give two major room temperature  $^1\text{H}$  peaks corresponding to "free" TMU and  $\text{TMU.BF}_3$ . The samples which show a significant  $\text{BF}_4^-$  peak in the  $^{19}\text{F}$  spectrum also show an abnormal low-field shift for the "free TMU"  $^1\text{H}$  peak. According to the above reaction, such samples should contain three TMU species, free TMU,  $\text{TMU.H}^+$ , and  $\text{TMU.BF}_3$ . Since proton exchange is generally very fast, we expect a single averaged signal for TMU and  $\text{TMU.H}^+$  which should shift to low field as the relative amount of  $\text{TMU.H}^+$  and  $\text{BF}_4^-$  increases, as is observed.  $\text{BF}_3$  exchanges much more slowly than  $\text{H}^+$  among TMU molecules, so that a separate  $\text{TMU.BF}_3$  peak, of fixed chemical shift, is visible up to about  $90^\circ\text{C}$ . A more complex case of proton transfer affecting chemical shifts of "free" base molecules has been reported in the methanol- $\text{BF}_3$  system (3).

The elimination of complications arising from HF contamination has made possible more exact studies of the surprisingly complex  $\text{TMU-BF}_3$  system.

Yours sincerely,

  
J. S. Hartman      G. J. Schrobilgen

JSH:GJS:hk

#### References:

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- (2) R.J. Gillespie, J.S. Hartman and M. Parekh, Can. J. Chem., 46, 1601 (1968).
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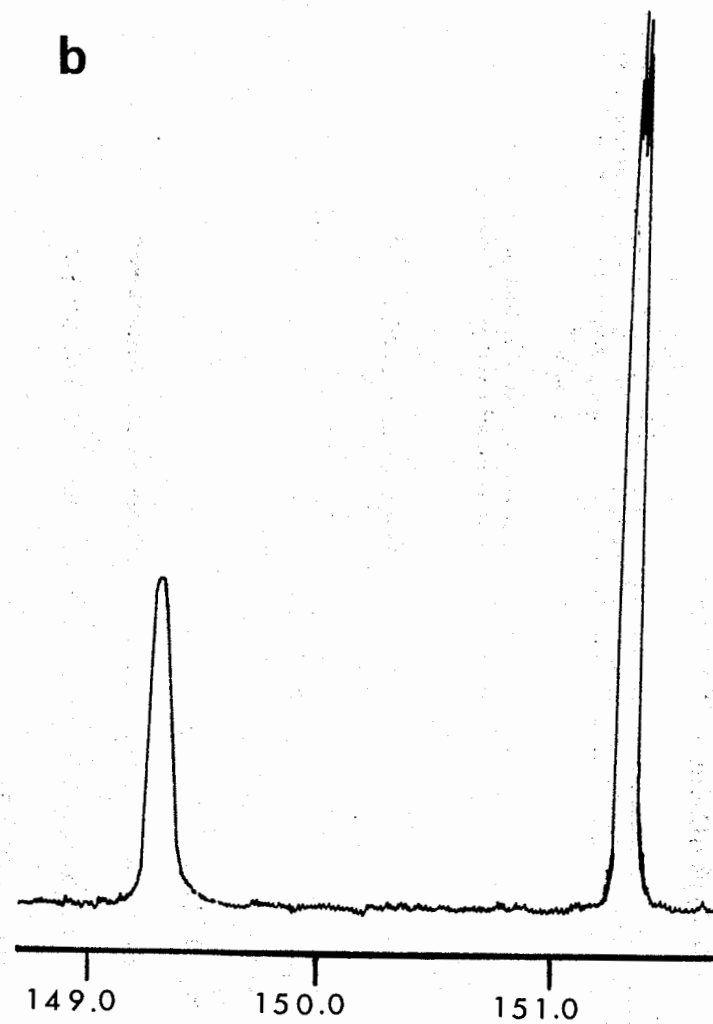
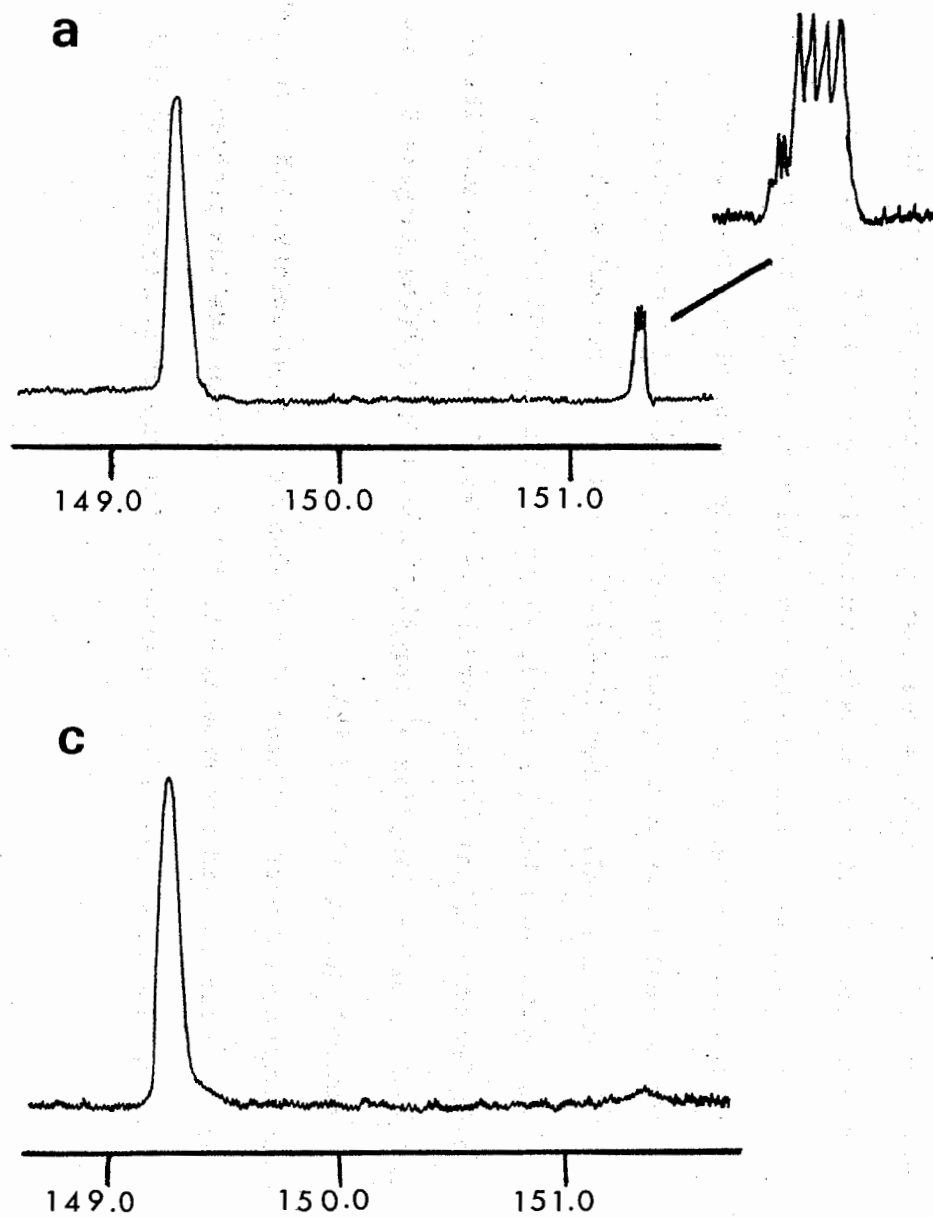


Fig. 1 -  $^{19}\text{F}$  spectra of TMU- $\text{BF}_3$  solutions in methylene chloride. TMU is present in excess. Calibrations are in ppm from  $\text{CFCl}_3$ .



DEPARTMENT OF CHEMISTRY  
LOS ANGELES, CALIFORNIA 90024

August 21, 1970

Professor B. L. Shapiro  
Texas A. M. University  
Department of Chemistry  
Collect Station, Texas 77843

Title: The Direct Determination of the Axial Conformer  
of Methylcyclohexane by 63.1 MHz <sup>13</sup>C NMR at Low  
Temperatures.

Dear Barry:

A great deal of work has been done on the conformational equilibria of substituted cyclohexanes. Perhaps the most unambiguous procedure for the determination of the relative amounts of equatorial and axial forms is to measure the nmr spectrum at such low temperatures (e.g. -100°) that separate spectra are given by the two forms.<sup>1,2</sup> Integration then gives the desired result, provided that peaks do not overlap for all the nuclei under observation. With proton nmr, this method is easy to apply to compounds such as chlorocyclohexane or cyclohexyl acetate, where (i) the methine proton on C-1 has quite different chemical shifts for the two forms, and (ii) the equilibrium constant ( $K = \frac{\text{equatorial form}}{\text{axial form}}$ ) is not too large (the largest K which has been measured<sup>1</sup> has a value of 29).

Methylcyclohexane meets neither of the above two requirements, and furthermore partial deuteration is not expected to solve the peak overlap problem. Thus there has been no report of the observation of the proton spectrum of the

August 21, 1970

axial form of methylcyclohexane. We now have been able to observe the 63 MHz  $^{13}\text{C}$  spectrum of the axial conformer of methylcyclohexane at  $-100^\circ$ . (Fig. 1). The work of Dalling and Grant, shows that the chemical shifts of the methyl carbon ( $\alpha$ ) and the methylene carbons 3 and 5 should be at about 8 ppm higher field than the corresponding carbons in the equatorial form. Therefore, at 63 MHz the  $\alpha$ , 3 and 5 carbons should occur in a region free of bands of the equatorial form, or of its  $^{13}\text{C}$  satellites ( $J_{\text{C}^{13}\text{C}^{13}} \approx 35$  Hz for  $\text{sp}^3\text{-sp}^3$  single bond). The bands in Figure 1 assigned to the axial isomer are not present at  $-75^\circ$ , and are thus unlikely to be due to chemical impurities. Furthermore their chemical shifts are in close agreement with the empirical chemical shift rules of Dalling and Grant, and their relative intensities ( $\approx 2:1$ ) are as expected.

The value of the equilibrium constant is about 100, corresponding to an A value for the methyl group of 1.6 kcal/mole, in good agreement with the accepted value of 1.7 kcal/mole.

Yours sincerely,

*Frank Craig*

F. A. L. Anet  
C. H. Bradley  
G. W. Buchanan\*

- 1) F. R. Jensen, C. H. Bushweller, and B. H. Beck, J. Amer. Chem. Soc., 91, 344 (1969).
- 2) F. A. L. Anet and P. M. Henrichs, Tetrahedron Letters, 741 (1969).
- 3) The spectrometer, which makes use of a 59 kgauss superconducting solenoid, was briefly described at the 11th Experimental NMR Conference, Pittsburgh, Pa. April 1970.
- 4) D. K. Dalling and D. Grant, J. Amer. Chem. Soc., 89, 6612 (1967).

\*Present address: Department of Chemistry  
University of Windsor  
Windsor, Ontario  
Canada



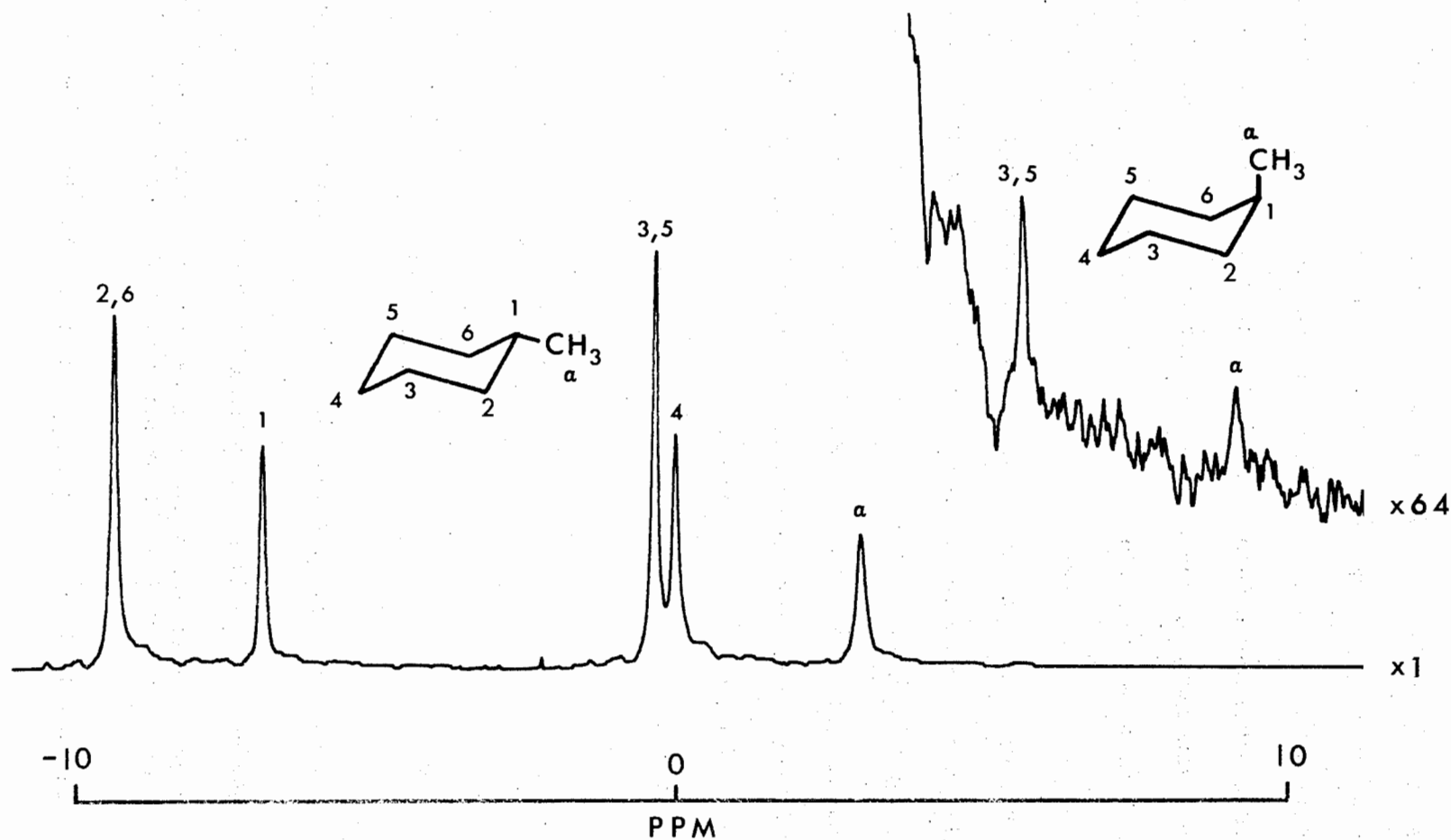


Figure 1. 63.1 MHz  $^{13}\text{C}$  spectrum (protons noise-decoupled at 251 MHz) of neat methylcyclohexane at  $-110^\circ$ . Carbon 4 which is expected to have the same chemical shift in both conformers, is used as an internal reference. Zero on this scale is about 102 ppm upfield from benzene and about 27 ppm downfield from TMS. Spinning 10 mm sample tubes were used and the sweep rate was 16 Hz/sec. The spectrum represents 96 scans time-averaged on a Varian CAT.



# Imperial Chemical Industries Limited

DYESTUFFS DIVISION

P.O. Box 42, Hexagon House, Blackley, Manchester, 9

Professor E.L. Shapiro,  
Department of Chemistry,  
Texas A and M University,  
College Station,  
Texas 77843,  
U.S.A.

Your Ref:

Our Ref: PL/ARS/B5

21 August, 1970.

Dear Professor Shapiro,

Thank you for your blue letter; alas, the moving pen having writ thrice last year must still write on.

## $^{14}\text{N}$ shifts in Substituted Anilines

We have continued our study of  $^{14}\text{N}$  chemical shifts obtained by the double resonance method reported previously <sup>1,2</sup>, our attention this time being directed to meta and para substituted anilines. Measurements were made on thirteen compounds in acetone solution. The shifts are quoted in p.p.m. upfield from the nitrate nitrogen signal a 4.5M solution of ammonium nitrate in aqueous 5N-hydrochloric acid and are shown in the following table.

Compound	$^{14}\text{N}$ shift	Compound	$^{14}\text{N}$ shift
Aniline	321.7 $\pm$ 0.5		
p-NO <sub>2</sub> "	306.5 $\pm$ 0.5	m-NO <sub>2</sub> Aniline	317.7 $\pm$ 0.5
p-CH <sub>3</sub> CO "	312.7 $\pm$ 0.5	m-CH <sub>3</sub> CO "	319.4 $\pm$ 0.5
p-CH <sub>3</sub> OCO "	312.6 $\pm$ 0.5	m-CH <sub>3</sub> OCO "	319.9 $\pm$ 2.0
p-Br "	320.1 $\pm$ 1.0	m-CH <sub>3</sub> "	319.8 $\pm$ 0.5
p-Cl "	320.5 $\pm$ 1.0		
p-CH <sub>3</sub> "	323.7 $\pm$ 0.5	m-CH <sub>3</sub> "	322.2 $\pm$ 0.5
p-NH <sub>2</sub> "	330.1 $\pm$ 1.0		

The shifts of both the meta and para substituted compounds correlate with the Hammett  $\sigma$  parameters of the substituents. These results will be discussed more fully in a paper now in preparation.

We are hoping to take delivery within the next two or three months of a Ferranti Argus 500 computer and a Fourier transform accessory for our RA-100 spectrometer and we anticipate having some results to report in our next contribution.

Yours sincerely,

F. HAMPSON, A. MATHIAS.

1. F. Hampson and A. Mathias *Molec. Phys.* **11**, 541, 1966.
2. F. Hampson and A. Mathias *Nucl. Newsletter* **25**, 1, 1966.

NATIONAL RESEARCH COUNCIL OF CANADA  
CONSEIL NATIONAL DE RECHERCHES DU CANADA

OTTAWA 7.

31 August 1970

Dr. B.L. Shapiro  
Department of Chemistry  
Texas A. and M. University  
College Station, Texas 77843

Dear Barry, "Solvent Effects on Fluoroform Shifts"

I was very interested in the results given by W.B. Smith in Newsletters #143 p22 August 1970 on Solvent Effects of fluoroform shifts and coupling for both H and F nuclei. The statement is made that "whereas the proton chemical shifts (except in benzene) are downfield from the value in cyclohexane the fluorine chemical shifts move upfield with increasing solvent polarity".

In our work on perturber effects on the F and H resonance in gaseous  $\text{HCF}_3$  [J. Chem. Phys. 38 1562 (1963)] we obtained the same result for F in  $\text{CHF}_3$  and attributed it to the fact that the coefficient of the electric field term is negative in the equation for the intermolecular effect on chemical shifts viz eqn (5) in the above reference. That is, in the equation

$$\sigma_E = -A\langle E_z \rangle - B\langle E^2 \rangle$$

A is positive for H in CH bonds and relatively small. B is also positive so increasing solvent polarity gives larger downfield shifts. For F in  $\text{HCF}_3$  however, A is negative and relatively large so that although B is positive the effect of increasing polarity is to shift the F resonance to higher field because of the dominating effect of the term in A.

Yours sincerely,

H.J. Bernstein

HJB:sc

Gerhard Binsch  
Department of Chemistry  
University of Notre Dame  
Notre Dame, Indiana 46556

Joseph B. Lambert  
Department of Chemistry  
Northwestern University  
Evanston, Illinois 60201

September 1, 1970

Professor Bernard L. Shapiro  
Department of Chemistry  
Texas A and M University  
College Station, Texas 77843

Symposium on Dynamic NMR Spectroscopy

Dear Barry:

We hereby wish to announce a two-day symposium on "Dynamic NMR Spectroscopy", to be held on March 30 and 31, 1971, jointly with the Los Angeles ACS meeting. This symposium will be sponsored by the Organic Division and possibly co-sponsored by the Division of Physical Chemistry. A decision regarding the co-sponsorship will be made in September, 1970.

The backbone of the symposium will consist of a series of invited papers. The following scientists have so far assented to contribute:

F. A. L. Anet: "Dynamic NMR Studies at 59 Kilogauss."  
C. H. Bushweller: "Rate Processes in Acyclic Trialkylamines."  
R. Freeman (and H. D. W. Hill): "Nuclear Spin Relaxation Studied by Fourier Transform Methods."  
P. Jesson (and P. Meakin): "Stereochemically Nonrigid Six-Coordinate Hydrides."  
J. Jonas: "High-Pressure NMR Relaxation Studies of Liquids."  
M. Saunders: "Dynamic NMR Studies of Carbonium-Ion Rearrangements."

In addition, there will be room for about a dozen 15-minute presentations. We solicit contributions from the following areas of current research: Theoretical and experimental aspects of exchange-broadened nmr line shapes, carbonium-ion rearrangements by dynamic nmr, theory and applications of nuclear-spin relaxation phenomena, dynamic aspects of multiple resonance, pseudorotation and

Professor Bernard L. Shapiro

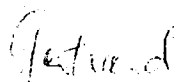
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other permutational phenomena, pulse techniques in dynamic nmr, the study of conformational interconversions, atomic inversions and valence isomerization by dynamic nmr, nuclear-spin exchange and relaxation processes in inorganic and organometallic compounds, and applications in biochemistry.

Those of your readers who are interested in contributing a short paper are asked to communicate with either of us. The Standard Abstract Form for ACS Meeting Papers is obtainable from us or from the ACS Divisional Offices. A 200 word abstract on the official form is all that is needed in advance of the meeting, but this abstract must be in our hands before November 25, 1970..

Sincerely yours,



Gerhard Binsch



Joseph B. Lambert

kjs

## CLEMSON UNIVERSITY

CLEMSON, SOUTH CAROLINA 29631

DEPARTMENT OF  
CHEMISTRY AND GEOLOGY

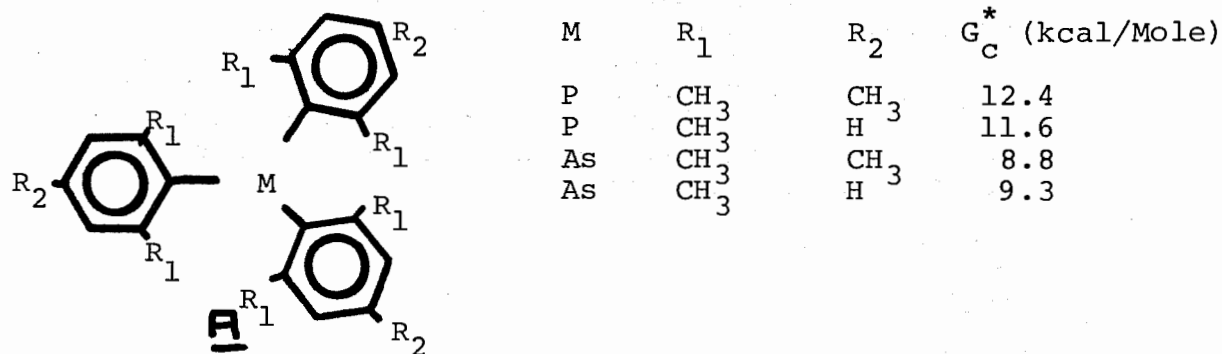
September 8, 1970

Dr. Bernard L. Shapiro  
 Department of Chemistry  
 Texas A & M University  
 College Station, Texas 77843

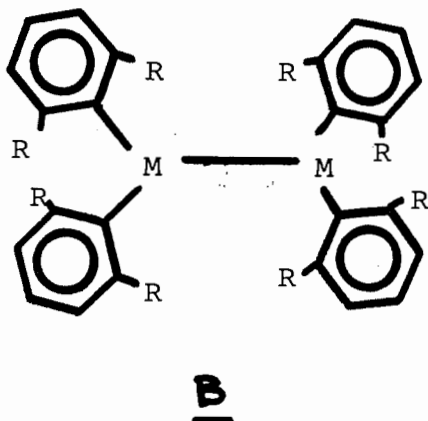
## ROTATIONAL BARRIERS IN DIPHOSPHINES

Dear Barry:

Substantial rotational barriers about C-M bonds in highly substituted derivatives of structure A have been observed (1,2). A substantial barrier to rotation



about the central M-M bond in compounds of type B must be invoked to rationalize the experimental results reported for tetrakis-(pentafluorophenyl)-diarsine (3).




In this connection, we have studied the temperature dependent spectra of tetrakis-(2,6-dimethylphenyl)-diphosphine ( $B; R=CH_3, M=P$ ). At room temperature in toluene  $d_8$  only a single methyl resonance is observed at 145.5 Hz below internal TMS (60 MHz). This observation is most consistent with a model in which all molecular torsions are rapid, or, a model in which only a single torsional M-M diastereomer is present (trans) and M-C rotation is rapid. A frozen gauche diastereomer should exhibit two methyl resonances even if C-M rotation is rapid.

At  $-48^\circ$ , four methyl resonances are observed at ca. 180, 157, 130, and 127 Hz, with areas 1:1:2 (sum of areas for resonances at 130 and 127 Hz), respectively. This observation is most succinctly rationalized by a model in which P-P rotation is rapid and P-C rotation is slow. The cant of the rings can be the same on both P atoms ( $C_2$  symmetry in the eclipsed form) or opposite on the  $^2P$  atoms ( $C_s$  symmetry in the eclipsed form). Alternatively, slow P-C rotation in any single P-P torsional diastereomer (preferably trans) will provide the same result; slow P-P rotation alone does not suffice to rationalize the experimental observation.

We are currently synthesizing systems which will hopefully afford information on phosphorous pyramidal inversion, P-C bond rotation, and P-P bond rotation simultaneously.

Sincerely yours,

  
John Jacobus

- (1) A. Rieker and H. Kessler, Tet. Lett., 1227(1969).
- (2) Unpublished work, J. J.
- (3) M. Green and D. Kirkpatrick, Chem. Commun., 57(1967).

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